

## The use of faecal microbiota transplant as treatment for recurrent or refractory *Clostridium difficile* infection and other potential indications:

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**The use of faecal microbiota transplant as treatment for recurrent or refractory *Clostridium difficile* infection and other potential indications: joint British Society of Gastroenterology (BSG) and Healthcare Infection Society (HIS) guidelines.**

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Complete List of Authors:	<p>Mullish, Benjamin; Imperial College London, Division of Integrative Systems Medicine and Digestive Disease, Department of Surgery and Cancer; Imperial College Healthcare NHS Trust, Department of Gastroenterology and Hepatology, St Mary's Hospital</p> <p>Quraishi, Mohammed; Queen Elizabeth Hospital Birmingham, Department of Gastroenterology</p> <p>Segal, Jonathan; Imperial College London, Division of Digestive Diseases/ Liver Unit; Imperial College Faculty of Medicine - Northwick Park and Saint Marks Campus, Inflammatory Bowel Disease Unit</p> <p>McCune, Victoria; Public Health Laboratory Birmingham, Public Health England; University of Birmingham, Institute of Microbiology and Infection</p> <p>Baxter, Melissa; Royal Devon and Exeter NHS Foundation Trust, Department of Microbiology</p> <p>Marsden, Gemma; Healthcare Infection Society</p> <p>Moore, David; University of Birmingham, School of Health and Population Sciences</p> <p>Colville, Alaric ; Royal Devon and Exeter NHS Foundation Trust, Department of Microbiology</p> <p>Bhala, Neeraj; University Hospital Birmingham NHS Foundation Trust, Gastroenterology; University of Birmingham, Institute of Translational Medicine</p> <p>Iqbal, Tariq; University Hospital Birmingham NHS Trust, Gastroenterology; University of Birmingham, Institute of Translational Medicine</p> <p>Settle, Christopher; City Hospitals Sunderland NHS Foundation Trust, Department of Microbiology</p> <p>Kontkowski, Graziella; C diff Support</p> <p>Hart, Ailsa; Imperial College Faculty of Medicine - Northwick Park and Saint Marks Campus, Inflammatory Bowel Disease Unit; Imperial College London, Division of Integrative Systems Medicine and Digestive Disease, Department of Surgery and Cancer</p> <p>Hawkey, Peter; University of Birmingham, Institute of Microbiology and Infection</p> <p>Goldenberg, Simon; King's College London, Centre for Clinical Infection and Diagnostics Research; Guy's and St Thomas' NHS Foundation Trust,</p>

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	Department of Microbiology Williams, Horace; Imperial College London, Division of Integrative Systems Medicine and Digestive Disease, Department of Surgery and Cancer; Imperial College Healthcare NHS Trust, Department of Gastroenterology and Hepatology, St Mary's Hospital
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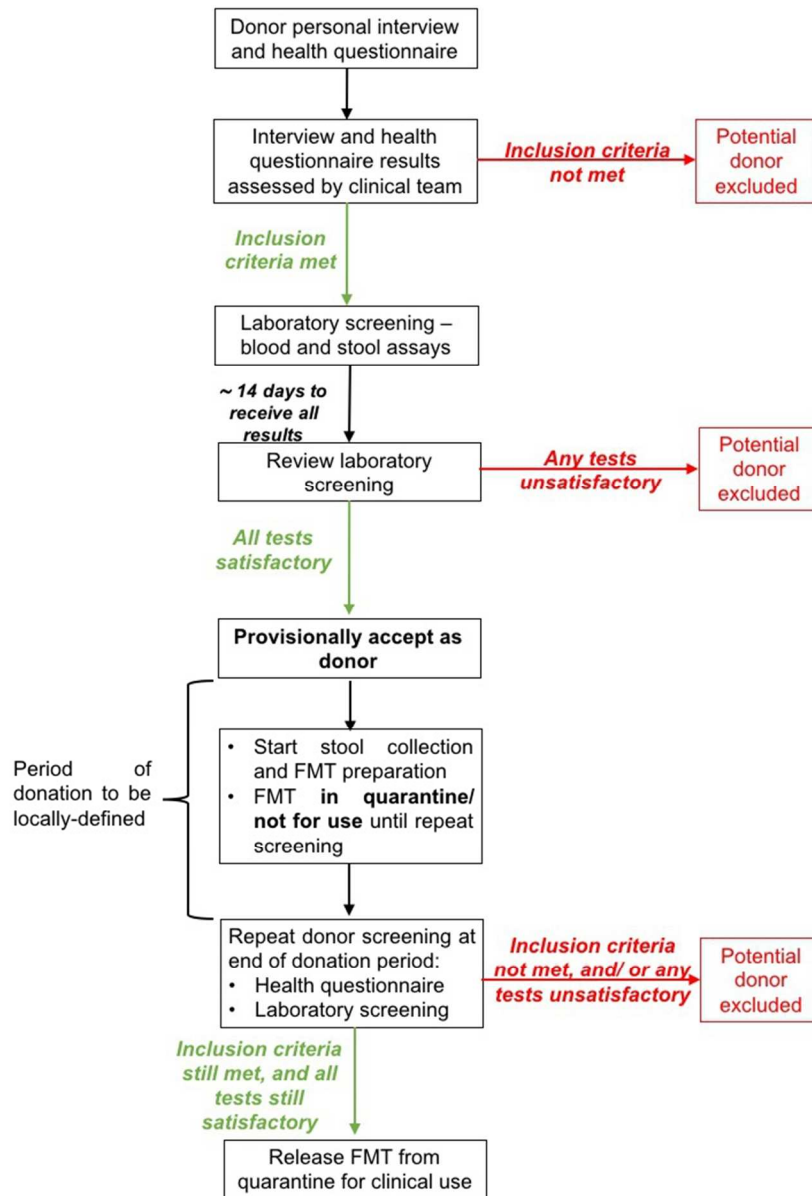


Figure 1: Proposed summary pathway for donor screening for centres preparing frozen FMT from recurring donors.

60x88mm (300 x 300 DPI)

HIS/ BSG FMT Guideline: Main Document, Gut version.

**The use of faecal microbiota transplant as treatment for recurrent or refractory *Clostridium difficile* infection and other potential indications: joint British Society of Gastroenterology (BSG) and Healthcare Infection Society (HIS) guidelines.**

Benjamin H Mullish<sup>\*1,2</sup>, Mohammed Nabil Quraishi<sup>\*3</sup>, Jonathan Segal<sup>\*1,4</sup>, Victoria L McCune<sup>5,6</sup>, Melissa Baxter<sup>7</sup>, Gemma L Marsden<sup>8</sup>, David Moore<sup>9</sup>, Alaric Colville<sup>7</sup>, Neeraj Bhala<sup>3,9,10</sup>, Tariq H Iqbal<sup>3,10</sup>, Christopher Settle<sup>11</sup>, Graziella Kontkowski<sup>12</sup>, Ailsa L Hart<sup>1,4</sup>, Peter M Hawkey<sup>6</sup>, Simon D Goldenberg<sup>○13,14</sup>, Horace RT Williams<sup>○□1,2</sup>.

1. Division of Integrative Systems Medicine and Digestive Disease, Department of Surgery and Cancer, Faculty of Medicine, Imperial College London, London, UK.
2. Departments of Gastroenterology and Hepatology, St Mary's Hospital, Imperial College Healthcare NHS Trust, Paddington, London, UK.
3. Department of Gastroenterology, Queen Elizabeth Hospital Birmingham, University Hospitals Birmingham NHS Foundation Trust, Birmingham, UK.
4. Inflammatory Bowel Disease Unit, St Mark's Hospital, Harrow, London, UK.
5. Public Health England, Public Health Laboratory Birmingham, Birmingham, UK.
6. Institute of Microbiology and Infection, University of Birmingham, Birmingham, UK.
7. Department of Microbiology, Royal Devon and Exeter NHS Foundation Trust, Exeter, UK.
8. Healthcare Infection Society, London, UK.
9. Institute of Applied Health Research, University of Birmingham, Birmingham, UK.
10. Institute of Translational Medicine, University of Birmingham, Edgbaston, Birmingham, UK.
11. Department of Microbiology, City Hospitals Sunderland NHS Foundation Trust, Sunderland, UK.
12. C diff Support, UK.
13. Centre for Clinical Infection and Diagnostics Research, King's College London, London, UK.
14. Department of Microbiology, Guy's and St Thomas' NHS Foundation Trust, London UK.

\*Joint first authors.

○Joint senior authors.

□Corresponding author.

HIS/ BSG FMT Guideline: Main Document, *Gut* version.

32 Contact via: Dr Horace Williams

33 Department of Gastroenterology

34 3<sup>rd</sup> Floor, Salton House

35 St Mary's Hospital, Imperial College Healthcare NHS Trust

36 London, W2 1NY

37 United Kingdom

38 Email: h.williams@imperial.ac.uk

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44 Abbreviations: FMT faecal microbiota transplant

45 CDI *Clostridium difficile* infection

46 EBV Epstein-Barr virus

47 CMV cytomegalovirus

48 BMI body mass index

49 GI gastrointestinal

50 RCT randomised controlled trial

51 NAAT nucleic acid amplification test

52 GDH glutamate dehydrogenase

53 EIA enzymes immunoassay

54 PCR polymerase chain reaction

55 IBD inflammatory bowel disease

56 IBS irritable bowel syndrome

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57	HIV	human immunodeficiency virus
58	AIDS	acquired immune deficiency syndrome
59	CPE	carbapenemase-producing <i>Enterobacteriaceae</i>
60	ESBL	extended-spectrum beta-lactamase
61	VRE	vancomycin-resistant <i>Enterococci</i>
62	MRSA	methicillin-resistant <i>Staphylococcus aureus</i>
63	PPI	proton pump inhibitor
64	UC	ulcerative colitis
65	HE	hepatic encephalopathy
66	MELD	Model for End-Stage Liver Disease

## 1. **Abstract:**

Interest in the therapeutic potential of faecal microbiota transplant (FMT) has been increasing globally in recent years, particularly as a result of randomised studies in which it has been used as an intervention. The main focus of these studies has been the treatment of recurrent or refractory *Clostridium difficile* infection (CDI), but there is also an emerging evidence base regarding potential applications in non-CDI settings. The key clinical stakeholders for the provision and governance of FMT services in the United Kingdom (UK) have tended to be in two major specialty areas: gastroenterology and microbiology/infectious diseases. Whilst the National Institute for Health and Care Excellence (NICE) guidance (2014) for use of FMT for recurrent or refractory CDI has become accepted in the UK, clear evidence-based UK guidelines for FMT have been lacking. This resulted in discussions between the British Society of Gastroenterology (BSG) and Healthcare Infection Society (HIS), and a joint BSG/HIS FMT working group was established. This guideline document is the culmination of that joint dialogue.

## 2. **Executive summary:**

### 2.1. **Overview:**

The remit of the British Society of Gastroenterology (BSG)/ Healthcare Infection Society (HIS) working group was to provide recommendations as to best practice in the provision of a faecal microbiota transplant (FMT) service. This guideline considers the use of FMT for the treatment of *Clostridium difficile* infection (CDI) – as well as for potential non-CDI indications – in adults. The working group have primarily targeted their report at clinicians involved in the use and provision of FMT services, but have also aimed it to be of interest to patients and their relatives.

### 2.2. **Summary of recommendations:**

#### 2.2.1. **Which patients with *Clostridium difficile* infection should be considered for faecal microbiota transplant, and how should they be followed up after treatment?**

##### 2.2.1.1. **Prior to faecal microbiota transplant. Patient selection:**

##### 2.2.1.1.1. **Recurrent *Clostridium difficile* infection:**

We recommend that FMT should be offered to patients with recurrent CDI who have had at least two recurrences, or those who have had one recurrence and have risk factors for further episodes, including severe and severe-complicated CDI (*GRADE of evidence: high; strength of recommendation: strong*).



**2.2.1.1.2. Refractory *Clostridium difficile* infection:**

We recommend that FMT should be considered in cases of refractory CDI (*GRADE of evidence: moderate; strength of recommendation: strong*).

**2.2.1.1.3. FMT as initial therapy for *Clostridium difficile* infection:**

We recommend that FMT should not be administered as initial treatment for CDI (*GRADE of evidence: low; strength of recommendation: strong*).

**2.2.1.1.4. Antimicrobial/ antitoxin therapy prior to considering FMT for patients with CDI:**

- i.* We recommend that FMT for recurrent CDI should only be considered after recurrence of symptoms following resolution of an episode of CDI that was treated with appropriate antimicrobials for at least 10 days (*GRADE of evidence: low; strength of recommendation: strong*).
- ii.* We recommend consideration of treatment with extended/ pulsed vancomycin and/or fidaxomicin before considering FMT as treatment for recurrent CDI (*GRADE of evidence: low; strength of recommendation: strong*).
- iii.* For those with severe or complicated CDI, which appears to be associated with reduced cure rates, we recommend that consideration should be given to offering patients treatment with medications which are associated with reduced risk of recurrence (e.g. fidaxomicin and bezlotoxumab), before offering FMT (*GRADE of evidence: low; strength of recommendation: strong*).

**2.2.1.2. Post-FMT follow-up, outcomes and adverse events:**

**2.2.1.2.1. Management of FMT failure:**

We recommend that FMT should be offered after initial FMT failure (*GRADE of evidence: high; strength of recommendation: strong*).

**2.2.1.2.2. General approach to follow-up post-FMT:**

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143 We recommend that all FMT recipients should routinely receive follow-up. Clinicians should  
144 follow-up FMT recipients for long enough to fully establish efficacy/adverse events, and for  
145 at least eight weeks in total (*GRADE of evidence: low; strength of recommendation: strong*).

#### 147 **2.2.1.2.3. Management of the FMT recipient:**

- 148 *i.* We recommend that immediate management after endoscopic administration of  
149 FMT should be as per endoscopy unit protocol (*GRADE of evidence: very low:*  
150 *strength of recommendation: strong*).
- 151 *ii.* We recommend that patients should be warned about short term adverse events, in  
152 particular the possibility of self-limiting GI symptoms. They should be advised that  
153 serious adverse events are rare (*GRADE of evidence: very low; strength of*  
154 *recommendation: strong*).
- 155 *iii.* After enteral tube administration, we recommend that patients may have the tube  
156 removed and oral water given from 30 minutes post-administration (*GRADE of*  
157 *evidence: very low; strength of recommendation: strong*).

#### 159 **2.2.1.2.4. Definition of cure post-FMT for CDI:**

160 We recommend that a decision regarding cure/remission from CDI should be recorded  
161 during follow-up. However, this has no uniformly-agreed definition, and should be decided  
162 on a case-by-case basis (*GRADE of evidence: very low; strength of recommendation: strong*).

#### 164 **2.2.1.2.5. Definition of treatment failure post-FMT for CDI:**

165 We recommend that treatment failure/recurrence should be defined on a case-by-case  
166 basis. Routine testing for *C. difficile* toxin after FMT is not recommended, but it is  
167 appropriate to consider in the case of persistent CDI symptoms/suspected relapse (*GRADE*  
168 *of evidence: low; strength of recommendation: strong*).

#### 170 **2.2.2. What recipient factors influence the outcome of faecal microbiota transplant when** 171 **treating people with *Clostridium difficile* infection?**

##### 172 **2.2.2.1. General approach to co-morbidities and FMT:**

- i. We recommend that FMT should be avoided in those with anaphylactic food allergy (GRADE of evidence: very low; strength of recommendation: strong).
- ii. We suggest that FMT should be offered with caution to patients with CDI and decompensated chronic liver disease (GRADE of evidence: very low; strength of recommendation: weak).

**2.2.2.2. Immunosuppression and FMT:**

- i. We recommend that FMT should be offered with caution to immunosuppressed patients, in whom FMT appears efficacious without significant additional adverse effects (GRADE of evidence: moderate; strength of recommendation: strong).
- ii. We recommend that immunosuppressed FMT recipients at risk of severe infection if exposed to EBV or CMV should only receive FMT from donors negative for EBV and CMV (GRADE of evidence: very low; strength of recommendation: strong).

**2.2.2.3. Other comorbidities and FMT:**

- i. We recommend that FMT should be offered to those with recurrent CDI and inflammatory bowel disease, but patients should be counselled about a small but recognised risk of exacerbation of IBD (GRADE of evidence: moderate; strength of recommendation: strong).
- ii. We recommend that FMT should be considered for appropriate patients with recurrent CDI regardless of other comorbidities (GRADE of evidence: moderate; strength of recommendation: strong).

**2.2.3. What donor factors influence the outcome of faecal microbiota transplant when treating people with Clostridium difficile infection?**

**2.2.3.1. General approach to donor selection:**

We recommend that related or unrelated donors should both be considered acceptable. However, where possible, FMT is best sourced from a centralised stool bank, from a healthy unrelated donor (GRADE of evidence: low; strength of recommendation: strong).

**2.2.3.2. Age and BMI restrictions for potential donors:**

204 We suggest that people should only be considered as potential FMT donors if they are  $\geq 18$   
205 and  $\leq 60$  years old, and have a BMI of  $\geq 18$  and  $\leq 30$  kg/m<sup>2</sup> (*GRADE of evidence: low; strength*  
206 *of recommendation: weak*).

### 2.2.3.3. General approach to the donor screening assessment:

209 It is mandatory to screen potential donors by questionnaire and personal interview, to  
210 establish risk factors for transmissible diseases and factors influencing the gut microbiota  
211 (**Table 1**) (*GRADE of evidence: low; strength of recommendation: strong*).

### 2.2.3.4. Laboratory screening of potential donors:

214 Blood and stool screening of donors is mandatory (**Tables 2 and 3**) (*GRADE of evidence: low;*  
215 *strength of recommendation: strong*).

### 2.2.3.5. Repeat donor checks, and donation pathway:

- 218 i. In centres using frozen FMT, before FMT may be used clinically, we recommend that  
219 donors should have successfully completed a donor health questionnaire and laboratory  
220 screening assays both before and after the period of stool donation. This is the  
221 preferred means of donor screening (*GRADE of evidence: low; strength of*  
222 *recommendation: strong*).
- 223 ii. In centres using fresh FMT, we recommend that a repeat health questionnaire should be  
224 assessed at the time of each stool donation. To ensure ongoing suitability for inclusion  
225 as a donor, the donor health questionnaire and laboratory screening should be repeated  
226 regularly (*GRADE of evidence: low; strength of recommendation: strong*).

## 2.2.4. What factors related to the preparation of the transplant influence the outcome of faecal microbiota transplant when treating people with *Clostridium difficile* infection?

### 2.2.4.1. General principles of FMT preparation:

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3 232 i. We recommend that stool collection should follow a standard protocol (*GRADE of*  
4 233 *evidence: low; strength of recommendation: strong*).  
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6 234 ii. We recommend that donor stool should be processed within 6 hours of defaecation  
7 235 (*GRADE of evidence: low; strength of recommendation: strong*).  
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9 236 iii. We recommend that both aerobically and anaerobically prepared FMT treatments  
10 237 should be considered suitable when preparing FMT for the treatment of recurrent  
11 238 CDI (*GRADE of evidence: moderate; strength of recommendation: strong*).  
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13 239 iv. We recommend that sterile 0.9% saline should be considered as an appropriate  
14 240 diluent for FMT production, and cryoprotectant such as glycerol should be added for  
15 241 frozen FMT (*GRADE of evidence: moderate: strength of recommendation: strong*).  
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17 242 v. We recommend using ≥50g of stool in each FMT preparation (*GRADE of evidence:*  
18 243 *moderate: strength of recommendation: strong*).  
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20 244 vi. We suggest that stool should be mixed 1:5 with diluent to make the initial faecal  
21 245 emulsion (*GRADE of evidence: low; strength of recommendation: weak*).  
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23 246 vii. We suggest that homogenisation and filtration of FMT should be undertaken in a  
24 247 closed disposable system (*GRADE of evidence: low; strength of recommendation:*  
25 248 *weak*).  
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45 255 **2.2.4.2. Fresh vs frozen FMT:**

46 251 We recommend that the use of banked frozen FMT material should be considered  
47 252 preferable to fresh preparations for CDI (*GRADE of evidence: high; strength of*  
48 253 *recommendation: strong*).  
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255 **2.2.4.3. Use of frozen FMT:**

- 256 i. We recommend that FMT material stored frozen at -80°C should be regarded as having a  
257 maximum shelf life of six months from preparation (*GRADE of evidence: low; strength of*  
258 *recommendation: strong*).  
259 ii. We suggest consideration of thawing frozen FMT at ambient temperature, and using  
260 within six hours of thawing (*GRADE of evidence: low; strength of recommendation:*  
261 *weak*).

iii. We suggest not thawing FMT in warm water baths, due to the risks of cross contamination with *Pseudomonas* (and other contaminants) and reduced bacterial viability (*GRADE of evidence: very low; strength of recommendation: weak*).

### **2.2.5. What factors related to administration of the transplant influence the outcome of faecal microbiota transplant when treating people with *Clostridium difficile* infection?**

#### **2.2.5.1. Use of specific medications in the period around FMT administration:**

##### **2.2.5.1.1. General principles of FMT administration:**

- i. We recommended that bowel lavage should be administered prior to FMT via the lower GI route, and that bowel lavage should be considered prior to FMT via the upper GI route; polyethylene glycol preparation is preferred (*GRADE of evidence: low; strength of recommendation: strong*).
- ii. For upper GI FMT administration, we suggest that a proton pump inhibitor should be considered, e.g. the evening before and morning of delivery (*GRADE of evidence: low; strength of recommendation: weak*).
- iii. We suggest that a single dose of loperamide (or other anti-motility drugs) should be considered following lower GI FMT delivery (*GRADE of evidence: low; strength of recommendation: weak*).
- iv. We suggest that prokinetics (such as metoclopramide) should be considered prior to FMT via the upper GI route (*GRADE of evidence: low; strength of recommendation: weak*).
- v. We recommend that best practice for prevention of further transmission of CDI should be applied throughout when administering FMT to patients with CDI (nursing with enteric precautions, sporicidal treatment of endoscope, etc) (*GRADE of evidence: high; strength of recommendation: strong*).

##### **2.2.5.1.2. Additional antibiotics pre-FMT:**

We recommend the administration of further antimicrobial treatment for CDI for at least 72 hours prior to FMT (*GRADE of evidence: low; strength of recommendation: strong*).

**2.2.5.1.3. Washout period between antibiotic use and FMT:**

- i.* To minimise any deleterious effect of antimicrobials on the FMT material, we recommend that there should be a minimum washout period of 24 hours between the last dose of antibiotic and treatment with FMT (*GRADE of evidence: low; strength of recommendation: strong*).
- ii.* We suggest considering consultation with infectious disease specialists or medical microbiologists for advice whenever FMT recipients also have an indication for long-term antibiotics, or have an indication for non-CDI antibiotics within eight weeks of FMT (*GRADE of evidence: very low; strength of recommendation: weak*).

**2.2.5.2. Route of FMT delivery:**

**2.2.5.2.1. Upper gastrointestinal tract administration of FMT:**

- i.* We recommend that upper GI administration of FMT as treatment for recurrent or refractory CDI should be used where clinically appropriate (*GRADE of evidence: high; strength of recommendation: strong*).
- ii.* Where upper GI administration is considered most appropriate, we recommend that FMT administration should be via nasogastric, nasoduodenal, or nasojejunal tube, or alternatively via upper GI endoscopy. Administration via a permanent feeding tube is also appropriate (*GRADE of evidence: high; strength of recommendation: strong*).
- iii.* We recommend that no more than 100ml of FMT is administered to the upper GI tract (*GRADE of evidence: low; strength of recommendation: strong*).
- iv.* We recommend that upper GI administration of FMT should be used with caution in those at risk of regurgitation and/ or those with swallowing disorders (*GRADE of evidence: low; strength of recommendation: strong*).

**2.2.5.2.2. Lower gastrointestinal tract administration of FMT:**

- i.* We recommend that colonoscopic administration of FMT as treatment for recurrent or refractory CDI should be used where appropriate (*GRADE of evidence: high; strength of recommendation: strong*).



- 322 *ii.* Where colonoscopic administration is used, we suggest considering preferential  
323 delivery to the caecum or terminal ileum, as this appears to give the highest efficacy  
324 rate (*GRADE of evidence: low; strength of recommendation: weak*).
- 325 *iii.* We recommend that FMT via enema should be used as a lower GI option when  
326 delivery using colonoscopy or flexible sigmoidoscopy is not possible (*GRADE of*  
327 *evidence: high; strength of recommendation: strong*).

#### 329 **2.2.5.2.3. Capsulised FMT:**

330 Capsulised FMT holds promise as a treatment option for recurrent CDI and we recommend  
331 that this should be offered to patients as a potential treatment modality where available.  
332 Capsule preparations should follow a standard protocol. Further evidence regarding  
333 optimal dosing and formulation is required (*GRADE of evidence: high; strength of*  
334 *recommendation: strong*).

#### 336 **2.2.6. What is the clinical effectiveness of FMT in treating conditions other than** 337 ***Clostridium difficile* infection?**

338 We do not currently recommended FMT as treatment for inflammatory bowel disease.  
339 Apart from CDI, there is insufficient evidence to recommend FMT for any other  
340 gastrointestinal or non-gastrointestinal disease (*GRADE of evidence: moderate; strength of*  
341 *recommendation: strong*).

#### 343 **2.2.7. Basic requirements for implementing a FMT service:**

##### 344 **2.2.7.1. General considerations:**

- 345 *i.* The development of FMT centres should be encouraged (*GRADE of evidence: very*  
346 *low; strength of recommendation: strong*).
- 347 *ii.* We suggest that FMT centres should work to raise awareness about FMT as a  
348 treatment option amongst clinicians caring for patients with CDI, and provide  
349 training to relevant healthcare professionals on the practicalities of delivering an  
350 FMT service (*GRADE of evidence: very low; strength of recommendation: weak*).



**2.2.7.2. Legal aspects and clinical governance:**

In the UK, FMT must be manufactured in accordance with MHRA guidance for human medicines regulation. When FMT is supplied on a named patient basis, within a single organisation, a pharmacy exemption may be used, subject to ensuring proper governance and traceability. All centres that are processing and distributing FMT should seek guidance from the MHRA and where necessary obtain appropriate licenses prior to establishing an FMT service. This is a legal requirement. In countries other than the UK, FMT should only be manufactured following appropriate approval from the national authority of that country (*GRADE of evidence: very low; strength of recommendation: strong*).

**2.2.7.3. Multidisciplinary teams:**

We recommend that a multidisciplinary team should be formed to deliver FMT services (*GRADE of evidence: very low; strength of recommendation: strong*).

**2.2.7.4. Infrastructure:**

We recommend utilisation of suitable laboratory facilities and infrastructure for FMT production (*GRADE of evidence: very low; strength of recommendation: strong*).

**2.2.7.5. FMT manufacturing:**

We recommend ensuring the traceability of supply (*GRADE of evidence: very low; strength of recommendation: strong*).

**2.2.7.6. FMT production quality control:**

We recommend monitoring, notification and investigation of all adverse events and reactions related to FMT (*GRADE of evidence: very low; strength of recommendation: strong*).

**2.2.7.7. Donor screening governance:**

We recommend ensuring the clinical governance of donor screening (*GRADE of evidence: very low; strength of recommendation: strong*).

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**3. Introduction:**

The aim of the BSG/ HIS FMT working group was to establish a guideline that defined best practice in all aspects of a FMT service, by providing evidence-based recommendations wherever possible, and consensus multi-disciplinary expert opinion where specific published evidence is currently lacking. This included the evaluation of the use of FMT in the treatment of *Clostridium difficile* infection (CDI; also referred to as *Clostridioides difficile*<sup>1</sup>), and also in potential non-CDI indications. Relevant guidance published to date includes the interventional procedure guidance from the National Institute for Health and Care Excellence (NICE)<sup>2</sup>, UK, European and US microbiological guidelines on the treatment of *Clostridium difficile* infection (CDI)<sup>3–5</sup>, and recent expert consensus documents on FMT in clinical practice<sup>6,7</sup>. Furthermore, there have also been national recommendations regarding FMT produced by working groups in several different countries<sup>8–10</sup>. Principally as a result of randomised studies that have been published in recent years<sup>11–18</sup>, FMT has become an accepted treatment for recurrent/refractory CDI.

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The unique remit and objectives of this guideline when commissioned by the BSG and HIS was:

i. To review the rapidly-growing body of randomised trial evidence for the efficacy of FMT in the treatment of adults (≥18 years), both in CDI and in other clinical conditions, much of which has been published after the publication of current CDI treatment algorithms<sup>3,4</sup>.

ii. To provide specific guidance about best practice for an FMT service within the context of the regulatory framework for the intervention as it currently exists in the UK<sup>19,20</sup>.

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The elucidation of the mechanisms underlying the efficacy of FMT in treating CDI remains an active area of global research, with the aim of rationalising FMT from its current crude form to a more targeted, refined therapeutic modality<sup>21</sup>. Previous research has demonstrated that commensal bacteria cultured from the stool of healthy donors<sup>22</sup>, sterile faecal filtrate<sup>23</sup>, and/ or spores of *Firmicutes* derived from ethanol-treated stool from healthy donors<sup>24</sup>, may have similar efficacy to conventional FMT in treating CDI, although results of the latter approach produced disappointing outcome data when extended to a Phase II clinical trial<sup>25</sup>. For the purposes of this guideline, the BSG/HIS working group considered only studies that used the administration of manipulated whole stool (including encapsulated faeces). They deemed studies using cultured microorganisms (or their

proteins, metabolites or other components), or microbiota suspensions, to be in the pre-clinical research stage, without firm evidence.

FMT has been shown to be very acceptable to patients, both in the setting of CDI<sup>11,26</sup> and in non-CDI settings, e.g. ulcerative colitis<sup>27</sup>. However, the absence of appropriate protocols<sup>28–31</sup> specifically taking into account UK clinical practice and regulation of FMT has been perceived as a barrier to the use of FMT in the UK and Ireland; these guidelines seek to rectify this problem.

**4. Guideline development:**

**4.1. Guideline development team**

BSG and HIS commissioned the authors to undertake the Working Party Report. The authors represent the membership of both societies. The working group included gastroenterologists, infectious diseases/microbiology clinicians, a clinical scientist, a systematic reviewer, and patient representatives. The views expressed in this publication are those of the authors, and have been endorsed by BSG and HIS following consultation.

**4.2. Scope of the guidelines**

The main scope of the guidelines is to provide guidance for the optimal provision of an effective and safe FMT service, principally for recurrent or refractory CDI, but non-CDI indications are also considered. These guidelines only apply to adult patients (≥18 years); the working party did not consider the role of FMT in the treatment of either CDI or non-CDI indications in children or young people. The guidelines were written with a focus upon UK practice, but also with consideration of more global practice as it applied. The diagnosis and management of *Clostridium difficile* infection in general are outside the remit of these guidelines.

**4.3. Evidence appraisal**

Questions for review were derived from the Working Party Group, which included patient representatives in accordance with the PICO process<sup>32</sup>. To prepare these recommendations, the working group collectively reviewed relevant peer-reviewed research.

#### 4.4. Data sources and search strategy

A systematic literature search was undertaken using MEDLINE, EMBASE databases and Cochrane Library for relevant articles published from 1<sup>st</sup> January 1980 to 1<sup>st</sup> January 2018. The MEDLINE and EMBASE strategy are shown in **Supplementary Material 1, Appendix 2ii**. Free text and MESH/ index terms for faecal microbial transplant and *Clostridium difficile* or other diseases of interest were combined. In addition, conference proceedings from microbiology, infectious disease, and gastroenterology conferences were also searched to identify additional studies.

#### 4.5. Study eligibility and selection criteria

The members of the guideline group determined criteria for study inclusion. Two reviewers (BHM, MNQ) screened the titles and abstracts of each article for relevance independently; any disagreements were resolved by discussion with a third reviewer (JPS). Copies of relevant articles were obtained and assessed for inclusion as evidence in the guideline by all three reviewers. The reason for not selecting studies was recorded. Only articles published in English and human clinical studies were included. For evidence on FMT for CDI, both randomised studies (including randomised controlled trials (RCTs)) and case series with at least 10 patients were selected. Only randomised trials were included as evidence for FMT for non-CDI indications. Conference abstracts were only included for CDI and non-CDI indications if they reported a randomised trial; where abstracts were available reporting data from a randomised trial that was subsequently published, only the published paper was reviewed.

#### 4.6. Data extraction and quality assessment

The initial search identified 2658 publications, and of these, 802 duplicates were excluded. 1856 studies were subsequently screened, from which 78 studies were assessed by reviewing the full text for eligibility (see **Supplementary Material 1, Appendix 2iii** and **Supplementary Material 2, Additional Appendix D**). Of these 78 studies, 58 studies were included as the basis of evidence for writing this guideline. In total, 39 were case studies in CDI including at least 10 patients (see **Supplementary Material 2, Additional Appendix C.1**), and ten were randomised studies in CDI (see **Supplementary Material 2, Additional Appendix C.2**). Nine were randomised trials for non-CDI indications (see **Supplementary Material 2, Additional Appendix C.3**). Data were extracted for patient demographics, disease characteristics, donor screening characteristics, stool preparation and administration, clinical outcomes and adverse events. The quality of randomised studies was

assessed with the Cochrane Collaboration’s risk of bias tool. Case series were assessed using the Centre for Reviews and Dissemination guidance.

**4.7. Rating of evidence and recommendations**

The BSG version of these guidelines was prepared in keeping with the BSG Clinical Services & Standards Committee (CSSC) advice document on the writing of clinical guidelines<sup>33</sup>. Evidence tables were presented and discussed by the working group, and guidelines were prepared according to the nature and applicability of the evidence regarding efficacy and patient preference and acceptability. For the BSG version of this guideline, the GRADE system (Grades of Recommendation Assessment, Development and Evaluation)<sup>34</sup> was used to assess the strength of evidence (high/ moderate/ low/ very low) and strength of recommendation (strong/ weak) (**Table 4**). The section entitled ‘Basic requirements for implementing an FMT service’ (**Supplementary Material 3**) was based on expert opinion, since this was a key area of the working party’s remit but not one amenable to evaluation by the PICO process. Face-to-face meetings and group teleconferences were held to agree on recommendations. Any disagreements on recommendations or the strength of recommendation were resolved by discussion and, where necessary, voting by the members of the working group, with consensus achieved when >80% were in agreement.

**4.8. Consultation process**

Feedback on draft guidelines was received from the Scientific Development Committee (SDC) of HIS, and changes made. These guidelines were then opened to consultation with relevant stakeholders (see **Supplementary Material 1, Appendix 3** of this document). The draft report was available on the HIS website for one month. Views were invited on format, content, local applicability, patient acceptability, and recommendations. The working group reviewed stakeholder comments, and collectively agreed revisions. Final changes were made after repeat reviews from HIS (Chair of the SDC and HIS Council) and BSG (BSG CSSC and BSG Council), and after further external peer review.

**4.9. Guideline accreditation and scheduled review**

The guidelines will be reviewed at least every four years and updated if change(s) in the evidence are sufficient to require a change in practice.

#### 4.0. Additional information:

Additional information related to this guideline (including a lay summary, background on the working party report, and information on the implementation of these guidelines) is contained within **Supplementary Material 1, Section 1**.

### 5. Rationale for recommendations:

#### 5.1. Which patients with *Clostridium difficile* infection should be considered for faecal microbiota transplant, and how should they be followed up after treatment?

##### 5.1.1. Prior to faecal microbiota transplant. Patient selection:

##### 5.1.1.1. Recurrent *Clostridium difficile* infection:

As already described, there is widespread consensus that FMT is an efficacious treatment for recurrent CDI. In defining recurrent CDI, some studies have relied on a minimum threshold of return of clinical symptoms (e.g. at least three unformed bowel movements within 24 hours, for at least two consecutive days)<sup>12,18</sup> following previous successful CDI treatment; most studies have also included a requirement for a positive microbiological test<sup>12,14,18,35–45</sup>. Other studies explicitly state that a positive test was not required<sup>46</sup>. Recommendations for CDI testing are beyond the scope of this guideline, and there are already well-established evidence-based guidelines<sup>47</sup>. These recommend testing with either a nucleic acid amplification test (NAAT) or GDH assay, followed by detection of free toxin (either by toxin A/B enzyme immunoassay (EIA) or cytotoxin neutralisation assay), which allows differentiation of patients with active disease as well as those who are likely colonised<sup>47</sup>. However, the working group discussed the importance of the accurate diagnosis of true recurrent CDI prior to consideration of FMT; in particular, they noted a study which observed that of 117 patients with presumed recurrent CDI referred for work-up for FMT, 25% ( $n=29/117$ ) were determined to have a non-CDI diagnosis, with irritable bowel syndrome ( $n=18$ ) and inflammatory bowel disease ( $n=3$ ) being the most common alternative diagnoses, and younger patients more likely to be misdiagnosed<sup>48</sup>.

All of the reviewed studies have included patients with recurrent CDI, however some studies offered FMT to patients at the first recurrence (second episode)<sup>12,15,16,18,35,37,42,43,46,49</sup>, whereas others offered FMT after the second recurrence (third episode)<sup>13,14,39,41,44,45,50,51</sup>. Some protocols offered FMT after three or more recurrences<sup>52</sup>, whilst others did not define the point at which it was administered<sup>40,53</sup>.

The severity of infection has been used as a parameter to decide at which stage FMT is offered. Youngster *et al.* offered FMT to patients with at least three episodes of mild to moderate CDI, or at least two episodes of severe CDI resulting in hospitalisation and associated with significant morbidity<sup>17</sup>. Another study selected patients for FMT using four categories of severity, which also accounted for prior anti-CDI therapy and requirement for hospitalisation<sup>54</sup>.

None of the studies directly compared the efficacy of FMT according to the stage at which it was offered (i.e. first recurrence vs.  $\geq$  two recurrences). A small number of studies<sup>55–57</sup> included patients with severe CDI (defined as hypoalbuminaemia with increased peripheral white cell count and/or abdominal tenderness) or complicated CDI (defined as admission to Intensive Care, altered mental status, hypotension, fever, ileus, white blood cell count  $> 30 \times 10^9/l$ , lactate  $> 2.2\text{mmol/l}$ , or evidence of end organ damage). A single study described an apparent lower rate of treatment success when FMT was used to treat patients with recurrent CDI with disease caused by ribotype 027<sup>43</sup>, but this is the case for all anti-CDI treatment modalities for this ribotype in comparison to others. The working group agreed that there was insufficient evidence to suggest that *C. difficile* ribotype should influence whether or not FMT is offered.

A lower primary cure rate was reported for complicated CDI (66%) compared with recurrent CDI (82%) and severe CDI (91%) in one study<sup>55</sup>; in a case series of 17 patients who all had severe and/or complicated CDI, a primary cure rate of 88% was described<sup>57</sup>. A cohort of 328 patients was analysed to determine which factors were associated with failure of FMT<sup>58</sup>. Higher early (one month) failure rates were found in patients with severe (72%,  $n=19/25$ ) or severe-complicated (52.9%,  $n=9/17$ ) CDI than for recurrent CDI (11.9%,  $n=34/286$ ). This study also identified that patients who were treated with FMT as an inpatient were nearly four times more likely to fail as those who had FMT as an outpatient; however, the working group noted that the authors of this study themselves identified that inpatient status is likely a proxy of severity of CDI and/or co-morbidities. A further similar study, including 64 patients treated with FMT as treatment for recurrent CDI, also identified severe CDI as the strongest independent risk factor for FMT failure on multivariate analysis<sup>59</sup>.

The working group discussed their experience of treating patients with CDI whose disease fitted an intermediate pattern to the typical descriptions given of recurrent or refractory CDI, e.g. patients with CDI who have some (but incomplete) symptomatic improvement with anti-CDI antibiotics and worsening of disease when these are stopped. The experience of the working group was that such



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571 patients experienced excellent responses to FMT, and that these patients should be considered for  
572 FMT.

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574 As FMT is currently an unlicensed medicine with poorly-studied long term sequelae, the working  
575 group considered that it should generally be reserved for patients who have had three or more  
576 episodes of infection. There are no studies directly comparing its effectiveness with some of the  
577 newer agents such as fidaxomicin or bezlotoxumab, hence this recommendation is made on the  
578 basis of safety. However, the working group agreed that it may be reasonable in certain patient  
579 groups with ongoing risk factors for further recurrence to offer FMT after the second episode.

580

581 **Recommendation:**

582 ***We recommend that FMT should be offered to patients with recurrent CDI who have had***  
583 ***at least two recurrences, or those who have had one recurrence and have risk factors for***  
584 ***further episodes, including severe and severe-complicated CDI (GRADE of evidence: high;***  
585 ***strength of recommendation: strong).***

586

587 **5.1.1.2. Refractory *Clostridium difficile* infection:**

588 Two randomised trials allowed the recruitment of patients with refractory CDI. The first defined this  
589 as at least three weeks of ongoing severe symptoms despite standard antimicrobial therapy for  
590 CDI<sup>17</sup>. The second required persistent or worsening diarrhoea and one of the following: ongoing  
591 abdominal pain, fever > 38°C, or white blood cell count > 15x 10<sup>9</sup>/l despite oral vancomycin at a dose  
592 of 500mg four times daily for at least five days<sup>16</sup>. Both studies included only small numbers of  
593 patients with refractory CDI (*n*=4/20 (20%) and *n*=15/219 (6.8%), respectively). There did not appear  
594 to be any significant difference in primary outcome measure (clinical cure) in patients with recurrent  
595 or refractory CDI, although neither study was designed to assess this difference. There are also a  
596 number of case series in which FMT was given to patients with refractory CDI; however, outcome  
597 measures were not reported for these groups individually in these studies<sup>37,38,54,60</sup>.

598

599 Overall, the working group concluded that there is little consensus on the definition of refractory  
600 CDI, with some studies using the terms 'refractory' and 'recurrent' interchangeably (as well as other  
601 terms, e.g. 'salvage therapy'). Consequently, the quality of evidence for the utility of FMT in



refractory cases of CDI is lower than for recurrent CDI. The standardisation of definitions will allow more robust comparison between patient cohorts.

**Recommendation:**

***We recommend that FMT should be considered in cases of refractory CDI (GRADE of evidence: moderate; strength of recommendation: strong).***

**5.1.1.3. FMT as initial therapy for *Clostridium difficile* infection:**

Experience of the use of FMT as initial therapy for CDI is very limited. In a case series of patients with CDI with ribotype 027, use of anti-CDI antibiotics together with nasogastric FMT within a week of diagnosis during an initial episode of CDI was associated with reduced mortality when compared to using FMT only after the failure of three courses of antibiotics (mortality of 18.75% ( $n=3/16$  patients) vs 64.4% ( $n=29/45$  patients))<sup>61</sup>. However, 37.5% ( $n=6/16$ ) of the patients treated with FMT within a week of CDI diagnosis required further antibiotics and a second FMT within one month of the first FMT because of relapse<sup>61</sup>. In a small pilot randomised trial, patients were randomised to either vancomycin or multi-donor FMT (administered either via upper or lower GI routes) as initial therapy for CDI; CDI resolution occurred in 88.9% ( $n=8/9$ ) patients with vancomycin, compared to 57.1% of patients ( $n=4/7$ ) patients with one FMT, and 71.4% of patients ( $n=5/7$ ) after two FMTs<sup>62</sup>. Given the small size of these studies and equivocal results, the working group concluded that the reviewed studies did not support FMT as initial therapy for CDI.

**Recommendation:**

***We recommend that FMT should not be administered as initial treatment for CDI (GRADE of evidence: low; strength of recommendation: strong).***

**5.1.1.4. Antimicrobial/ antitoxin therapy prior to considering FMT for patients with CDI:**

There are now at least two licensed agents (fidaxomicin and bezlotoxumab) which have been shown to significantly reduce the risk of recurrence compared with vancomycin<sup>63,64</sup>. There is also some evidence that pulsed/tapered dosing of vancomycin and fidaxomicin (including pulsed fidaxomicin<sup>65</sup>) results in fewer recurrences than with standard dosing of these agents<sup>66,67</sup> (although this finding has

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not been replicated in all studies<sup>68</sup>). Pre-planned subgroup analysis of patients with severe CDI in a randomised trial demonstrated a significantly lower recurrence rate when treated with fidaxomicin (13.0%,  $n=12/92$ ) than when treated with vancomycin (26.6%,  $n=29/209$ )<sup>63</sup>; this finding was replicated in another randomised controlled trial, with 8.3% ( $n=4/48$ ) and 32.6% ( $n=14/43$ ) experiencing a recurrence respectively<sup>69</sup>. In a further randomised trial, bezlotoxumab (together with standard of care antibiotics) was shown to reduce recurrence of severe CDI compared to standard of care antibiotics alone (10.9% ( $n=6/55$ ) vs 20% ( $n=13/65$ ) respectively)<sup>64</sup>.

As discussed above, the working group noted that there are no studies comparing FMT to fidaxomicin or bezlotoxumab, and only one study comparing a vancomycin taper to FMT<sup>12</sup>. The working group agreed that in the absence of this evidence, on the balance of safety and potential risks, consideration should be given to using antimicrobial/antitoxin therapy associated with reduced CDI recurrence prior to considering the use of FMT.

Several studies specify that patients should be treated with anti-*C. difficile* antibiotics for a minimum period of 10 days before diagnosing recurrent CDI and offering FMT<sup>12,15,16,18</sup>.

#### **Recommendations:**

- i. We recommend that FMT for recurrent CDI should only be considered after recurrence of symptoms following resolution of an episode of CDI that was treated with appropriate antimicrobials for at least 10 days (GRADE of evidence: low; strength of recommendation: strong).***
- ii. We recommend consideration of treatment with extended/ pulsed vancomycin and/or fidaxomicin before considering FMT as treatment for recurrent CDI (GRADE of evidence: low; strength of recommendation: strong).***
- iii. For those with severe or complicated CDI, which appears to be associated with reduced cure rates, we recommend that consideration should be given to offering patients treatment with medications which are associated with reduced risk of recurrence (e.g. fidaxomicin and bezlotoxumab), before offering FMT (GRADE of evidence: low; strength of recommendation: strong).***

#### **5.1.2. Post-FMT follow-up, outcomes and adverse events:**

##### **5.1.2.1. Management of FMT failure:**

Where patients were deemed not to have responded to an initial FMT, many studies have offered repeat FMT and success rates have been excellent even in patients with modest response to a first FMT<sup>14,15,17,18,35,43,46,51,54,70,71</sup>. The success of a second FMT appears to be high whether treatment failure represents non-response to the first FMT, or a late failure (i.e. further relapse of CDI after an initial response); however, these terms have been defined variably between different studies (also see **Section 5.1.2.5**). Second FMTs have been offered as soon as 24-72 hours after an initial FMT for presumed non-response<sup>37,72,73</sup>. For FMT failure in patients with pseudomembranous colitis, repeat FMT every three days until resolution of pseudomembranes has been a successful approach<sup>18</sup>. Good outcomes in pseudomembranous disease have also been achieved through a protocol that routinely restarted five days of vancomycin if FMT failed, before offering another FMT<sup>73</sup>. Other studies have demonstrated potential success in treating initial FMT failure with further antibiotics, including repeat FMT with vancomycin between procedures<sup>42</sup>, or anti-CDI antibiotics alone<sup>35,42,43,45,51,70,71</sup>. Patients unresponsive to two FMTs have been offered further FMT or antibiotic therapy<sup>16</sup>, or even the administration of intravenous immunoglobulin<sup>35</sup>. Whilst the working group collectively agreed that there was strong evidence to recommend repeat FMT after initial FMT failure, they were not able to recommend a specific protocol for administering repeat FMT and/ or maximum number of FMTs, given the wide heterogeneity of approach described within the reviewed literature.

**Recommendation:**

***We recommend that FMT should be offered after initial FMT failure (GRADE of evidence: high; strength of recommendation: strong).***

**5.1.2.2. General approach to follow-up post-FMT:**

Follow-up post-FMT (in terms of duration, modality and regimen for follow-up) varies considerably between studies, and is largely dependent upon study design. Follow-up regimens vary not only between studies but within them too, reflecting the retrospective nature of many early FMT studies in CDI, where follow-up mostly reflected pragmatic routine clinical care.

Modalities of follow-up have included outpatient review<sup>14,43,58,71,74-76</sup>, telephone interview<sup>17,39,43,46,58,71,74</sup> and case note/ database review<sup>35,39,70,71,74,40,42,43,45,46,49,51,54</sup>. Follow-up duration has varied from 60 days<sup>45</sup> to 8 years<sup>36</sup>, with very different durations used in each study. Once again, however, this variability in follow-up largely reflects the retrospective analysis of case

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series rather than being justified by any specific methodology. The working group decided by consensus that at least eight weeks of follow-up was appropriate post-FMT to fully assess efficacy and potential adverse events; this figure was also influenced by discussions regarding the timepoint after FMT at which a decision could be made regarding cure/ remission of CDI (see **Section 5.1.2.4**).

**Recommendation:**

***We recommend that all FMT recipients should routinely receive follow-up. Clinicians should follow-up FMT recipients for long enough to fully establish efficacy/adverse events, and for at least eight weeks in total (GRADE of evidence: low; strength of recommendation: strong).***

**5.1.2.3. Management of the FMT recipient:**

Procedural adverse events during administration of FMT have predominantly occurred with colonoscopic administration of FMT. These have included mild nausea and vomiting attributed to sedation for the colonoscopy, minor mucosal tears during colonoscopy<sup>49,60</sup>, and microperforation following biopsy of an area of presumed ischaemic small bowel injury in a patient with chronically dilated small bowel (which resolved with conservative management<sup>46</sup>). One death occurred due to witnessed aspiration at the time of colonoscopy<sup>60</sup>. Faecal regurgitation and vomiting with temporal association to upper GI FMT administration has also been described (discussed further in **Section 5.5.2.2**)<sup>77</sup>.

The predominant short term adverse events post-FMT for CDI are mild: self-limiting GI symptoms have been the most frequently reported adverse events. These may be related to the route of administration and include belching<sup>15</sup>, nausea<sup>15,16,49,60</sup>, abdominal cramps/ discomfort/ bloating/ pain<sup>15,18,49,60,72</sup>, and diarrhoea<sup>15,16,18,60</sup>. One patient with a history of autonomic dysfunction experienced dizziness with diarrhoea after FMT<sup>15</sup>. These symptoms are typically short-lived, resolving in hours to days<sup>15,16,18,49,72</sup>. Minor subsequent adverse events have included a range of GI side effects including self-limiting abdominal discomfort<sup>14,17,57,76</sup>, nausea<sup>14,49,70</sup>, flatulence<sup>14,16,17,41,42,49,57</sup>, self-limiting irregular bowel movements<sup>41</sup>, *C. difficile*-toxin negative diarrhoea<sup>52,55</sup>, constipation<sup>14,15,42,55,70</sup> and constitutional symptoms/ temperature disturbance<sup>14,17</sup>.

As such, immediately post-endoscopic administration of FMT, most FMT centres typically manage patients using standard protocols for an endoscopic procedure<sup>41,49</sup>, without any specific adaptations (apart from to reiterate advice about the possibility of self-limiting GI side effects, and the use of departmental infection control protocols). There is often a relatively short period of post-procedural observation<sup>15,18</sup>. Most studies allow patients to leave the administration site after the period of observation, although overnight observation was the protocol used for a cohort of very elderly patients with multiple comorbidities<sup>51</sup>. Where enteral tube administration is used, post-procedure management has ranged between removal of the tube after 30 minutes (following nasoenteral administration of 500ml of FMT<sup>15</sup>) to prompt post-procedure removal and oral water administration (after nasogastric administration of 90ml of FMT<sup>72</sup>), with no direct adverse outcomes in either case. The working group felt that removal of the tube at 30 minutes, with administration of water at this point, was a pragmatic approach.

The definition of post-FMT serious adverse events has varied between studies, but has included significant morbidity necessitating hospital admission and death in the follow up period. Many of these events are described as not directly caused by the FMT, including the scenario of post-FMT severe CDI recurrences<sup>72</sup> and probable or certain CDI-related deaths<sup>16,60,70</sup> occurring in the context of FMT failure, or deaths related to patient comorbidities<sup>17,55</sup>. One patient was admitted to hospital with self-limiting abdominal pain post-FMT<sup>60</sup>, and four patients with flares of inflammatory bowel disease<sup>60</sup>. Three patients underwent colectomy during the post-FMT follow-up period, with all related to ulcerative colitis and not believed to be due to CDI<sup>60</sup>. Other reported serious adverse events include recurrent urinary tract infection<sup>15</sup>, fever during haemodialysis<sup>15</sup> and upper gastrointestinal haemorrhage after nasogastric FMT (in a patient taking NSAIDs<sup>51</sup>), none of which were thought to be strongly linked to FMT. There have also been a number of new onset autoimmune, inflammatory and metabolic conditions described post-FMT, although these have been described from single centres only, with these findings not replicated elsewhere. Such conditions include microscopic colitis, Sjögren's syndrome, follicular lymphoma, peripheral neuropathy, immune thrombocytopenia and rheumatoid arthritis<sup>53,55</sup>.

Significant adverse events are therefore rare but well-described. Furthermore, the procedure is relatively novel, and longer-term follow-up data regarding safety are required. Therefore, the working group opined that formal follow-up post-FMT to assess outcome and possible adverse events is essential.

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The use of questionnaires to compare symptoms pre- and post-FMT is common. Specifically, data collected have included clinical response to symptom severity<sup>55</sup>, stool frequency<sup>15,17,46,55,57,72</sup>, stool consistency<sup>14,15,72</sup>, abdominal pain or tenderness<sup>55,57</sup>, rating of gastrointestinal symptoms<sup>72</sup>, general well-being<sup>55,72</sup>, days to improvement post-FMT<sup>57</sup>, weight change<sup>72</sup>, functional status<sup>55</sup>, and changes in medication/use of antibiotics<sup>57,72</sup>. Additionally, certain patients have been given specific advice post-FMT to contact their clinical team if there is recurrence of diarrhoea or symptoms<sup>14,35,41,43</sup>. Where patients underwent outpatient clinical evaluation, this was generally undertaken relatively early post-FMT<sup>39,52,76</sup>. In one study, patients were additionally given instructions for cleaning and disinfection at home, with the aim of reducing the possibility of *C. difficile* reinfection<sup>43</sup>, and counselling on the risk of recurrent CDI with future antibiotic courses<sup>76</sup>.

#### **Recommendations:**

- i. ***We recommend that immediate management after endoscopic administration of FMT should be as per endoscopy unit protocol (GRADE of evidence: very low; strength of recommendation: strong).***
- ii. ***We recommend that patients should be warned about short term adverse events, in particular the possibility of self-limiting GI symptoms. They should be advised that serious adverse events are rare (GRADE of evidence: very low; strength of recommendation: strong).***
- iii. ***After enteral tube administration, we recommend that patients may have the tube removed and oral water given from 30 minutes post-administration (GRADE of evidence: very low; strength of recommendation: strong).***

#### **5.1.2.4. Definition of cure post-FMT for CDI:**

It is recognised that symptoms of CDI resolve relatively promptly post-successful FMT, although this has been variably described (within hours in some studies<sup>52</sup>, at an average of 4-5 days in others<sup>57,71</sup>). Treatment success post-FMT for CDI has no uniformly-agreed definition, with the time point at which cure/ remission is defined on clinical grounds varying between 3-5 days<sup>36</sup> up to six months<sup>42</sup>. A consensus document from the USA recommends 'resolution of symptoms as a primary end point; absence within eight weeks of FMT as a secondary end point'<sup>78</sup>. The working group recommended that this definition should be made on a case-by-case basis; however, they agreed that an

assessment for cure/ remission of CDI within eight weeks post-FMT was reasonable in most cases, and therefore that this was also a reasonable minimum length of time to undertake follow-up post-FMT (see **Section 5.1.2.2**).

**Recommendation:**

***We recommend that a decision regarding cure/remission from CDI should be recorded during follow-up. However, this has no uniformly-agreed definition, and should be decided on a case-by-case basis (GRADE of evidence: very low; strength of recommendation: strong).***

**5.1.2.5. Definition of treatment failure post-FMT for CDI:**

There is no uniformly-agreed definition of treatment failure/recurrence post-FMT for CDI, with varied definitions used in studies. The use of *C. difficile* toxin as a marker of treatment success or failure is variable, with some studies opting not to test for CDT unless symptoms consistent with CDI recurred<sup>49,52–54,60,72,74</sup>. Some studies have routinely performed CDT testing without specifying any action taken after a positive result<sup>14,15,18,36,39,41</sup>, whilst others have tested for *C. difficile* PCR but relied on clinical criteria (even if PCR was positive) post-FMT for evaluating FMT efficacy<sup>14</sup>. A recent prospective study from the USA identified that only 3% (3/129) of patients who were asymptomatic at four weeks post-FMT for recurrent CDI had positive *C. difficile* PCR, again emphasising that symptoms rather than laboratory assays are more useful contributors to establishing FMT success<sup>79</sup>.

**Recommendation:**

***We recommend that treatment failure/recurrence should be defined on a case-by-case basis. Routine testing for C. difficile toxin after FMT is not recommended, but it is appropriate to consider in the case of persistent CDI symptoms/suspected relapse (GRADE of evidence: low; strength of recommendation: strong).***

**5.2. What recipient factors influence the outcome of faecal microbiota transplant when treating people with *Clostridium difficile* infection?**

**5.2.1. General approach to co-morbidities and FMT:**

Most published studies had a core set of general recipient exclusions which included: significant/ anaphylactic food allergy<sup>14,17</sup>, pregnancy<sup>12–15,17,18</sup>, breastfeeding<sup>14</sup>, admission to Intensive Care or the



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requirement for vasopressors<sup>12,15,18</sup>, chronic diarrhoea or other infectious cause of diarrhoea<sup>12,14,18,50</sup>, inflammatory bowel disease (IBD) and irritable bowel syndrome (IBS)<sup>14,36</sup>, immunodeficiency due to recent chemotherapy and/ or neutropenia<sup>12,14–18,50</sup>, HIV/AIDS<sup>14,17,18</sup>, prolonged use of corticosteroids<sup>15,17,18</sup>, graft versus host disease<sup>12</sup>, and decompensated cirrhosis<sup>14,15,17,18</sup>.

The working group discussed the reported practice of several centres of treating patients with recurrent CDI and food allergies through the use of FMT prepared from a patient-directed donor instructed to avoid trigger foods before stool donation. They agreed that this seemed reasonable for patients with true adverse immunological reactions to defined food groups (e.g. gluten-free diet donor for a recipient with coeliac disease). However, the working group noted that food allergies are often poorly-defined clinically, and also expressed concerns that there was no means to verify how closely a donor had followed an exclusion diet; as such, they felt unable to make any specific recommendation about FMT in patients with food allergies in general. In contrast, whilst the working group were unaware of any reports in the literature of anaphylaxis attributable to FMT, they felt that the theoretical risk of a serious adverse outcome in patients with anaphylactic food allergy merited a specific recommendation that such individuals should not be offered FMT. Similarly, the working group expressed concern about the theoretical risk of adverse outcomes when administering FMT to patients with advanced decompensated chronic liver disease (including translocation of microbial material from the intestinal tract into the portal and systemic circulations, and theoretical risk of sepsis), and felt that FMT should be used with caution in this patient group.

#### **Recommendations:**

- i. We recommend that FMT should be avoided in those with anaphylactic food allergy (GRADE of evidence: very low; strength of recommendation: strong).**
- ii. We suggest that FMT should be offered with caution to patients with CDI and decompensated chronic liver disease (GRADE of evidence: very low; strength of recommendation: weak).**

#### **5.2.2. Immunosuppression and FMT:**

One randomised study<sup>16</sup> included patients with immunodeficiency (treatment with immunosuppressive therapy (azathioprine, ciclosporin, infliximab, methotrexate alone, or in combination with corticosteroids) ( $n=18$ ), renal transplant ( $n=5$ ), chronic haemodialysis ( $n=5$ ), solid organ tumours ( $n=3$ ) and haematological malignancy ( $n=4$ )) at the time of FMT. Clinical resolution



rates after up to two FMTs were high: 27/29 (93%) for immunocompromised individuals, 5/6 (83%) for patients with IBD.

There are also limited data from case series and single case reports describing the use of FMT in patients with immunocompromise. Agrawal and colleagues<sup>55</sup> included 46/146 (32%) patients with a history of cancer, and an additional 15/146 (10%) patients with non-cancer-related immunologic dysfunction, although primary outcome measures were not specifically reported for these groups. Overall cure at 12 weeks in a case series of 80 patients with immunocompromise was reported in 71 (89%) of patients<sup>60</sup>. Adverse events occurred in 12 (15%) immunocompromised patients; this included two deaths (one due to respiratory failure and another due to pneumonia resulting from aspiration at the time of FMT administration)<sup>60</sup>; however, such adverse events have also been reported in non-immunocompromised patient populations<sup>80</sup>. Hefazi and coauthors described high efficacy rates in a case series of FMT for recurrent CDI and a range of haematological or solid organ malignancies (remission after one FMT in 11/12 with haematological patients, and 8/10 in solid organ malignancy patients). No significant FMT-related complications were reported<sup>81</sup>. A further case series<sup>45</sup> reported FMT treatment for 75 patients with recurrent CDI and found no significant difference in primary cure rates for patients with diabetes mellitus, malignancy, or steroid use in the preceding three months.

The working group discussed the potential impact of donor EBV and CMV status for the immunocompromised FMT recipient at risk of severe infection if exposed to these viruses. Their opinion was that such recipients should only receive FMT from donors with negative EBV and CMV status.

**Recommendations:**

- i. ***We recommend that FMT should be offered with caution to immunosuppressed patients, in whom FMT appears efficacious without significant additional adverse effects (GRADE of evidence: moderate; strength of recommendation: strong).***
- ii. ***We recommend that immunocompromised FMT recipients at risk of severe infection if exposed to EBV or CMV should only receive FMT from donors negative for EBV and CMV (GRADE of evidence: very low; strength of recommendation: strong).***

**5.2.3. Other comorbidities and FMT:**

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Only a limited number of cited studies included specific detail about the presence of comorbidities in patients receiving FMT. However, several studies reported median Charlson comorbidity scores<sup>12,14,15,18,50</sup>. One randomised study reported the presence of IBD in 10/17 (59%) FMT recipients<sup>16</sup>, and there did not appear to be any significant difference in primary outcome measures in this group. Another randomised trial included 14/72 (33%) patients with IBD and reported clinical cure of CDI in 12/14 (86%) of these patients<sup>13</sup>. This study also included 64/72 (89%) patients with cardiac, respiratory, renal, central nervous system or multi-organ system comorbidities<sup>13</sup>; however outcomes were not stratified according to co-morbidity. Kelly and coauthors<sup>60</sup> reported an overall cure rate of 94% in a subset of CDI patients with IBD. A meta-analysis of studies in which patients with IBD received FMT (either primarily as treatment for concurrent recurrent CDI, or with the aim of treating IBD) noted a small risk of exacerbation of IBD in association with the use of FMT<sup>82</sup>. The working group noted the complexity of the relationship between IBD and CDI, given that IBD is itself a risk factor for CDI.

Other exclusions have been more directly related to the mode of administration. For upper gastrointestinal delivery, exclusion criteria have included delayed gastric emptying, chronic aspiration, 'swallow dysfunction', and dysphagia<sup>17,50</sup>. Exclusions for lower GI administration have included colostomy/ileostomy<sup>16,50</sup>, significant bleeding disorders<sup>12</sup>, untreated colorectal cancer<sup>14,36,54</sup>, and ileus/small bowel obstruction<sup>50</sup>.

In summary, the working group noted that co-morbidities amongst patients with recurrent CDI are common. Most studies did not analyse primary outcome measures according to co-morbidity; however, a small number of studies have analysed primary outcome measures (clinical cure) for patients with IBD receiving FMT for recurrent CDI and have found no significant difference compared to those without IBD, along with no overall significant worsening of IBD activity.

#### **Recommendations:**

- i. We recommend that FMT should be offered to those with recurrent CDI and inflammatory bowel disease, but patients should be counselled about a small but recognised risk of exacerbation of IBD (GRADE of evidence: moderate; strength of recommendation: strong).***

**ii. We recommend that FMT should be considered for appropriate patients with recurrent CDI regardless of other comorbidities (GRADE of evidence: moderate; strength of recommendation: strong).**

**5.3. What donor factors influence the outcome of faecal microbiota transplant when treating people with *Clostridium difficile* infection?**

**5.3.1. General approach to donor selection:**

Excellent efficacy has been shown in treating recurrent CDI using FMT derived from both related<sup>14,36,54,57,59,61,83,38,40,41,43,45,46,49,53</sup> and unrelated<sup>14,15,57,59,61,72,74,83–87,16,17,35,37,38,41,43,53</sup> donors. To date, there have been no randomised studies comparing differences in efficacy. Case series have tended to rely more on donation of stool from healthy family members. In randomised studies using FMT, all donors were healthy unrelated individuals<sup>12–18,88</sup>. Three case series used donor stool from healthcare professionals<sup>39,61,85</sup>; no randomised studies have used stool from this cohort. However, the working group noted that there were clear advantages to using FMT from a screened anonymous donor, in particular with regards to monitoring and traceability, as discussed further later.

**Recommendation:**

**We recommend that related or unrelated donors should both be considered acceptable. However, where possible, FMT is best sourced from a centralised stool bank, from a healthy unrelated donor (GRADE of evidence: low; strength of recommendation: strong).**

**5.3.2. Age and BMI restrictions for potential donors:**

There are no well-defined age restrictions on donors. Randomised studies have used donors of  $\geq 18^{12,72}$  and  $\leq 60$  years old<sup>15,17,18</sup> with satisfactory outcomes. Two of the case series defined age limitations for donors as  $\geq 18$  and  $\leq 50$  years<sup>72,89</sup>. A recent study demonstrated that *Bacteroides: Firmicutes* ratio and microbial diversity was similar for donors above and below 60 years, and their stool donations had similar clinical efficacy as FMT; however, there were loss of the phylum *Actinobacteria* and family *Bifidobacteriaceae* from donors older than 60 years<sup>90</sup>. On balance, the working group agreed that an age range of 18 – 60 years was appropriate for donors.

A widely-reported case study noted apparent weight gain in a recipient of FMT for treatment of CDI when an overweight donor was used<sup>91</sup>, but any association between a donor with a raised BMI and weight gain post-FMT has not been replicated elsewhere in the literature<sup>92</sup>. Whereas most randomised studies did not report donor-specific BMIs, some have excluded those without a 'normal' BMI<sup>13,17</sup>. The working group considered an acceptable BMI for donors as between  $\geq 18$  to  $\leq 30$  kg/m<sup>2</sup>.

**Recommendation:**

***We suggest that people should only be considered as potential FMT donors if they are  $\geq 18$  and  $\leq 60$  years old, and have a BMI of  $\geq 18$  and  $\leq 30$  kg/m<sup>2</sup> (GRADE of evidence: low; strength of recommendation: weak).***

**5.3.3. General approach to the donor screening assessment:**

There is a clear theoretical risk of the transmission of infection by FMT; furthermore, given the large number of conditions in which perturbation of the gut microbiota has been described<sup>93</sup>, there is a concern regarding a risk of transmission of microbiota associated with vulnerability to disease. Whilst FMT is efficacious for recurrent CDI, adverse events may be associated with its use (discussed further later), and long-term safety follow-up is lacking. The aim of a donor screening questionnaire and interview is to minimise post-FMT adverse events by excluding potential donors from whom FMT may be associated with risk to recipients. Randomised studies performed to date used various pre-screening questionnaires, including self-screening questionnaires which focused on high risk behaviours for blood-borne infections<sup>12-16</sup>, questionnaires that focused on previous potential transferable medical conditions<sup>18</sup>, and adaptations from the American Association of Blood Banks Donor Questionnaire<sup>14,17</sup>. One randomised study used the OpenBiome questionnaire as a screening questionnaire<sup>94</sup>. Some studies have suggested excluding potential donors who have recently travelled to defined regions (typically tropical areas), varying between 3-6 months prior to donation<sup>38,39,49,52,55,59,74,87</sup>; this is also the protocol employed in randomised studies<sup>14,16,18</sup>. Another important point for assessment is recent use of medications by potential donors. In particular, given the profound effects of antimicrobials on the gut microbiota<sup>95-98</sup> (along with the theoretical concern that recent antimicrobials might precipitate gut colonisation with antimicrobial-resistant bacteria that could be transferred during FMT), studies advocate either a three month<sup>14,46,53-55,57,61,74</sup> or six month<sup>16-18,35,38,39,43,49,85,99,100</sup> period without antimicrobial use prior to FMT donation.

986

987 The working group agreed that, given the growing evidence for the contribution of the gut  
988 microbiota to the aetiopathogenesis of colorectal carcinoma, patients with a significant personal or  
989 family history of (or risk factors for) this condition should be excluded as donors (**Table 1**). However,  
990 the working group noted an added complexity, in that their recommendation was that potential  
991 donors may be up to 60 years of age, but bowel scope screening for colorectal carcinoma currently  
992 begins within the UK at 55 years of age, and formal NHS bowel cancer screening starts at the age of  
993 60 years<sup>101</sup>. The working group agreed that potential donors living in countries with bowel cancer  
994 screening programmes that start before the age of 60 years should have therefore completed  
995 appropriate screening with negative/ normal tests before they are considered further as donors.

996

997 The working group was of the opinion that a screening process is mandatory; any positive responses  
998 should usually result in exclusion from donation, although this will depend upon the particular  
999 circumstances/ answers given. A donor screening questionnaire should be performed both prior to  
1000 considering a person as a donor, and also at a further point in time (discussed further in **Section**  
1001 **5.3.5**).

1002

1003 **Recommendation:**

1004 ***It is mandatory to screen potential donors by questionnaire and personal interview, to***  
1005 ***establish risk factors for transmissible diseases and factors influencing the gut microbiota***  
1006 ***(Table 1) (GRADE of evidence: low; strength of recommendation: strong).***

1007

1008 **5.3.4. Laboratory screening of potential donors:**

1009 Currently, there are no known confirmed cases of blood-borne pathogens being transmitted by FMT,  
1010 but strict preventative measures are important, as the potential risk of transmission is unknown.  
1011 Many of the suggestions are extended from established blood screening guidelines<sup>102</sup>. Case series  
1012 almost universally screen for HIV, hepatitis B and hepatitis C as a minimum<sup>35,36,52–</sup>  
1013 <sup>55,59,61,72,74,84,86,37,87,103,39–43,46,49</sup>; other studies (including the randomised trials) have a more thorough  
1014 blood screening process<sup>14–18</sup>. Many studies have also included a ‘metabolic/general blood screen’, to  
1015 select out donors with hitherto undiagnosed chronic illness. **Table 2** shows the suggested blood  
1016 screening protocol of the BSG/HIS working group.

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1017

1018 The working group specifically discussed the role of screening donors for their EBV and CMV status;  
1019 the importance of the rationale for this is discussed in **Section 5.2.2**. They agreed that EBV and CMV  
1020 testing was only required where there is the potential that the FMT prepared from that donor would  
1021 be administered to immunosuppressed patients at risk of severe infection if exposed to CMV and  
1022 EBV.

1023

1024 The primary aim of stool screening of potential donors is to minimise the risk of transmission of  
1025 pathogens; again, the relative novelty of FMT for CDI means that these risks are not currently well-  
1026 defined. Stool screening protocols are universal amongst published studies, though widely-variable  
1027 protocols have been used. **Table 3** displays the suggested stool screening protocol of the working  
1028 group. The working group discussed stool screening for multi-drug resistant bacteria carriage, and  
1029 agreed that carbapenemase-producing *Enterobacteriaceae* (CPE) should be screened for. Although  
1030 these bacteria are carried only by a minority of the UK population, transfer into debilitated patients  
1031 with CDI is clearly undesirable given that CPE are potentially so difficult to treat. They also agreed  
1032 that extended-spectrum beta-lactamase (ESBL)-producing organisms could also potentially cause  
1033 severe disease (with limited antimicrobial options) if transplanted into patients with CDI, and so  
1034 should also be screened for. Whilst vancomycin-resistant *Enterococci* (VRE) carriage is relatively  
1035 common in the community (probably related to food consumption)<sup>104</sup>, community strains of VRE are  
1036 genetically distinct from (and generally of much lower pathogenicity than) those found  
1037 nosocomially<sup>105</sup>; as such, the working group thought that routine screening was not justified. The  
1038 working group also noted that methicillin-resistant *Staphylococcus aureus* (MRSA) carriage is very  
1039 rare in healthy adults in non-healthcare settings (with significant intestinal carriage even rarer), so  
1040 did not justify routine screening. However, the working group acknowledged that the potential  
1041 infection risk from VRE and MRSA would vary regionally dependent upon local prevalence and  
1042 pathogenicity, and as such recommended that a risk assessment is performed to assess whether  
1043 screening for these organisms should be considered.

1044

1045 A donor laboratory screening should be performed both prior to considering a person as a donor,  
1046 and also at a further point in time (discussed further in **Section 5.3.5**).

1047

1048 **Recommendation:**

**Blood and stool screening of donors is mandatory (Tables 2 and 3) (GRADE of evidence: low; strength of recommendation: strong).**

**5.3.5. Repeat donor checks, and donation pathway:**

Almost all reviewed studies have repeated at least some elements of the initial donor screening process either at the time of donation of each stool sample used to prepare FMT, or at the end of a period of donation to assess ongoing suitability for inclusion. However, protocols have differed widely between studies.

The opinion of the working group was that when a donor had met criteria for donation (both with an acceptable health questionnaire and satisfactory laboratory tests), they were suitable to begin donation of stool that may be prepared into FMT. Repeat donor screening was also deemed necessary. In centres where frozen FMT is being prepared, stool may be collected and processed immediately after the first donor screen is successfully completed, but should be stored in ‘quarantine’ pending further donor screening, rather than used immediately for clinical use. At the end of the locally-defined period of donation, potential donors should undergo repeat testing, with a further health questionnaire and laboratory screening. If the donor’s health questionnaire remains acceptable and repeat laboratory screening is negative at this point, then the frozen FMT may be released from ‘quarantine’, and used. The working group thought that donor screening both before and after donation was the safest route possible, and that this represented the preferred scenario. A proposed summary pathway for donor screening in this scenario is provided in **Figure 1**.

In centres using fresh FMT, the working group agreed that a repeat health questionnaire should be completed at the time of donation of each stool sample used to prepare FMT. Formal repetition of both the personal interview/ health questionnaire and laboratory screening tests should occur at regular intervals to ensure ongoing suitability for inclusion as a donor. The working group’s opinion was that this repetition of the screening process should occur at least once every four months.

**Recommendations:**

- i. In centres using frozen FMT, before FMT may be used clinically, we recommend that donors should have successfully completed a donor health questionnaire and***



laboratory screening assays both before and after the period of stool donation. This is the preferred means of donor screening (GRADE of evidence: low; strength of recommendation: strong).

ii. In centres using fresh FMT, we recommend that a repeat health questionnaire should be assessed at the time of each stool donation. To ensure ongoing suitability for inclusion as a donor, the donor health questionnaire and laboratory screening should be repeated regularly (GRADE of evidence: low; strength of recommendation: strong).

#### **5.4. What factors related to the preparation of the transplant influence the outcome of faecal microbiota transplant when treating people with *Clostridium difficile* infection?**

##### **5.4.1. General principles of FMT preparation:**

There is very little evidence or guidance on the collection of donor stool. Critical steps during this process centre on the reduction of environmental cross-contamination risk, so the use of clean collection devices and clean collection procedures is advocated. To promote standardised practice and a safe and effective product, clear instructions should be provided to the donor for stool collection (Table 5).

Regardless of the methods used to prepare FMT, stool donations should be processed within six hours of defaecation. The period of six hours has been generally applied across many successful studies of FMT treatment in CDI<sup>14,18,35,39,43,52</sup>, although no formal comparative study has been undertaken. This strategy aims to minimise sample degradation and alteration over time, which may occur due to the complex metabolic and environmental requirements of the faecal microbiota.

There are no comparative trials of anaerobically versus aerobically prepared FMT in the treatment of recurrent CDI. With the exception of small observational studies<sup>41,74</sup>, the vast majority of FMT preparation has been undertaken aerobically for the treatment of CDI and has proved highly efficacious. There appears to be no clear need to process anaerobically, a method which introduces complexity and cost for the treatment of CDI.



1110 The reviewed randomised studies reported variable amounts of stool used in the preparation of  
1111 each FMT aliquot, and the lack of comparative data means that it is not possible to link stool mass to  
1112 outcome from these studies. However, a previous systematic review of case series using FMT as  
1113 treatment for recurrent CDI reported similar rates of treatment efficacy, but an approximate  
1114 fourfold increase in recurrence rates, if <50g of stool was used compared to ≥50g<sup>106</sup>. Similarly, the  
1115 initial volume of diluent used to create the faecal emulsion is variable between studies, although the  
1116 most common practice appears to be creation of a stool: diluent ratio of approximately 1:5. The  
1117 overwhelming majority of the reviewed studies used stool from only a single donor per FMT (rather  
1118 than stool pooled from a mixture of donors), and there are no comparative studies of outcomes of  
1119 CDI from single donor vs pooled donor FMT; as such, the working group found no justification to  
1120 recommend donor stool pooling for FMT for CDI.

1122 The majority of studies have used preservative-free sterile 0.9% saline as the diluent for FMT  
1123 production, although there have been a handful of reports of other diluents including potable  
1124 water<sup>16,35,43</sup>. There have been no comparative studies of FMT diluent. In cases where frozen FMT is  
1125 prepared, an appropriate cryoprotective substance should be added prior to freezing. Most studies  
1126 use glycerol at a final concentration of ~10%<sup>16,41</sup>. It has been demonstrated that storing stool at -  
1127 80°C for up to six months in saline without glycerol decreases viable aerobic and anaerobic bacterial  
1128 counts; the reduction was statistically significant in all bacterial groups with the exception of *E. coli*  
1129 and total anaerobes. When stored with glycerol, no significant reduction in viable counts was  
1130 observed<sup>74</sup>.

1132 A variety of homogenisation and open filtration systems have been used, with no apparent major  
1133 variation in efficacy. Open filtration systems such as gauze<sup>16,37,40,55</sup>, filter paper<sup>39</sup> and strainers/  
1134 sieves<sup>17,41</sup> are unpleasant to use and pose a risk of external contamination. In order to best comply  
1135 with GMP standards, a sterile, single-use closed homogenisation and filtration system is  
1136 recommended. An example of such a system includes the use of sterile filter bags inside a  
1137 laboratory paddle homogeniser.

1139 **Recommendations:**

- 1140 *i. We recommend that donor stool collection should follow a standard protocol*  
1141 *(GRADE of evidence: low; strength of recommendation: strong).*

- 1142 **ii. We recommend that donor stool should be processed within 6 hours of defaecation**  
 1143 **(GRADE of evidence: low; strength of recommendation: strong).**
- 1144 **iii. We recommend that both aerobically and anaerobically prepared FMT treatments**  
 1145 **should be considered suitable when preparing FMT for the treatment of recurrent**  
 1146 **CDI (GRADE of evidence: moderate; strength of recommendation: strong).**
- 1147 **iv. We recommend that sterile 0.9% saline should be considered as an appropriate**  
 1148 **diluent for FMT production, and cryoprotectant such as glycerol should be added**  
 1149 **for frozen FMT (GRADE of evidence: moderate: strength of recommendation:**  
 1150 **strong).**
- 1151 **v. We recommend using  $\geq 50\text{g}$  of stool in each FMT preparation (GRADE of evidence:**  
 1152 **moderate: strength of recommendation: strong).**
- 1153 **vi. We suggest that stool should be mixed 1:5 with diluent to make the initial faecal**  
 1154 **emulsion (GRADE of evidence: low; strength of recommendation: weak).**
- 1155 **vii. We suggest that homogenisation and filtration of FMT should be undertaken in a**  
 1156 **closed disposable system (GRADE of evidence: low; strength of recommendation:**  
 1157 **weak).**

#### 1159 **5.4.2. Fresh vs frozen FMT:**

1160 Two randomised studies have examined this area. One double-blind randomised study concluded  
 1161 that enema frozen FMT ( $n=91$ ) was non-inferior for clinical resolution of diarrhoea to fresh FMT  
 1162 ( $n=87$ ) for the treatment of recurrent or refractory CDI<sup>16</sup> (with frozen FMT in this study stored at -  
 1163 20°C for up to 30 days). A further randomised study demonstrated statistically comparable  
 1164 remission rates for recurrent CDI with fresh or frozen FMT delivered colonoscopically ( $n=25/25$  vs  
 1165 20/24 respectively,  $p=0.233$ ) (using frozen FMT stored at -80°C for up to six months)<sup>13</sup>. These data  
 1166 support the findings of earlier small observational studies<sup>35,41</sup>. Frozen FMT is preferable to fresh FMT  
 1167 on logistical and cost grounds<sup>16</sup>. Banked frozen FMT also enables the window period for donor  
 1168 screening to be minimised, allowing centres to more closely to meet regulatory requirements (also  
 1169 see **Section 5.3.5**).

#### 1171 **Recommendation:**

***We recommend that the use of banked frozen FMT material should be considered preferable to fresh preparations for CDI (GRADE of evidence: high; strength of recommendation: strong).***

**5.4.3. Use of frozen FMT:**

Frozen FMT has been used up to six months after storage at -80°C<sup>17,41,74</sup>, with high efficacy rates (>70%) observed in the cases treated. However, there have been no comparative trials investigating storage durations. A trend towards decrease in the viability of certain gut microbiota taxa was noted when faecal aliquots were frozen in 10% glycerol for six months<sup>74</sup>, and as such, the working group agreed that six months was the acceptable limit for freezing of an FMT in glycerol. Storage at -80°C is recommended rather than -20°C to minimise sample degradation.

Warm water baths have been recommended to speed thawing<sup>6</sup>; however, the working group thought that this should be strongly discouraged, as this may introduce risks of cross contamination by *Pseudomonas* species (and other contaminants) from the water bath<sup>107,108</sup>, and may reduce bacterial viability in the FMT. Repetitive freeze thawing of FMT samples should be avoided as bacterial numbers will be reduced during this process<sup>109</sup>.

***Recommendations:***

- i. We recommend that FMT material stored frozen at -80°C should be regarded as having a maximum shelf life of six months from preparation (GRADE of evidence: low; strength of recommendation: strong).***
- ii. We suggest consideration of thawing frozen FMT at ambient temperature, and using within six hours of thawing (GRADE of evidence: low; strength of recommendation: weak).***
- iii. We suggest not thawing FMT in warm water baths, due to the risks of cross contamination with Pseudomonas (and other contaminants) and reduced bacterial viability (GRADE of evidence: very low; strength of recommendation: weak).***

**5.5. What factors related to administration of the transplant influence the outcome of faecal microbiota transplant when treating people with *Clostridium difficile* infection?**

**5.5.1. Use of specific medications in the period around FMT administration:**

**5.5.1.1. General principles of FMT administration:**

Bowel purgatives have been proposed pre-FMT as a means of removing residual antibiotics that may affect engraftment of transplanted microorganisms, and as a means of removing any residual *C. difficile* toxin, spores and vegetative cells<sup>110–114</sup>. Furthermore, bowel purgatives pre-colonoscopy FMT delivery facilitate safe endoscopy. Various bowel purgatives have been used in colonoscopic FMT studies, including polyethylene glycol (PEG) (often 4 litres)<sup>14,17,115–117,35,41,43,46,54–56,100</sup>, Moviprep<sup>®35,41</sup>, and macrogol<sup>13,15,18,59</sup>. In those studies that used an upper GI route for FMT, PEG<sup>54,55,84</sup> and Klean-Prep<sup>®15,61</sup> were used. FMT without bowel preparation has also been used as treatment for recurrent CDI without any apparent reduction in efficacy, including in randomised studies<sup>16</sup>.

The rationale for the use of proton pump inhibitors (PPI) prior to upper GI FMT is to minimise acidity which may impair engraftment of transplanted microorganisms; however, PPIs have been shown to alter the gut microbiota<sup>118,119</sup>, and have also been associated with primary and recurrent CDI<sup>120,121</sup>. Some studies advocate the use of PPI prior to receiving FMT via the upper GI route<sup>37,39,45,84,85,122,123</sup>, but there appears to be comparable efficacy data in studies where it has not been used. Certain studies have also given recipients PPI prior to receiving colonoscopic FMT<sup>17,87</sup>.

The use of prokinetics (such as metoclopramide) has been described prior to FMT delivery via the upper GI tract route, but only in a very small number of studies<sup>85</sup>. Given the potential risk of regurgitation/aspiration associated with upper GI administration of FMT, the working group felt that its use should be considered where appropriate.

A single dose/ short course of loperamide has been used following FMT (predominantly for lower GI administration) in an attempt to prolong the exposure of the FMT to the mucosa, and to aid retention of the FMT within the GI tract<sup>13,46,49,55,84,123</sup>. One study utilised diphenoxylate with atropine<sup>54</sup> instead. However, no studies have compared FMT with and without anti-motility drugs.

The working group also discussed infection control aspects as they apply to FMT administration. Specifically, they agreed that recipients should ideally be cared for in a single room with en-suite bathroom facilities and, where appropriate, be placed at the end of an endoscopy list, to facilitate enhanced environmental decontamination and prevention of transmission of *C. difficile* spores. Protocols for decontamination of endoscopes should follow national guidance<sup>124,125</sup>, using a sporicidal agent. Best practice for prevention of transmission of healthcare-associated infections, as described in national guidelines<sup>126</sup>, should also be applied throughout.

**Recommendations:**

- i. We recommend that bowel lavage should be administered prior to FMT via the lower GI route, and bowel lavage should be considered prior to FMT via the upper GI route; polyethylene glycol preparation is preferred (GRADE of evidence: low; strength of recommendation: strong).*
- ii. For upper GI FMT administration, we suggest that a proton pump inhibitor should be considered, e.g. the evening before and morning of delivery (GRADE of evidence: low; strength of recommendation: weak).*
- iii. We suggest that a single dose of loperamide (or other anti-motility drugs) should be considered following lower GI FMT delivery (GRADE of evidence: low; strength of recommendation: weak).*
- iv. We suggest that prokinetics (such as metoclopramide) should be considered prior to FMT via the upper GI route (GRADE of evidence: low; strength of recommendation: weak).*
- v. We recommend that best practice for prevention of further transmission of CDI should be applied throughout when administering FMT to patients with CDI (nursing with enteric precautions, sporicidal treatment of endoscope, etc) (GRADE of evidence: high; strength of recommendation: strong).*

**5.5.1.2. Additional antibiotics pre-FMT:**

Many studies have given further courses of conventional antimicrobial *C. difficile* treatment prior to FMT. Regimens have included vancomycin alone<sup>12,14,18,35,39,55,59,86,117</sup>, metronidazole or

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vancomycin<sup>40,41,43,122</sup>, or alternatively vancomycin, fidaxomicin or metronidazole<sup>56</sup>, with one study using a range of regimens which included rifaximin<sup>123</sup>. The length of treatment was also variable, ranging from 24 hours<sup>54</sup> up to four days prior to receiving FMT<sup>39,45</sup>; however, comparative studies have not been undertaken.

**Recommendation:**

***We recommend the administration of further antimicrobial treatment for CDI for at least 72 hours prior to FMT (GRADE of evidence: low; strength of recommendation: strong).***

**5.5.1.3. Washout period between antibiotic use and FMT:**

Nearly all studies specified a washout period after completing anti-CDI antibiotics and before administration of FMT. However, this time period appeared to be arbitrarily selected and varied from as little as four<sup>46</sup> or 12 hours<sup>51</sup>, up to 72 hours<sup>36</sup>. The majority of studies specified either 24 hours<sup>15,37,39,40,45,54,127</sup> or 48 hours<sup>41,42,49,60</sup>, however some allowed a range from 1-3 days<sup>16,44,52,53,55</sup>. One study appeared to allow co-administration of vancomycin with bowel preparation, without a washout period<sup>18</sup>.

The working group discussed the challenging scenario of providing FMT to patients with recurrent CDI, but who also had a strong indication for long-term non-anti-CDI antibiotics (e.g. splenectomy, osteomyelitis, or infective endocarditis), or patients who develop an indication for antibiotics for a reason other than CDI shortly after receiving FMT. The concern in this instance is that the use of antibiotics may limit engraftment of microbial communities derived from the FMT, and therefore reduce its effectiveness. The working group discussed a recent retrospective study demonstrating that exposure to non-anti-CDI antimicrobials within eight weeks of FMT is associated with an approximate threefold risk of FMT failure ( $n=8/29$  failures with antibiotic exposure vs  $36/320$  failures without antibiotic exposure)<sup>128</sup>. Similarly, the experience of the large pan-Netherlands stool bank<sup>129</sup> was that ~50% of their failures of FMT in the treatment of recurrent CDI occurred in patients who had received antibiotics within one month of their FMT. For patients requiring long-term antibiotics, the working group's expert opinion was that such patients should still be eligible for FMT, but that the regimen for the use of non-anti-CDI antibiotics should be decided on a case-by-case basis, based on factors including response to FMT and/or strength of indication of antibiotics. Both in this scenario, and the scenario in which antibiotics are required shortly after receiving FMT, the working

1295 party agreed that infectious diseases specialists/medical microbiologists should be involved in  
1296 making decisions regarding the choice of agents used.

1298 **Recommendations:**

- 1299 *iii. To minimise any deleterious effect of antimicrobials on the FMT material, we*  
1300 *recommend that there should be a minimum washout period of 24 hours between the*  
1301 *last dose of antibiotic and treatment with FMT (GRADE of evidence: low; strength of*  
1302 *recommendation: strong).*
- 1303 *iv. We suggest considering consultation with infectious disease specialists or medical*  
1304 *microbiologists for advice whenever FMT recipients also have an indication for long-*  
1305 *term antibiotics, or have an indication for non-CDI antibiotics within eight weeks of*  
1306 *FMT (GRADE of evidence: very low; strength of recommendation: weak).*

1308 **5.5.2. Route of FMT delivery:**

1309 **5.5.2.1. Introduction:**

1310 FMT can be delivered via the lower GI route (retention enema, colonoscopy), upper GI route  
1311 (endoscopically, or via nasogastric tube, nasoduodenal or nasojejunal tube), or via capsules  
1312 (containing either frozen FMT or lyophilised faecal material). Systematic reviews with meta-analysis  
1313 suggest that FMT for recurrent CDI via colonoscopy may have slightly higher efficacy compared to  
1314 upper GI administration<sup>127,130–132</sup> with similar safety profiles, but also note the trend towards using  
1315 larger amounts of stool or ‘higher concentration’ FMT in lower GI administration. One systematic  
1316 review (reviewing principally case series, and including only one randomised study) compared  
1317 remission rates for CDI using FMT delivered to different areas of the GI tract, and reported that for  
1318 FMT infused into the stomach, duodenum/jejunum, caecum/ascending colon, and rectum the rates  
1319 of cure rate were 81%, 86%, 93%, and 84%, respectively<sup>131</sup>.

1321 In the only randomised study that directly compared upper and lower GI administration, there was  
1322 no significant difference in overall cure rate ( $p = 0.53$ )<sup>17</sup>.

1324 **5.5.2.2. Upper gastrointestinal tract administration of FMT:**



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FMT has been shown to be safe and efficacious in the treatment of *C. difficile* when administered via nasogastric tube<sup>37,39,45,61,83,123</sup>, nasoduodenal tube<sup>15,84,85</sup>, enteroscopy<sup>122,123</sup>, or via the infusion channel on a gastroscope<sup>40,45</sup>. In a randomised trial, nasoduodenal donor FMT has been shown to be more efficacious than vancomycin in treating recurrent CDI<sup>15</sup>. Furthermore, it has been shown that FMT can also be safely and effectively delivered via a percutaneous endoscopic gastrectomy tube<sup>45,83</sup>. The working group noted that upper GI administration of FMT may be particularly suitable for certain patient groups, such as those in whom there are contraindications or who would find it difficult to tolerate lower GI endoscopy, and/ or patients unlikely to be unable to retain enemas.

1333

Typically, smaller volumes of faecal suspension are administered to the upper GI tract compared to lower GI administration, with quoted volumes ranging from 25ml<sup>39</sup> up to 150ml<sup>84</sup>- 250ml<sup>37,85</sup>. Up to 500ml of suspension has been given safely and effectively via the upper GI route<sup>15,77</sup>. However, the working group expressed concerns regarding the risk of regurgitation and aspiration if large volumes of FMT are administered to the upper GI tract, and also discussed cases in which this has been described with adverse outcomes<sup>80</sup>. This included a reported death from aspiration, after 100-150ml of FMT was delivered by enteroscope into the distal duodenum under general anaesthetic as attempted treatment for recurrent CDI<sup>133</sup>. A further reported case described a case of fatal aspiration pneumonitis likely related to a 500ml FMT via nasoduodenal tube; this patient had a swallowing disorder following oropharyngeal radiation after surgical removal of a maxillary carcinoma two years previously<sup>77</sup>. Based on their expert opinion, the working group recommended that upper GI FMT should be used with caution in those at risk of regurgitation (e.g. known large hiatus hernia, severe gastro-oesophageal reflux disease, etc) and/ or with swallowing disorders (although administration via a gastrostomy tube would be acceptable). They also recommended that no more than 100ml of FMT should be administered to the upper GI tract to minimise these risks.

1350

#### 1351 **Recommendations:**

- 1352 **i. We recommend that upper GI administration of FMT as treatment for recurrent or**
- 1353 **refractory CDI should be used where clinically appropriate (GRADE of evidence:**
- 1354 **high; strength of recommendation: strong).**
- 1355 **ii. Where upper GI administration is considered most appropriate, we recommend**
- 1356 **that FMT administration should be via nasogastric, nasoduodenal, or nasojejunal**

- 1357 ***tube, or alternatively via upper GI endoscopy. Administration via a permanent***  
1358 ***feeding tube is also appropriate (GRADE of evidence: high; strength of***  
1359 ***recommendation: strong).***  
1360 ***v. We recommend that no more than 100ml of FMT is administered to the upper GI***  
1361 ***tract (GRADE of evidence: low; strength of recommendation: strong).***  
1362 ***vi. We recommend that upper GI administration of FMT should be used with caution***  
1363 ***in those at risk of regurgitation and/ or those with swallowing disorders (GRADE of***  
1364 ***evidence: low; strength of recommendation: strong).***  
1365

1366 **5.5.2.3. Lower gastrointestinal tract administration of FMT:**

1367 **FMT via enema:** Successful treatment of *C. difficile* with FMT enema has been  
1368 demonstrated<sup>16,38,42,53,55,83,86</sup> but enema appears to have a lower efficacy than other routes of FMT  
1369 administration. Specifically, in a randomised study primarily comparing the efficacy of fresh and  
1370 frozen FMT in the treatment of recurrent CDI, only 52.8% of patients in the ‘frozen’ arm and 50.5%  
1371 of patients in the ‘fresh’ arm of the study ( $n=57/108$  and  $56/111$  respectively) experienced  
1372 resolution of symptoms after a single enema, by modified intention to treat analysis<sup>16</sup>. However,  
1373 resolution rates in both arms only reached >80% after at least three enemas<sup>16</sup>. A recent randomised  
1374 study demonstrated similar rates of recurrence of CDI in patients with recurrent CDI treated with  
1375 either a single FMT enema or a six week vancomycin taper ( $n=9/16$  patients with recurrence vs  $5/12$   
1376 respectively)<sup>12</sup>. Notwithstanding this, enemas do have specific advantages, such as being a  
1377 treatment option where full colonoscopy is contraindicated. It is also possible to give multiple  
1378 infusions relatively easily and outside a hospital setting.

1380 **FMT via colonoscopy:** Randomised study evidence has demonstrated that colonoscopic FMT has  
1381 higher efficacy in treating recurrent CDI than vancomycin<sup>18</sup>. Efficacy is similar whether FMT is fresh  
1382 or frozen, but modestly reduced when using a lyophilised FMT product<sup>13</sup>. Colonoscopic delivery of  
1383 donor FMT into the ileum or caecum was associated with a 91% cure rate for recurrent CDI<sup>14</sup>.  
1384 Observational studies highlighted similar success, describing cure rates of 88% ( $n=14/16$ )<sup>74</sup> and 91%<sup>46</sup>  
1385 ( $n=21/23$ ) in response to infusion of donor FMT into the caecum or terminal ileum. A further  
1386 advantage of using colonoscopy to administer FMT has been to allow assessment for the presence of  
1387 pseudomembranes; in certain reviewed studies, the presence or absence of pseudomembranes has  
1388 influenced the FMT regimen used<sup>18,73</sup>. However, the working group noted that that many patients

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with CDI are frail and elderly, and as such it will not always be safe or feasible to undertake colonoscopy in this particular group of patients. Flexible sigmoidoscopy appears to be an feasible option where full colonoscopy cannot be performed e.g. unable to tolerate colonoscopy, severity of colitis<sup>56,60</sup>.

The amount of faecal suspension via enema has varied between 150-500mls<sup>16,38,42,55,86</sup>. The amount of faecal suspension delivered via colonoscopy has been similarly variable, with some studies suggesting as little as 100ml can be used with success rates of 94%<sup>43</sup>. 250ml-400ml had a success rate of 100%<sup>36</sup>, whereas infusions of up to 500-700ml were associated with cure rates of 92%<sup>46</sup>. However, the working group noted that it is difficult to compare 'concentration' of FMT in different studies as different protocols used varied starting amounts of faecal material. Currently, there are no randomised studies that compare concentration/ volume of colonoscopic or enema FMT. As such, no recommendation was made to this regard.

#### **Recommendations:**

- i. We recommend that colonoscopic administration of FMT as treatment for recurrent or refractory CDI should be used where appropriate (GRADE of evidence: high; strength of recommendation: strong).*
- ii. Where colonoscopic administration is used, we suggest considering preferential delivery to the caecum or terminal ileum, as this appears to give the highest efficacy rate (GRADE of evidence: low; strength of recommendation: weak).*
- iii. We recommend that FMT via enema should be used as a lower GI option when delivery using colonoscopy or flexible sigmoidoscopy is not possible (GRADE of evidence: high; strength of recommendation: strong).*

#### **5.5.2.4. Capsulised FMT:**

Capsulised FMT aims to remove some of the concerns regarding conventional FMT, such as the invasive means of administration and palatability. The largest case series describing the use of capsules as treatment for recurrent CDI<sup>72,89</sup> noted clinical resolution at eight weeks off antibiotics for CDI in 82% of cases ( $n=147/180$ ) after one course of capsules, and 91% ( $n=164/180$ ) after two courses. The capsules contained frozen FMT prepared in a diluent of saline with 10% glycerol; 15

capsules were administered each day for two consecutive days (equating to a mean 48g of original crude stool). Other smaller case series have demonstrated comparable results<sup>87,123,134</sup>, including when lyophilised stool is used instead of frozen whole FMT<sup>134</sup>.

The working group reviewed two randomised studies which have examined the efficacy of capsulised FMT in treating recurrent CDI. In one study, published in abstract form<sup>94</sup>, a 'high dose' regimen of frozen FMT capsules (30 capsules each day for two days) was compared to 'low dose' (30 capsules in one day). CDI resolution was comparably high in both arms with one treatment course (77% (n=7/9) in the 'high dose' arm vs 70% (n=7/10) in the 'low dose arm'). 4/5 initial non-responders entered remission after a second capsule course with the 'high dose' regimen<sup>94</sup>. In a recent large randomised trial, patients with recurrent CDI were randomised to receive either thawed frozen FMT either via colonoscopy or via capsules (one treatment of 40 capsules)<sup>11</sup>. On per protocol analysis, remission at 12 weeks after a single treatment occurred in 96% in both arms (n=51/53 by capsule, n=50/52 by colonoscopy).

The working group discussed certain unresolved issues regarding capsules. Specifically, capsules are often large, and swallowing 30 capsules in a single day may be a significant undertaking for patients with CDI, such as the frail elderly with an existing high pill burden. They also noted that follow-up data post-capsule administration is relatively short compared to other modalities of FMT.

**Recommendation:**

***Capsulised FMT holds promise as a treatment option for recurrent CDI and we recommend that this should be offered to patients as a potential treatment modality where available. Capsule preparations should follow a standard protocol. Further evidence regarding optimal dosing and formulation is required (GRADE of evidence: high; strength of recommendation: strong).***

**5.6. What is the clinical effectiveness of FMT in treating conditions other than *Clostridium difficile* infection?**

**5.6.1. Introduction:**

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In current clinical practice, FMT is used predominantly in the treatment of recurrent CDI. Its success has led to exploration of its efficacy in other GI diseases, primarily ulcerative colitis (UC), where perturbation of the gut microbiota has been observed and implicated in disease pathogenesis<sup>135</sup>. Due to variability of the quality, methodology and cohorts of patients recruited in trials of FMT for non-CDI indications, and in order to control for significant confounding factors, the working group only included randomised trials involving patients with well-defined conditions and in which there was a primary clinical outcome. To date, there have been a total of 71 such studies investigating the role of FMT in IBD; of these, only four are prospective randomised controlled trials, limited to patients with ulcerative colitis<sup>136–139</sup>. Five other reviewed randomised studies investigated the use of FMT in irritable bowel syndrome<sup>140</sup>, slow transit constipation<sup>141</sup>, hepatic encephalopathy<sup>142</sup> and metabolic syndrome<sup>143,144</sup>.

## **5.6.2. Use of FMT for ulcerative colitis:**

### **5.6.2.1. Efficacy:**

All four RCTs, with a total of 277 subjects, included patients with mild to moderate UC (Mayo score 3-11 and endoscopic sub-score of at least 1). Participants were aged between 27 and 56 years and largely included patients on stable immunosuppressive therapy (only one study excluded patients using biologic treatments and methotrexate within the preceding two months)<sup>136</sup>. Three studies included patients on oral corticosteroids at the time of FMT, however only two required a mandatory wean of these to meet eligibility. Studies generally included patients with all disease distributions found in UC. Time to evaluation of response to FMT in these studies varied between seven and twelve weeks. Two studies used autologous FMT as placebo<sup>136,139</sup>. Three of the four studies demonstrated that patients receiving donor FMT were significantly more likely to achieve clinical and endoscopic remission compared to placebo<sup>137–139</sup>. The pooled rate of combined clinical and endoscopic remission was 27.9% for donor FMT and 9.5% for placebo. A pooled risk ratio for failure of FMT to achieve these combined outcomes was 0.8 (95% CI: 0.7-0.9). Deep remission (histological) was only reported in one RCT: 18.4% of patients receiving FMT achieved this outcome compared to 2.7% of those receiving placebo<sup>137</sup>.

### **5.6.2.2. Characteristics of FMT preparation and delivery:**

The four RCTs varied in their FMT preparation and delivery methodology. Two RCTs delivered frozen FMT, one fresh FMT, and one used a combination. Three RCTs with a positive outcome delivered the FMT via the lower GI route; these studies used a high intensity protocol ranging from a total of three

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1483 infusions in one week to 40 FMTs over an eight week period<sup>137–139</sup>. The other RCT (that failed to  
1484 show efficacy of FMT for UC) had adopted a low intensity protocol of two nasoduodenal infusions  
1485 given three weeks apart<sup>136</sup>. Interestingly, the only RCT that prepared stool in anaerobic conditions  
1486 demonstrated the highest rate of steroid-free clinical remission and steroid-free clinical response  
1487 with donor FMT<sup>139</sup>. A further interesting observation in one study was a trend towards higher rates  
1488 of remission with one particular donor<sup>137</sup>.

1489

#### 1490 **5.6.2.3. Adverse events:**

1491 Short-lived GI symptoms such as abdominal bloating, cramps, diarrhoea and fever were reported in  
1492 patients receiving FMT for UC. There were no significant differences in serious adverse events  
1493 between patients receiving FMT compared to placebo (10 vs 7 respectively). Most of the serious  
1494 adverse events were a consequence of worsening colitis: one patient who received FMT required a  
1495 colectomy<sup>136</sup>. In addition, one patient developed concurrent CDI<sup>137</sup>. No deaths were reported in any  
1496 of the studies.

1497

#### 1498 **5.6.3. Use of FMT in functional bowel disorders:**

1499 Two RCTs have investigated the role of FMT in functional bowel disorders. In a double-blind placebo  
1500 controlled RCT that recruited 90 patients with IBS with diarrhoea or with diarrhoea and  
1501 constipation<sup>140</sup>, the primary endpoint only just reached statistical significance in inducing symptom  
1502 relief (as assessed by 75 point reduction in IBS-severity scoring system at three months following a  
1503 single infusion FMT by colonoscopy) ( $p=0.049$ ). The second RCT randomised 60 patients with slow  
1504 transit constipation to either six consecutive days of nasogastric-delivered FMT or conventional  
1505 treatment<sup>141</sup>. This demonstrated that a significant proportion of patients achieved the primary  
1506 endpoint of a mean of at least three complete spontaneous bowel movements per week (53.3% vs.  
1507 20.0%,  $p=0.009$ ) along with improvement in stool consistency score and colonic transit time.  
1508 However, the intervention group had more treatment-related adverse events than did the control  
1509 group (total of 50 vs 4 cases).

1510

#### 1511 **5.6.4. Use of FMT in hepatic encephalopathy:**

1512 One small study has investigated the role of FMT in the management of hepatic encephalopathy  
1513 (HE)<sup>142</sup>. This RCT randomised 20 male patients with cirrhosis with refractory HE to receive either five  
1514 days of broad-spectrum antibiotic pre-treatment followed by a single FMT enema or standard of

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care. Patients in the FMT arm had a significantly lower incidence of serious adverse events and improved cognition. The Model for End-Stage Liver Disease (MELD) score, however, transiently worsened post-antibiotics in the FMT arm. The study was potentially confounded as patients in the FMT arm continued to receive lactulose and/or rifaximin for treatment of their HE.

#### **5.6.5. Use of FMT for metabolic syndrome:**

Two randomised studies<sup>143,144</sup>, with a combined total of 56 patients, demonstrated an improvement in peripheral (but not hepatic) insulin sensitivity in Caucasian male obese patients with metabolic syndrome following one or two infusions via nasoduodenal tube of FMT obtained from lean donors. This improvement was observed at six weeks post-FMT, but was no longer present by 18 weeks. No improvement in insulin sensitivity was identified in patients transplanted with autologous FMT (i.e. patients transplanted with their own collected faeces). The improvement in peripheral insulin sensitivity in the lean donor FMT group was accompanied by a small but significant improvement in HbA1c at six weeks<sup>144</sup>, but no improvements in other metabolic parameters, such as weight. Whilst these data are of interest, the working group felt that the limited, transient nature of the benefits seen and small size of the studies meant that FMT could not be recommended as treatment for metabolic syndrome.

#### **5.6.6. Future directions for randomised trials of FMT for non-CDI indications:**

Currently there are a large number of randomised trials (including RCTs) being undertaken globally, to evaluate the potential role of FMT as treatment for a wide range of conditions. The working group concluded that until there are more reliable data to inform decision-making, the best practice principles described in this document for the governance of an FMT service for recurrent CDI should also be applied to FMT clinical trials for other conditions. However, specific adaptations may be considered depending on the condition being studied, e.g. consideration of using anaerobic conditions for the preparation of FMT in trials for the treatment of UC, as described above.

In conclusion, FMT has the potential to be an effective treatment option for mild to moderate ulcerative colitis, and appears to be safe despite the use of immunosuppressive therapy. FMT may also have a potential role in the treatment of functional bowel disorders. However, recommendations for clinical use for both these indications cannot be made until there is clearer evidence of the most appropriate patient characteristics, preparation methodology, route of delivery



1547 and intensity of administration of FMT. The evidence for the use of FMT in hepatic encephalopathy  
1548 and metabolic syndrome is currently limited, and further well-designed RCTs are needed to evaluate  
1549 its potential role here.

1551 **Recommendation:**

1552 ***We do not currently recommended FMT as treatment for inflammatory bowel disease.***  
1553 ***Apart from CDI, there is insufficient evidence to recommend FMT for any other***  
1554 ***gastrointestinal or non-gastrointestinal disease (GRADE of evidence: moderate; strength***  
1555 ***of recommendation: strong).***

1557 **6. Basic requirements for implementing a FMT service:**

1558 As discussed above, there is an absence of published studies to support the recommendations in this  
1559 section (although the experience of setting up a nationwide stool bank has recently been reported  
1560 from the Netherlands<sup>129</sup>). This section is therefore based on the working group’s expert opinion and  
1561 experience of developing FMT services. The working group considered best practice in this area as it  
1562 applied to legal and clinical governance aspects, the relevant professionals required to establish an  
1563 FMT service, the infrastructure of a service, and appropriate practices for FMT manufacturing and  
1564 quality control monitoring where relevant. The full text of this section is in **Supplementary Material**  
1565 **3.**

1567 **7. Key performance indicators:**

- 1568 • All donors to have completed initial screening questionnaires and blood and stool screening  
1569 results, as well as final health check prior to each stool donation processed to FMT. Results from  
1570 each subsequent serial round of screening also to be documented.
- 1571 • All FMT recipients to have clear documentation of details of their disease course and  
1572 preparation prior to FMT, including whether recurrent or refractory disease, previous  
1573 antimicrobial courses, and use of bowel purgatives/other preparatory medications pre-FMT.
- 1574 • All FMT recipients to have sufficient documentation to allow clear traceability of the exact FMT  
1575 aliquot transfused. Records should include identification of the donor, as well as a frozen FMT  
1576 aliquot (and original faecal sample) - as well as serum - from that donor (see **Supplementary**  
1577 **Material 3**).

- All FMT recipients for recurrent or refractory CDI to have documentation during follow-up of treatment success or failure (and subsequent treatment plan if failure), together with clear documentation of any adverse events that may be attributable to FMT.

## 8. Further research:

- As described within this guideline, many aspects of the terminology of CDI are used variably between studies, and end-points in FMT trials are inconsistent. The working group noted the need to standardise this terminology to allow more robust comparisons between studies.
- Given the relative novelty of FMT as a procedure, any potential long-term adverse events associated with its use are poorly-defined. The establishment of formal FMT registries should be considered. Whilst this would primarily act as an important tool for defining the safety and efficacy of FMT, it would also be a valuable database for researchers within the field. Standardisation of other key documentation related to FMT administration (e.g. establishment of a proforma for assessing eligibility for FMT and/or follow-up after FMT) would also be advantageous for the same reasons.
- The working group noted the lack of consistency in definitions related to the severity of CDI disease and to response or failure to FMT. This limited interpretation of the published studies. As such, the working group thought that standardisation of these definitions would allow more accurate delineation of the factors influencing the efficacy of FMT for CDI. The working group also noted that only one reviewed study had reported the relationship between *C difficile* ribotype and FMT outcome, and that recording of this information should be encouraged better to evaluate its influence.
- Further well-designed clinical trials (in particular, RCTs) are required to identify the optimal means of administration of FMT as treatment for recurrent and/or refractory CDI.
- The working group noted that even capsulised FMT may be associated with potential drawbacks. They also noted that there are many patients with recurrent CDI for whom FMT (or any form of 'bacteriotherapy') may be inappropriate, including those with very marked immunosuppression, and/or multi-organ disease. Despite high levels of efficacy, there is a small but appreciable FMT failure rate and it is not currently understood whether this is due to underlying donor or recipient factors. Therefore, a research priority should be in basic and translational studies better to define the mechanisms underlying the efficacy of FMT in CDI. This includes comparing the structure and function of the microbiota of donors to patients pre-FMT and post-FMT, via techniques including next-generation microbial sequencing, metabolic profiling, and

immunological assays. This would allow the refinement of FMT from its current state to a more targeted therapy, removing the concerns associated with FMT.

- The working group identified a need for further well-designed RCTs to investigate the potential role of FMT for non-CDI indications.

**9. Conclusions:**

FMT has become an accepted, efficacious treatment for recurrent and/or refractory CDI. In developing this guideline, the evidence for the technique has been reviewed in the context of other available treatments. Specific guidance for best practice for an FMT service is provided.

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**11. Competing interests:**

- THI: Acted as consultant, advisor or speaker for Pharmacosmos and Shield Therapeutics.
- ALH: Acted as consultant, advisory board member or speaker for AbbVie, Atlantic, Bristol-Myers Squibb, Celltrion, Falk, Ferring, Janssen, MSD, Napp Pharmaceuticals, Pfizer, Pharmacosmos, Shire and Takeda. ALH also serves on the Global Steering Committee for Genentech.
- SDG: Received consultancy fees, speaker fees and research grant support from Astellas between 2015-2017; received consultancy fees and speaker fees from MSD between 2015-2017; and received consultancy fees in 2017 from Pfizer.
- All other authors declared no conflict of interest.

**12. Provenance and peer review:**

Commissioned. Peer review through stakeholder consultation, HIS (SDC and Council), BSG (CSSC and Council) and externally.

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### **14.References:**

1. Lawson PA, Citron DM, Tyrrell KL, Finegold SM. Reclassification of *Clostridium difficile* as *Clostridioides difficile* (Hall and O'Toole 1935) Prévot 1938. *Anaerobe*. 2016;40:95-99. doi:10.1016/j.anaerobe.2016.06.008.
2. Faecal microbiota transplant for recurrent *Clostridium difficile* infection | Guidance and guidelines | NICE. <https://www.nice.org.uk/guidance/ipg485>. Accessed October 2, 2017.

1  
2  
3 1665 3. Health England P. Updated guidance on the management and treatment of Clostridium  
4 1666 difficile infection. 2013. <http://www.gov.uk/phe>. Accessed March 20, 2017.  
5  
6 1667 4. Debast SB, Bauer MP, Kuijper EJ. European Society of Clinical Microbiology and Infectious  
7 1668 Diseases: Update of the Treatment Guidance Document for Clostridium difficile Infection. *Clin*  
8 1669 *Microbiol Infect*. 2014;20(s2):1-26. doi:10.1111/1469-0691.12418.  
9  
10 1670 5. McDonald LC, Gerding DN, Johnson S, et al. Clinical Practice Guidelines for Clostridium  
11 1671 difficile Infection in Adults and Children: 2017 Update by the Infectious Diseases Society of  
12 1672 America (IDSA) and Society for Healthcare Epidemiology of America (SHEA). *Clin Infect Dis*.  
13 1673 February 2018. doi:10.1093/cid/cix1085.  
14  
15 1674 6. Cammarota G, Ianiro G, Tilg H, et al. European consensus conference on faecal microbiota  
16 1675 transplantation in clinical practice. *Gut*. 2017;66(4):569-580. doi:10.1136/gutjnl-2016-  
17 1676 313017.  
18  
19 1677 7. König J, Siebenhaar A, Högenauer C, et al. Consensus report: faecal microbiota transfer -  
20 1678 clinical applications and procedures. *Aliment Pharmacol Ther*. 2017;45(2):222-239.  
21 1679 doi:10.1111/apt.13868.  
22  
23 1680 8. Treating Clostridium difficile Infection With Fecal Microbiota Transplantation. *Clin*  
24 1681 *Gastroenterol Hepatol*. 2011;9(12):1044-1049. doi:10.1016/J.CGH.2011.08.014.  
25  
26 1682 9. Kump P, Krause R, Steininger C, et al. Empfehlungen zur Anwendung der fäkalen  
27 1683 Mikrobiotatransplantation „Stuhltransplantation“: Konsensus der Österreichischen  
28 1684 Gesellschaft für Gastroenterologie und Hepatologie (ÖGGH) in Zusammenarbeit mit der  
29 1685 Österreichischen Gesellschaft für Infektiologie und. *Z Gastroenterol*. 2014;52(12):1485-1492.  
30 1686 doi:10.1055/s-0034-1385562.  
31  
32 1687 10. Faecal microbiota transplantation in recurrent Clostridium difficile infection:  
33 1688 Recommendations from the French Group of Faecal microbiota Transplantation. *Dig Liver Dis*.  
34 1689 2016;48(3):242-247. doi:10.1016/J.DLD.2015.08.017.  
35  
36 1690 11. Kao D, Roach B, Silva M, et al. Effect of Oral Capsule– vs Colonoscopy-Delivered Fecal  
37 1691 Microbiota Transplantation on Recurrent *Clostridium difficile* Infection. *JAMA*.  
38 1692 2017;318(20):1985. doi:10.1001/jama.2017.17077.  
39  
40 1693 12. Hota SS, Sales V, Tomlinson G, et al. Oral Vancomycin Followed by Fecal Transplantation  
41 1694 Versus Tapering Oral Vancomycin Treatment for Recurrent *Clostridium difficile* Infection: An  
42 1695 Open-Label, Randomized Controlled Trial. *Clin Infect Dis*. 2017;64(3):265-271.  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60

HIS/ BSG FMT Guideline: Main Document, *Gut* version.

- doi:10.1093/cid/ciw731.
13. Jiang ZD, Ajami NJ, Petrosino JF, et al. Randomised clinical trial: faecal microbiota transplantation for recurrent *Clostridium difficile* infection – fresh, or frozen, or lyophilised microbiota from a small pool of healthy donors delivered by colonoscopy. *Aliment Pharmacol Ther.* 2017;45(7):899-908. doi:10.1111/apt.13969.
14. Kelly CR, Khoruts A, Staley C, et al. Effect of Fecal Microbiota Transplantation on Recurrence in Multiply Recurrent *Clostridium difficile* Infection. *Ann Intern Med.* 2016;165(9):609. doi:10.7326/M16-0271.
15. van Nood E, Vrieze A, Nieuwdorp M, et al. Duodenal Infusion of Donor Feces for Recurrent *Clostridium difficile*. *N Engl J Med.* 2013;368(5):407-415. doi:10.1056/NEJMoa1205037.
16. Lee CH, Steiner T, Petrof EO, et al. Frozen vs Fresh Fecal Microbiota Transplantation and Clinical Resolution of Diarrhea in Patients With Recurrent *Clostridium difficile* Infection. *JAMA.* 2016;315(2):142. doi:10.1001/jama.2015.18098.
17. Youngster I, Sauk J, Pindar C, et al. Fecal Microbiota Transplant for Relapsing *Clostridium difficile* Infection Using a Frozen Inoculum From Unrelated Donors: A Randomized, Open-Label, Controlled Pilot Study. *Clin Infect Dis.* 2014;58(11):1515-1522. doi:10.1093/cid/ciu135.
18. Cammarota G, Masucci L, Ianiro G, et al. Randomised clinical trial: faecal microbiota transplantation by colonoscopy vs. vancomycin for the treatment of recurrent *Clostridium difficile* infection. *Aliment Pharmacol Ther.* 2015;41(9):835-843. doi:10.1111/apt.13144.
19. Faecal Microbiota Transplantation (FMT) MHRA's position. [http://www.bsg.org.uk/images/stories/docs/clinical/guidance/fmt\\_mhra\\_position\\_june2015.pdf](http://www.bsg.org.uk/images/stories/docs/clinical/guidance/fmt_mhra_position_june2015.pdf). Accessed October 3, 2017.
20. Thomas A. HTA Policy on the Regulation of Faecal Microbiota Transplant. 2015. [http://www.bsg.org.uk/images/stories/docs/clinical/guidance/hta\\_pol\\_030\\_policy\\_regulation\\_of\\_fmt.pdf](http://www.bsg.org.uk/images/stories/docs/clinical/guidance/hta_pol_030_policy_regulation_of_fmt.pdf). Accessed October 3, 2017.
21. Khoruts A, Sadowsky MJ. Understanding the mechanisms of faecal microbiota transplantation. *Nat Rev Gastroenterol Hepatol.* 2016;13(9):508-516. doi:10.1038/nrgastro.2016.98.
22. Petrof EO, Gloor GB, Vanner SJ, et al. Stool substitute transplant therapy for the eradication of *Clostridium difficile* infection: 'RePOOPulating' the gut. *Microbiome.* 2013;1(1):3. doi:10.1186/2049-2618-1-3.

HIS/ BSG FMT Guideline: Main Document, *Gut* version.

- 1727 23. Ott SJ, Waetzig GH, Rehman A, et al. Efficacy of Sterile Fecal Filtrate Transfer for Treating
- 1728 Patients With *Clostridium difficile* Infection. *Gastroenterology*. 2017;152(4):799-811.e7.
- 1729 doi:10.1053/j.gastro.2016.11.010.
- 1730 24. Khanna S, Pardi DS, Kelly CR, et al. A Novel Microbiome Therapeutic Increases Gut Microbial
- 1731 Diversity and Prevents Recurrent *Clostridium difficile* Infection. *J Infect Dis*. 2016;214(2):173-
- 1732 181. doi:10.1093/infdis/jiv766.
- 1733 25. Martin J, Wilcox M. New and emerging therapies for *Clostridium difficile* infection. *Curr Opin*
- 1734 *Infect Dis*. 2016;29(6):546-554. doi:10.1097/QCO.0000000000000320.
- 1735 26. Zipursky JS, Sidorsky TI, Freedman CA, Sidorsky MN, Kirkland KB. Patient Attitudes Toward the
- 1736 Use of Fecal Microbiota Transplantation in the Treatment of Recurrent *Clostridium difficile*
- 1737 Infection. *Clin Infect Dis*. 2012;55(12):1652-1658. doi:10.1093/cid/cis809.
- 1738 27. Kahn SA, Vachon A, Rodriguez D, et al. Patient perceptions of fecal microbiota
- 1739 transplantation for ulcerative colitis. *Inflamm Bowel Dis*. 2013;19(7):1506-1513.
- 1740 doi:10.1097/MIB.0b013e318281f520.
- 1741 28. Quraishi MN, Segal J, Mullish B, et al. National survey of practice of faecal microbiota
- 1742 transplantation for *Clostridium difficile* infection in the UK. *J Hosp Infect*. 2016.
- 1743 doi:10.1016/j.jhin.2016.10.023.
- 1744 29. Porter RJ, Fogg C. Faecal microbiota transplantation for *Clostridium difficile* infection in the
- 1745 United Kingdom. *Clin Microbiol Infect*. 2015;21(6):578-582. doi:10.1016/j.cmi.2015.01.020.
- 1746 30. Ding NS, Mullish BH, McLaughlin J, Hart A, Marchesi JR. Meeting update: faecal microbiota
- 1747 transplantation—bench, bedside, courtroom? *Frontline Gastroenterol*. November
- 1748 2016:flgastro-2016-100752. doi:10.1136/flgastro-2016-100752.
- 1749 31. Prior AR, Kevans D, McDowell L, Cudmore S, Fitzpatrick F. Treatment of *Clostridium difficile*
- 1750 infection: a national survey of clinician recommendations and the use of faecal microbiota
- 1751 transplantation. *J Hosp Infect*. 2017;95(4):438-441. doi:10.1016/j.jhin.2016.10.004.
- 1752 32. 1995 - The well-built clinical question: a key to evidence-based decisions (Editorial) | 1995
- 1753 Nov-Dec : Volume 123, Number 3, Page A12 | ACP Journal Club Archives.
- 1754 <https://acpjc.acponline.org/Content/123/3/issue/ACPJC-1995-123-3-A12.htm>. Accessed
- 1755 October 18, 2017.
- 1756 33. British Society of Gastroenterology CS and SC. Guideline Development Within the BSG Clinical
- 1757 Services and Standards Committee Policies. <https://www.bsg.org.uk/resource/guideline->



HIS/ BSG FMT Guideline: Main Document, *Gut* version.

- development-within-the-bsg-clinical-services-and-standards-committee-policies.html.
- Accessed April 25, 2018.
34. Guyatt GH, Oxman AD, Vist GE, et al. GRADE: an emerging consensus on rating quality of evidence and strength of recommendations. *BMJ*. 2008;336(7650):924-926. doi:10.1136/bmj.39489.470347.AD.
35. Satokari R, Mattila E, Kainulainen V, Arkkila PET. Simple faecal preparation and efficacy of frozen inoculum in faecal microbiota transplantation for recurrent *Clostridium difficile* infection - an observational cohort study. *Aliment Pharmacol Ther*. 2015;41(1):46-53. doi:10.1111/apt.13009.
36. Yoon SS, Brandt LJ. Treatment of Refractory/Recurrent *C. difficile*-associated Disease by Donated Stool Transplanted Via Colonoscopy. *J Clin Gastroenterol*. 2010;44(8):562-566. doi:10.1097/MCG.0b013e3181dac035.
37. Zainah H, Hassan M, Shiekh-Sroujeh L, Hassan S, Alangaden G, Ramesh M. Intestinal microbiota transplantation, a simple and effective treatment for severe and refractory *Clostridium difficile* infection. *Dig Dis Sci*. 2014;60(1):181-185. doi:10.1007/s10620-014-3296-y.
38. Kassam Z. Fecal Transplant via Retention Enema for Refractory or Recurrent *Clostridium difficile* Infection. *Arch Intern Med*. 2012;172(2):191. doi:10.1001/archinte.172.2.191.
39. Aas J, Gessert CE, Bakken JS. Recurrent *Clostridium difficile* Colitis: Case Series Involving 18 Patients Treated with Donor Stool Administered via a Nasogastric Tube. *Clin Infect Dis*. 2003;36(5):580-585. doi:10.1086/367657.
40. Garborg K, Waagsbø B, Stallemo A, Matre J, Sundøy A. Results of faecal donor instillation therapy for recurrent *Clostridium difficile*-associated diarrhoea. *Scand J Infect Dis*. 2010;42(11-12):857-861. doi:10.3109/00365548.2010.499541.
41. Hamilton MJ, Weingarden AR, Sadowsky MJ, Khoruts A. Standardized Frozen Preparation for Transplantation of Fecal Microbiota for Recurrent *Clostridium difficile* Infection. *Am J Gastroenterol*. 2012;107(5):761-767. doi:10.1038/ajg.2011.482.
42. Lee CH, Belanger JE, Kassam Z, et al. The outcome and long-term follow-up of 94 patients with recurrent and refractory *Clostridium difficile* infection using single to multiple fecal microbiota transplantation via retention enema. *Eur J Clin Microbiol Infect Dis*.

1789 2014;33(8):1425-1428. doi:10.1007/s10096-014-2088-9.

1790 43. Mattila E, Uusitalo-Seppälä R, Wuorela M, et al. Fecal transplantation, through colonoscopy,  
1791 is effective therapy for recurrent Clostridium difficile infection. *Gastroenterology*.  
1792 2012;142(3):490-496. doi:10.1053/j.gastro.2011.11.037.

1793 44. Rohlke F, Surawicz CM, Stollman N. Fecal Flora Reconstitution for Recurrent Clostridium  
1794 difficile Infection: Results and Methodology. *J Clin Gastroenterol*. 2010;44(8):567-570.  
1795 doi:10.1097/MCG.0b013e3181dadadb10.

1796 45. Rubin TA, Gessert CE, Aas J, Bakken JS. Fecal microbiome transplantation for recurrent  
1797 Clostridium difficile infection: Report on a case series. *Anaerobe*. 2013;19:22-26.  
1798 doi:10.1016/j.anaerobe.2012.11.004.

1799 46. Patel NC, Griesbach CL, DiBaise JK, Orenstein R. Fecal Microbiota Transplant for Recurrent  
1800 Clostridium difficile Infection: Mayo Clinic in Arizona Experience. *Mayo Clin Proc*.  
1801 2013;88(8):799-805. doi:10.1016/j.mayocp.2013.04.022.

1802 47. Crobach MJT, Planche T, Eckert C, et al. European Society of Clinical Microbiology and  
1803 Infectious Diseases: update of the diagnostic guidance document for Clostridium difficile  
1804 infection. *Clin Microbiol Infect*. 2016;22 Suppl 4:S63-81. doi:10.1016/j.cmi.2016.03.010.

1805 48. Jackson M, Olefson S, Machan JT, Kelly CR. A High Rate of Alternative Diagnoses in Patients  
1806 Referred for Presumed Clostridium difficile Infection. *J Clin Gastroenterol*. 2016;50(9):742-  
1807 746. doi:10.1097/MCG.0000000000000447.

1808 49. Ray A, Smith R, Breaux J. Fecal Microbiota Transplantation for Clostridium difficile Infection:  
1809 The Ochsner Experience. *Ochsner J*. 2014;14(4):538-544.  
1810 <http://www.ncbi.nlm.nih.gov/pubmed/25598718>. Accessed October 9, 2017.

1811 50. Kao, D., Roach B., Hotte, N., Silva, M., Madsen, K., Beck, P., Louie T, Canadian Association for  
1812 the Study of the Liver. A Prospective, Dual Center, Randomized Trial Comparing Colonoscopy  
1813 versus Capsule Delivered Fecal Microbiota Transplantation (FMT) in the Management of  
1814 Recurrent Clostridium Difficile Infection (RCDI). In: *Canadian Journal of Gastroenterology and*  
1815 *Hepatology*. Vol 2016. Hindawi; 2016:1-204. doi:10.1155/2016/4792898.

1816 51. MacConnachie AA, Fox R, Kennedy DR, Seaton RA. Faecal transplant for recurrent Clostridium  
1817 difficile-associated diarrhoea: a UK case series. *QJM*. 2009;102(11):781-784.  
1818 doi:10.1093/qjmed/hcp118.

1819 52. Kelly CR, de Leon L, Jasutkar N. Fecal Microbiota Transplantation for Relapsing Clostridium

HIS/ BSG FMT Guideline: Main Document, Gut version.

- 1820 difficile Infection in 26 Patients. *J Clin Gastroenterol*. 2012;46(2):145-149.
- 1821 doi:10.1097/MCG.0b013e318234570b.
- 1822 53. Brandt LJ, Aroniadis OC, Mellow M, et al. Long-Term Follow-Up of Colonoscopic Fecal
- 1823 Microbiota Transplant for Recurrent Clostridium difficile Infection. *Am J Gastroenterol*.
- 1824 2012;107(7):1079-1087. doi:10.1038/ajg.2012.60.
- 1825 54. Pathak R, Enuh HA, Patel A, Wickremesinghe P. Treatment of relapsing Clostridium difficile
- 1826 infection using fecal microbiota transplantation. *Clin Exp Gastroenterol*. 2013;7:1-6.
- 1827 doi:10.2147/CEG.S53410.
- 1828 55. Agrawal M, Aroniadis OC, Brandt LJ, et al. The Long-term Efficacy and Safety of Fecal
- 1829 Microbiota Transplant for Recurrent, Severe, and Complicated Clostridium difficile Infection
- 1830 in 146 Elderly Individuals. *J Clin Gastroenterol*. 2015;50(5):1.
- 1831 doi:10.1097/MCG.0000000000000410.
- 1832 56. Fischer M, Kao D, Kelly C, et al. Fecal Microbiota Transplantation is Safe and Efficacious for
- 1833 Recurrent or Refractory Clostridium difficile Infection in Patients with Inflammatory Bowel
- 1834 Disease. *Inflamm Bowel Dis*. 2016;22(10):2402-2409. doi:10.1097/MIB.0000000000000908.
- 1835 57. Aroniadis OC, Brandt LJ, Greenberg A, et al. Long-term Follow-up Study of Fecal Microbiota
- 1836 Transplantation for Severe and/or Complicated Clostridium difficile Infection. *J Clin*
- 1837 *Gastroenterol*. 2015;50(5):1. doi:10.1097/MCG.0000000000000374.
- 1838 58. Fischer M, Kao D, Mehta SR, et al. Predictors of Early Failure After Fecal Microbiota
- 1839 Transplantation for the Therapy of Clostridium Difficile Infection: A Multicenter Study. *Am J*
- 1840 *Gastroenterol*. 2016;111(7):1024-1031. doi:10.1038/ajg.2016.180.
- 1841 59. Ianaro G, Valerio L, Masucci L, et al. Predictors of failure after single faecal microbiota
- 1842 transplantation in patients with recurrent Clostridium difficile infection: results from a 3-year,
- 1843 single-centre cohort study. *Clin Microbiol Infect*. 2017;23(5):337.e1-337.e3.
- 1844 doi:10.1016/j.cmi.2016.12.025.
- 1845 60. Kelly CR, Ihunnah C, Fischer M, et al. Fecal microbiota transplant for treatment of Clostridium
- 1846 difficile infection in immunocompromised patients. *Am J Gastroenterol*. 2014;109(7):1065-
- 1847 1071. doi:10.1038/ajg.2014.133.
- 1848 61. Lagier J-C, Delord M, Million M, et al. Dramatic reduction in Clostridium difficile ribotype 027-
- 1849 associated mortality with early fecal transplantation by the nasogastric route: a preliminary
- 1850 report. *Eur J Clin Microbiol Infect Dis*. 2015;34(8):1597-1601. doi:10.1007/s10096-015-2394-x.

HIS/ BSG FMT Guideline: Main Document, *Gut* version.

- 1851 62. Camacho-Ortiz A, Gutiérrez-Delgado EM, Garcia-Mazcorro JF, et al. Randomized clinical trial  
1852 to evaluate the effect of fecal microbiota transplant for initial *Clostridium difficile* infection in  
1853 intestinal microbiome. Green J, ed. *PLoS One*. 2017;12(12):e0189768.  
1854 doi:10.1371/journal.pone.0189768.
- 1855 63. Louie TJ, Miller MA, Mullane KM, et al. Fidaxomicin versus Vancomycin for *Clostridium*  
1856 *difficile* Infection. *N Engl J Med*. 2011;364(5):422-431. doi:10.1056/NEJMoa0910812.
- 1857 64. Wilcox MH, Gerding DN, Poxton IR, et al. Bezlotoxumab for Prevention of Recurrent  
1858 *Clostridium difficile* Infection. *N Engl J Med*. 2017;376(4):305-317.  
1859 doi:10.1056/NEJMoa1602615.
- 1860 65. Guery B, Menichetti F, Anttila V-J, et al. Extended-pulsed fidaxomicin versus vancomycin for  
1861 *Clostridium difficile* infection in patients 60 years and older (EXTEND): a randomised,  
1862 controlled, open-label, phase 3b/4 trial. *Lancet Infect Dis*. December 2017.  
1863 doi:10.1016/S1473-3099(17)30751-X.
- 1864 66. McFarland L V., Elmer GW, Surawicz CM. Breaking the cycle: treatment strategies for 163  
1865 cases of recurrent *Clostridium difficile* disease. *Am J Gastroenterol*. 2002;97(7):1769-1775.  
1866 doi:10.1111/j.1572-0241.2002.05839.x.
- 1867 67. Sirbu BD, Soriano MM, Manzo C, Lum J, Gerding DN, Johnson S. Vancomycin Taper and Pulse  
1868 Regimen With Careful Follow-up for Patients With Recurrent *Clostridium difficile* Infection.  
1869 *Clin Infect Dis*. 2017;65(8):1396-1399. doi:10.1093/cid/cix529.
- 1870 68. Gentry CA, Giancola SE, Thind S, Kurdgelashvili G, Skrepnek GH, Williams RJ. A Propensity-  
1871 Matched Analysis Between Standard Versus Tapered Oral Vancomycin Courses for the  
1872 Management of Recurrent *Clostridium difficile* Infection. *Open Forum Infect Dis*.  
1873 2017;4(4):ofx235. doi:10.1093/ofid/ofx235.
- 1874 69. Cornely OA, Crook DW, Esposito R, et al. Fidaxomicin versus vancomycin for infection with  
1875 *Clostridium difficile* in Europe, Canada, and the USA: a double-blind, non-inferiority,  
1876 randomised controlled trial. *Lancet Infect Dis*. 2012;12(4):281-289. doi:10.1016/S1473-  
1877 3099(11)70374-7.
- 1878 70. Tauxe WM, Haydek JP, Rebolledo PA, et al. Fecal microbiota transplant for *Clostridium*  
1879 *difficile* infection in older adults. *Therap Adv Gastroenterol*. 2016;9(3):273-281.  
1880 doi:10.1177/1756283X15622600.
- 1881 71. Khan MA, Sofi AA, Ahmad U, et al. Efficacy and safety of, and patient satisfaction with,

HIS/ BSG FMT Guideline: Main Document, *Gut* version.

- 1882 colonoscopic-administered fecal microbiota transplantation in relapsing and refractory
- 1883 community- and hospital-acquired *Clostridium difficile* infection. *Can J Gastroenterol Hepatol*.
- 1884 2014;28(8):434-438. <http://www.ncbi.nlm.nih.gov/pubmed/25014180>. Accessed October 9,
- 1885 2017.
- 1886 72. Youngster I, Russell GH, Pindar C, Ziv-Baran T, Sauk J, Hohmann EL. Oral, Capsulized, Frozen
- 1887 Fecal Microbiota Transplantation for Relapsing *Clostridium difficile* Infection. *JAMA*.
- 1888 2014;312(17):1772. doi:10.1001/jama.2014.13875.
- 1889 73. Fischer M, Sipe BW, Rogers NA, et al. Faecal microbiota transplantation plus selected use of
- 1890 vancomycin for severe-complicated *Clostridium difficile* infection: description of a protocol
- 1891 with high success rate. *Aliment Pharmacol Ther*. 2015;42(4):470-476. doi:10.1111/apt.13290.
- 1892 74. Costello SP, Conlon MA, Vuaran MS, Roberts-Thomson IC, Andrews JM. Faecal microbiota
- 1893 transplant for recurrent *Clostridium difficile* infection using long-term frozen stool is
- 1894 effective: Clinical efficacy and bacterial viability data. *Aliment Pharmacol Ther*.
- 1895 2015;42(8):1011-1018. doi:10.1111/apt.13366.
- 1896 75. Hui J, Kench JG, Chitturi S, et al. Long-Term outcomes of cirrhosis in nonalcoholic
- 1897 steatohepatitis compared with hepatitis C. *Hepatology*. 2003;38(2):420-427.
- 1898 doi:10.1053/jhep.2003.50320.
- 1899 76. Allegretti JR, Korzenik JR, Hamilton MJ. Fecal microbiota transplantation via colonoscopy for
- 1900 recurrent *C. difficile* Infection. *J Vis Exp*. 2014;(94). doi:10.3791/52154.
- 1901 77. van Beurden YH, de Groot PF, van Nood E, Nieuwdorp M, Keller JJ, Goorhuis A. Complications,
- 1902 effectiveness, and long term follow-up of fecal microbiota transfer by nasoduodenal tube for
- 1903 treatment of recurrent *Clostridium difficile* infection. *United Eur Gastroenterol J*.
- 1904 2017;5(6):868-879. doi:10.1177/2050640616678099.
- 1905 78. Bakken JS, Borody T, Brandt LJ, et al. Treating *Clostridium difficile* Infection With Fecal
- 1906 Microbiota Transplantation. *Clin Gastroenterol Hepatol*. 2011;9(12):1044-1049.
- 1907 doi:10.1016/j.cgh.2011.08.014.
- 1908 79. Allegretti JR, Allegretti AS, Phelps E, Xu H, Kassam Z, Fischer M. Asymptomatic *Clostridium*
- 1909 *difficile* carriage rate post-fecal microbiota transplant is low: a prospective clinical and stool
- 1910 assessment. *Clin Microbiol Infect*. November 2017. doi:10.1016/J.CMI.2017.10.022.
- 1911 80. Baxter M, Colville A. Adverse events in faecal microbiota transplant: a review of the
- 1912 literature. *J Hosp Infect*. 2016;92(2):117-127. doi:10.1016/j.jhin.2015.10.024.

1  
2  
3 1913 81. Hefazi M, Patnaik MM, Hogan WJ, Litzow MR, Pardi DS, Khanna S. Safety and Efficacy of Fecal  
4 1914 Microbiota Transplant for Recurrent Clostridium difficile Infection in Patients With Cancer  
5 1915 Treated With Cytotoxic Chemotherapy: A Single-Institution Retrospective Case Series. *Mayo*  
6 1916 *Clin Proc.* 2017;92(11):1617-1624. doi:10.1016/j.mayocp.2017.08.016.  
7  
8  
9  
10 1917 82. Qazi T, Amaratunga T, Barnes EL, Fischer M, Kassam Z, Allegretti JR. The risk of inflammatory  
11 1918 bowel disease flares after fecal microbiota transplantation: Systematic review and meta-  
12 1919 analysis. *Gut Microbes.* July 2017:1-15. doi:10.1080/19490976.2017.1353848.  
13  
14  
15 1920 83. Meighani A, Hart BR, Mittal C, Miller N, John A, Ramesh M. Predictors of fecal transplant  
16 1921 failure. *Eur J Gastroenterol Hepatol.* 28:826-830. doi:10.1097/MEG.0000000000000614.  
17  
18  
19 1922 84. Alrabaa S, Jariwala R, Zeitler K, Montero J. Fecal microbiota transplantation outcomes in  
20 1923 immunocompetent and immunocompromised patients: A single-center experience. *Transpl*  
21 1924 *Infect Dis.* 2017;19(4):e12726. doi:10.1111/tid.12726.  
22  
23  
24 1925 85. Cohen NA, Livovsky DM, Yaakobovitch S, et al. A Retrospective Comparison of Fecal Microbial  
25 1926 Transplantation Methods for Recurrent Clostridium Difficile Infection. *Isr Med Assoc J.*  
26 1927 2016;18(10):594-599.  
27  
28  
29 1928 86. Orenstein R, Dubberke E, Hardi R, et al. Safety and Durability of RBX2660 (Microbiota  
30 1929 Suspension) for Recurrent *Clostridium difficile* Infection: Results of the PUNCH CD Study. *Clin*  
31 1930 *Infect Dis.* 2016;62(5):596-602. doi:10.1093/cid/civ938.  
32  
33  
34 1931 87. Hirsch BE, Saraiya N, Poeth K, Schwartz RM, Epstein ME, Honig G. Effectiveness of fecal-  
35 1932 derived microbiota transfer using orally administered capsules for recurrent Clostridium  
36 1933 difficile infection. *BMC Infect Dis.* 2015;15(1):191. doi:10.1186/s12879-015-0930-z.  
37  
38  
39 1934 88. Kao D, Roach B, Beck P, Hotte N, Madsen K, Louie T. A dual center, randomized trial  
40 1935 comparing colonoscopy and oral capsule delivered fecal microbiota transplantation in the  
41 1936 treatment of recurrent clostridium difficile infection: Preliminary results. *Am J Gastroenterol.*  
42 1937 2015;110:S553. doi:http://dx.doi.org/10.1038/ajg.2015.294.  
43  
44  
45 1938 89. Youngster I, Mahabamunuge J, Systrom HK, et al. Oral, frozen fecal microbiota transplant  
46 1939 (FMT) capsules for recurrent Clostridium difficile infection. *BMC Med.* 2016;14(1):134.  
47 1940 doi:10.1186/s12916-016-0680-9.  
48  
49  
50 1941 90. Anand R, Song Y, Garg S, et al. Effect of Aging on the Composition of Fecal Microbiota in  
51 1942 Donors for FMT and Its Impact on Clinical Outcomes. *Dig Dis Sci.* 2017;62(4):1002-1008.  
52 1943 doi:10.1007/s10620-017-4449-6.  
53  
54  
55  
56  
57  
58  
59  
60



HIS/ BSG FMT Guideline: Main Document, *Gut* version.

- 1944 91. Alang N, Kelly CR. Weight gain after fecal microbiota transplantation. *Open forum Infect Dis.*  
1945 2015;2(1):ofv004. doi:10.1093/ofid/ofv004.
- 1946 92. Fischer M, Kao D, Kassam Z, et al. Stool Donor Body Mass Index Does Not Affect Recipient  
1947 Weight After a Single Fecal Microbiota Transplantation for *C. difficile* Infection. *Clin*  
1948 *Gastroenterol Hepatol.* December 2017. doi:10.1016/J.CGH.2017.12.007.
- 1949 93. Marchesi JR, Adams DH, Fava F, et al. The gut microbiota and host health: a new clinical  
1950 frontier. *Gut.* 2016;65(2):330-339. doi:10.1136/gutjnl-2015-309990.
- 1951 94. Allegretti JR, Fischer M, Papa E, et al. Su1738 Fecal Microbiota Transplantation Delivered via  
1952 Oral Capsules Achieves Microbial Engraftment Similar to Traditional Delivery Modalities:  
1953 Safety, Efficacy and Engraftment Results From a Multi-Center Cluster Randomized Dose-  
1954 Finding Study. *Gastroenterology.* 2016;150(4):S540. doi:10.1016/S0016-5085(16)31855-8.
- 1955 95. Langdon A, Crook N, Dantas G. The effects of antibiotics on the microbiome throughout  
1956 development and alternative approaches for therapeutic modulation. *Genome Med.*  
1957 2016;8(1):39. doi:10.1186/s13073-016-0294-z.
- 1958 96. Lange K, Buerger M, Stallmach A, Bruns T. Effects of Antibiotics on Gut Microbiota. *Dig Dis.*  
1959 2016;34(3):260-268. doi:10.1159/000443360.
- 1960 97. Becattini S, Taur Y, Pamer EG. Antibiotic-Induced Changes in the Intestinal Microbiota and  
1961 Disease. *Trends Mol Med.* 2016;22(6):458-478. doi:10.1016/j.molmed.2016.04.003.
- 1962 98. Ianiro G, Tilg H, Gasbarrini A. Antibiotics as deep modulators of gut microbiota: between  
1963 good and evil. *Gut.* 2016;65(11):1906-1915. doi:10.1136/gutjnl-2016-312297.
- 1964 99. Boostrom SY, Mathis KL, Pendlimari R, et al. Burden of *Clostridium difficile* on the healthcare  
1965 system. *Inflamm Bowel Dis.* 2012;18(1):994-1002. doi:10.1126/science.1241214.
- 1966 100. J.R. A, J.R. K, M.J. H. Intestinal microbiome restoration for recurrent *clostridium difficile*  
1967 infection in patients with concurrent inflammatory bowel disease. *Gastroenterology.*  
1968 2015;148(4 SUPPL. 1):S869.
- 1969 101. National Health Service. Population screening programmes: NHS bowel cancer screening  
1970 (BCSP) programme - GOV.UK. [https://www.gov.uk/topic/population-screening-](https://www.gov.uk/topic/population-screening-programmes/bowel)  
1971 [programmes/bowel](https://www.gov.uk/topic/population-screening-programmes/bowel). Accessed June 10, 2018.
- 1972 102. London: TSO Guidelines for the Blood Transfusion Services in the United Kingdom 7 th Edition  
1973 2005 TSO Accredited Agents Web Access.



1  
2  
3 1974 103. Emanuelsson F, Claesson BEB, Ljungström L, Tvede M, Ung K-A. Faecal microbiota  
4 1975 transplantation and bacteriotherapy for recurrent Clostridium difficile infection: A  
5 1976 retrospective evaluation of 31 patients. *Scand J Infect Dis*. 2014;46(2):89-97.  
6 1977 doi:10.3109/00365548.2013.858181.  
7  
8  
9  
10 1978 104. Endtz HP, van den Braak N, van Belkum A, et al. Fecal carriage of vancomycin-resistant  
11 1979 enterococci in hospitalized patients and those living in the community in The Netherlands. *J*  
12 1980 *Clin Microbiol*. 1997;35(12):3026-3031. <http://www.ncbi.nlm.nih.gov/pubmed/9399488>.  
13 1981 Accessed February 15, 2018.  
14  
15  
16 1982 105. Willems RJL, Top J, van Santen M, et al. Global spread of vancomycin-resistant Enterococcus  
17 1983 faecium from distinct nosocomial genetic complex. *Emerg Infect Dis*. 2005;11(6):821-828.  
18 1984 doi:10.3201/eid1106.041204.  
19  
20  
21  
22 1985 106. Gough E, Shaikh H, Manges AR. Systematic Review of Intestinal Microbiota Transplantation  
23 1986 (Fecal Bacteriotherapy) for Recurrent Clostridium difficile Infection. *Clin Infect Dis*.  
24 1987 2011;53(10):994-1002. doi:10.1093/cid/cir632.  
25  
26  
27 1988 107. Casewell MW, Slater NGP, Cooper JE. Operating theatre water-baths as a cause of  
28 1989 pseudomonas septicaemia. *J Hosp Infect*. 1981;2:237-240. doi:10.1016/0195-6701(81)90043-  
29 1990 8.  
30  
31  
32  
33 1991 108. Muyltermans G, de Smet F, Pierard D, et al. Neonatal infections with Pseudomonas  
34 1992 aeruginosa associated with a water-bath used to thaw fresh frozen plasma. *J Hosp Infect*.  
35 1993 1998;39(4):309-314. <http://www.ncbi.nlm.nih.gov/pubmed/9749402>. Accessed June 11,  
36 1994 2018.  
37  
38  
39  
40 1995 109. Sleight SC, Wigginton NS, Lenski RE. Increased susceptibility to repeated freeze-thaw cycles in  
41 1996 Escherichia coli following long-term evolution in a benign environment. *BMC Evol Biol*.  
42 1997 2006;6(1):104. doi:10.1186/1471-2148-6-104.  
43  
44  
45 1998 110. O'Brien CL, Allison GE, Grimpen F, Pavli P. Impact of colonoscopy bowel preparation on  
46 1999 intestinal microbiota. *PLoS One*. 2013;8(5):e62815. doi:10.1371/journal.pone.0062815.  
47  
48  
49 2000 111. Mai V, Stine OC. Bowel preparation for colonoscopy: relevant for the gut's microbiota? *Gut*.  
50 2001 2015;64(10):1504-1505. doi:10.1136/gutjnl-2014-308937.  
51  
52  
53 2002 112. Jalanka J, Salonen A, Salojärvi J, et al. Effects of bowel cleansing on the intestinal microbiota.  
54 2003 *Gut*. 2015;64(10):1562-1568. doi:10.1136/gutjnl-2014-307240.  
55  
56 2004 113. Mai V, Greenwald B, Glenn Morris J, Raufman J-P, Stine OC. Effect of bowel preparation and  
57  
58  
59  
60

HIS/ BSG FMT Guideline: Main Document, *Gut* version.

- 2005 colonoscopy on post-procedure intestinal microbiota composition. *Gut*. 2006;55(12):1822-1823. doi:10.1136/gut.2006.108266.
- 2006
- 2007 114. Harrell L, Wang Y, Antonopoulos D, et al. Standard Colonic Lavage Alters the Natural State of Mucosal-Associated Microbiota in the Human Colon. Singh SR, ed. *PLoS One*. 2012;7(2):e32545. doi:10.1371/journal.pone.0032545.
- 2008
- 2009
- 2010 115. Chin SM, Sauk J, Mahabamunuge J, Kaplan JL, Hohmann EL, Khalili H. Fecal Microbiota Transplantation for Recurrent *Clostridium difficile* Infection in Patients With Inflammatory Bowel Disease: A Single-Center Experience. *Clin Gastroenterol Hepatol*. 2017;15(4):597-599. doi:10.1016/j.cgh.2016.11.028.
- 2011
- 2012
- 2013
- 2014 116. Wang S, Xu M, Wang W, et al. Systematic Review: Adverse Events of Fecal Microbiota Transplantation. *PLoS One*. 2016;11(8):e0161174. doi:10.1371/journal.pone.0161174.
- 2015
- 2016 117. Khoruts A, Rank KM, Newman KM, et al. Inflammatory Bowel Disease Affects the Outcome of Fecal Microbiota Transplantation for Recurrent *Clostridium difficile* Infection. *Clin Gastroenterol Hepatol*. 2016;14(10):1433-1438. doi:10.1016/j.cgh.2016.02.018.
- 2017
- 2018
- 2019 118. de Jager CPC, Wever PC, Gemen EFA, et al. Proton pump inhibitor therapy predisposes to community-acquired *Streptococcus pneumoniae* pneumonia. *Aliment Pharmacol Ther*. 2012;36(10):941-949. doi:10.1111/apt.12069.
- 2020
- 2021
- 2022 119. Imhann F, Bonder MJ, Vich Vila A, et al. Proton pump inhibitors affect the gut microbiome. *Gut*. 2016;65(5):740-748. doi:10.1136/gutjnl-2015-310376.
- 2023
- 2024 120. McDonald EG, Milligan J, Frenette C, Lee TC. Continuous Proton Pump Inhibitor Therapy and the Associated Risk of Recurrent *Clostridium difficile* Infection. *JAMA Intern Med*. 2015;175(5):784. doi:10.1001/jamainternmed.2015.42.
- 2025
- 2026
- 2027 121. Janarthanan S, Ditah I, Adler DG, Ehrinpreis MN. *Clostridium difficile*-Associated Diarrhea and Proton Pump Inhibitor Therapy: A Meta-Analysis. *Am J Gastroenterol*. 2012;107(7):1001-1010. doi:10.1038/ajg.2012.179.
- 2028
- 2029
- 2030 122. Girotra M, Garg S, Anand R, Song Y, Dutta SK. Fecal Microbiota Transplantation for Recurrent *Clostridium difficile* Infection in the Elderly: Long-Term Outcomes and Microbiota Changes. *Dig Dis Sci*. 2016;61(10):3007-3015. doi:10.1007/s10620-016-4229-8.
- 2031
- 2032
- 2033 123. Hagel S, Fischer A, Ehlermann P, et al. Fecal Microbiota Transplant in Patients With Recurrent *Clostridium Difficile* Infection. *Dtsch Arztebl Int*. 2016;113(35-36):583-589. doi:10.3238/arztebl.2016.0583.
- 2034
- 2035

2036 124. Department of Health (UK). Management and decontamination of flexible endoscopes (HTM  
2037 01-06) - GOV.UK. [https://www.gov.uk/government/publications/management-and-](https://www.gov.uk/government/publications/management-and-decontamination-of-flexible-endoscopes)  
2038 decontamination-of-flexible-endoscopes. Accessed December 19, 2017.

2039 125. British Society of Gastroenterology. Guidance on Decontamination of Equipment for  
2040 Gastrointestinal Endoscopy: 2017 Edition. [https://www.bsg.org.uk/resource/guidance-on-](https://www.bsg.org.uk/resource/guidance-on-decontamination-of-equipment-for-gastrointestinal-endoscopy-2017-edition.html)  
2041 decontamination-of-equipment-for-gastrointestinal-endoscopy-2017-edition.html. Accessed  
2042 December 19, 2017.

2043 126. Loveday HP, Wilson JA, Pratt RJ, et al. epic3: National Evidence-Based Guidelines for  
2044 Preventing Healthcare-Associated Infections in NHS Hospitals in England. *J Hosp Infect.*  
2045 2014;86(1):S1-S70. doi:10.1016/S0195-6701(13)60012-2.

2046 127. Kassam Z, Lee CH, Yuan Y, Hunt RH. Fecal Microbiota Transplantation for Clostridium difficile  
2047 Infection: Systematic Review and Meta-Analysis. *Am J Gastroenterol.* 2013;108(4):500-508.  
2048 doi:10.1038/ajg.2013.59.

2049 128. Allegretti JR, Kao D, Sitko J, Fischer M, Kassam Z. Early antibiotic use post-fecal microbiota  
2050 transplantation increases the risk of treatment failure. *Clin Infect Dis.* August 2017.  
2051 doi:10.1093/cid/cix684.

2052 129. Terveer EM, van Beurden YH, Goorhuis A, et al. How to: Establish and run a stool bank. *Clin*  
2053 *Microbiol Infect.* 2017;23(12):924-930. doi:10.1016/j.cmi.2017.05.015.

2054 130. Quraishi MN, Widlak M, Bhala N, et al. Systematic review with meta-analysis: the efficacy of  
2055 faecal microbiota transplantation for the treatment of recurrent and refractory *Clostridium*  
2056 *difficile* infection. *Aliment Pharmacol Ther.* 2017;46(5):479-493. doi:10.1111/apt.14201.

2057 131. Cammarota G, Ianiro G, Gasbarrini A. Fecal microbiota transplantation for the treatment of  
2058 Clostridium difficile infection: a systematic review. *J Clin Gastroenterol.* 2014;48(8):693-702.  
2059 doi:10.1097/MCG.000000000000046.

2060 132. D. D, J. R, S. G, et al. Fecal microbiota transplantation for clostridium difficile infection a  
2061 systematic review. *Ann Intern Med.* 2015;162(9):630-638.

2062 133. Baxter M, Ahmad T, Colville A, Sheridan R. Fatal Aspiration Pneumonia as a Complication of  
2063 Fecal Microbiota Transplant. *Clin Infect Dis.* 2015;61(1):136-137. doi:10.1093/cid/civ247.

2064 134. Hecker MT, Obrenovich ME, Cadnum JL, et al. Fecal Microbiota Transplantation by Freeze-  
2065 Dried Oral Capsules for Recurrent Clostridium difficile Infection. *Open forum Infect Dis.*  
2066 2016;3(2):ofw091. doi:10.1093/ofid/ofw091.

HIS/ BSG FMT Guideline: Main Document, Gut version.

- 2067 135. Ni J, Wu GD, Albenberg L, Tomov VT. Gut microbiota and IBD: causation or correlation? *Nat*  
2068 *Rev Gastroenterol Hepatol.* 2017;14(10):573-584. doi:10.1038/nrgastro.2017.88.
- 2069 136. Rossen NG, Fuentes S, van der Spek MJ, et al. Findings From a Randomized Controlled Trial of  
2070 Fecal Transplantation for Patients With Ulcerative Colitis. *Gastroenterology.* 2015;149(1):110-  
2071 118.e4. doi:10.1053/j.gastro.2015.03.045.
- 2072 137. Moayyedi P, Surette MG, Kim PT, et al. Fecal Microbiota Transplantation Induces Remission in  
2073 Patients With Active Ulcerative Colitis in a Randomized Controlled Trial. *Gastroenterology.*  
2074 2015;149(1):102-109.e6. doi:10.1053/j.gastro.2015.04.001.
- 2075 138. Paramsothy S, Kamm MA, Kaakoush NO, et al. Multidonor intensive faecal microbiota  
2076 transplantation for active ulcerative colitis: a randomised placebo-controlled trial. *Lancet*  
2077 *(London, England).* 2017;389(10075):1218-1228. doi:10.1016/S0140-6736(17)30182-4.
- 2078 139. Costello S, Waters O, Bryant R. Short Duration, Low Intensity, Pooled Fecal Microbiota  
2079 Transplantation Induces Remission in Patients with Mild-Moderately Active Ulcerative Colitis:  
2080 A Randomised Controlled Trial. (Abstract). *Gastroenterology.* 152(5):S198-S199.
- 2081 140. Johnsen PH, Hilpüsch F, Cavanagh JP, et al. Faecal microbiota transplantation versus placebo  
2082 for moderate-to-severe irritable bowel syndrome: a double-blind, randomised, placebo-  
2083 controlled, parallel-group, single-centre trial. *lancet Gastroenterol Hepatol.* 2018;3(1):17-24.  
2084 doi:10.1016/S2468-1253(17)30338-2.
- 2085 141. Tian H, Ge X, Nie Y, et al. Fecal microbiota transplantation in patients with slow-transit  
2086 constipation: A randomized, clinical trial. Green J, ed. *PLoS One.* 2017;12(2):e0171308.  
2087 doi:10.1371/journal.pone.0171308.
- 2088 142. Bajaj JS, Kassam Z, Fagan A, et al. Fecal Microbiota Transplant from a Rational Stool Donor  
2089 Improves Hepatic Encephalopathy: A Randomized Clinical Trial. *Hepatology.* June 2017.  
2090 doi:10.1002/hep.29306.
- 2091 143. Vrieze A, Van Nood E, Holleman F, et al. Transfer of Intestinal Microbiota From Lean Donors  
2092 Increases Insulin Sensitivity in Individuals With Metabolic Syndrome. *Gastroenterology.*  
2093 2012;143(4):913-916.e7. doi:10.1053/j.gastro.2012.06.031.
- 2094 144. Kootte RS, Levin E, Salojärvi J, et al. Improvement of Insulin Sensitivity after Lean Donor Feces  
2095 in Metabolic Syndrome Is Driven by Baseline Intestinal Microbiota Composition. *Cell Metab.*  
2096 2017;26(4):611-619.e6. doi:10.1016/j.cmet.2017.09.008.
- 2097

**15. Figure legends and tables:**

**Figure 1: Proposed summary pathway for donor screening for centres preparing frozen FMT from recurring donors.**

**Table 1: Recommended donor history/ questionnaire:** A positive response to any of these questions would usually result in exclusion from further consideration as a donor, although this would depend upon the particular circumstances/ answers given.

1. Receipt of antimicrobials within the past three months.
2. Known prior exposure to HIV and/ or viral hepatitis, and known previous or latent tuberculosis.
3. Risk factors for blood-borne viruses - including high risk sexual behaviours, use of illicit drugs, any tattoo/ body piercing/ needlestick injury/ blood transfusion/ acupuncture, all within the previous six months.
4. Receipt of a live attenuated virus within the past six months.
5. Underlying gastrointestinal conditions/ symptoms (e.g. history of IBD, IBS, chronic diarrhoea, chronic constipation, coeliac disease, bowel resection or bariatric surgery) - also including acute diarrhoea/ gastrointestinal symptoms within the past two weeks.
6. Family history of any significant gastrointestinal conditions (e.g. family history of IBD, or colorectal cancer).
7. History of atopy (e.g. asthma, eosinophilic disorders).
8. Any systemic autoimmune conditions.
9. Any metabolic conditions, including diabetes and obesity.
10. Any neurological or psychiatric conditions, or known risk of prion disease.
11. History of chronic pain syndromes, including chronic fatigue syndrome and fibromyalgia.
12. History of any malignancy.
13. Taking particular regular medications, or such medications within the past three months, i.e. antimicrobials, proton pump inhibitors, immunosuppression, chemotherapy
14. History of receiving growth hormone, insulin from cows, or clotting factor concentrates.
15. History of receiving an experimental medicine or vaccine within the past six months.
16. History of travel to tropical countries within the past six months.

**Table 2: Recommended blood screening for stool donors:** \*EBV and CMV testing is only recommended where there is the potential that the FMT prepared from that donor will be administered to immunosuppressed patients at risk of severe infection if exposed to CMV and EBV.

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*Pathogen screening:*

- Hepatitis A IgM
- Hepatitis B (HBsAg and HBcAb)
- Hepatitis C antibody
- Hepatitis E IgM
- HIV -1 and -2 antibodies
- HTLV-1 and -2 antibodies
- *Treponema pallidum* antibodies (TPHA, VDRL)
- Epstein-Barr virus IgM and IgG\*
- Cytomegalovirus IgM and IgG\*
- *Strongyloides stercoralis* IgG
- *Entamoeba histolytica* serology

*General/ metabolic screening:*

- Full blood count with differential.
- Creatinine and electrolytes
- Liver enzymes (including albumin, bilirubin, aminotransferases, gamma-glutamyltransferase and alkaline phosphatase).
- C-reactive protein

**Table 3: Recommended stool screening for stool donors:** \*Whilst CPE and ESBL are the only multi-drug resistant bacteria that are recommended to be screened for universally, consider testing for other resistant organisms (including vancomycin-resistant *Enterococci* (VRE) and/ or methicillin-resistant *Staphylococcus aureus* (MRSA)) based upon risk assessment and local prevalence.



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- *Clostridium difficile* PCR
- *Campylobacter*, *Salmonella*, and *Shigella* by standard stool culture and/ or PCR
- Shiga toxin-producing *Escherichia coli* by PCR.
- Multi-drug resistant bacteria, at least carbapenemase-producing *Enterobacteriaceae* (CPE) and extended-spectrum beta-lactamases (ESBL)\*.
- Stool ova, cysts and parasite analysis, including for *Microsporidia*.
- Faecal antigen for *Cryptosporidium* and *Giardia*.
- Acid fast stain for *Cyclospora* and *Isospora*.
- *Helicobacter pylori* faecal antigen.
- Norovirus, Rotavirus PCR.

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2179 **Table 4: A summary of the GRADE system:**

<b>GRADE - strength of evidence:</b>	<b>GRADE - strength of recommendation:</b>
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<i>High quality:</i> Further research is very unlikely to change our confidence in the estimate of effect.	<i>The trade-offs:</i> Taking into account the estimate size of the effect for main outcomes, the confidence limits around those estimates and the relative value placed on each outcome.
<i>Moderate quality:</i> Further research is likely to have an important impact on our confidence in the estimate of effect and may change the estimate.	<i>The quality of the evidence.</i>
<i>Low quality:</i> Further research is very likely to have an important impact on our confidence in the estimate of effect and is likely to change the estimate.	<i>Translation of the evidence into practice in a particular setting:</i> Taking into consideration important factors that could be expected to modify the size of expected effects.
<i>Very low quality:</i> Any estimate of effect is very uncertain.	<i>Uncertainty about the baseline risk for the population of interest.</i>

**Table 5: Criteria for stool collection:**

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Clear instructions should be given to donors regarding hand hygiene.

Collect stool donations in a sealable clean container. A number of specifically designed devices are available commercially.

Stool should ideally be passed directly into the clean container for collection; alternatively, it may be collected in clean tissue and transferred to the clean container.

Stool should be transported to the FMT production site as soon as possible post defaecation (and within six hours); however, if a short period of storage is necessary, this should be at 4°C.

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**The use of faecal microbiota transplant as treatment for recurrent or refractory *Clostridium difficile* infection and other potential indications: joint British Society of Gastroenterology (BSG) and Healthcare Infection Society (HIS) guidelines.**

Benjamin H Mullish<sup>\*1,2</sup>, Mohammed Nabil Quraishi<sup>\*3</sup>, Jonathan Segal<sup>\*1,4</sup>, Victoria L McCune<sup>5,6</sup>, Melissa Baxter<sup>7</sup>, Gemma L Marsden<sup>8</sup>, David Moore<sup>9</sup>, Alaric Colville<sup>7</sup>, Neeraj Bhala<sup>3,9,10</sup>, Tariq H Iqbal<sup>3,10</sup>, Christopher Settle<sup>11</sup>, Graziella Kontkowski<sup>12</sup>, Ailsa L Hart<sup>1,4</sup>, Peter M Hawkey<sup>6</sup>, Simon D Goldenberg<sup>○13,14</sup>, Horace RT Williams<sup>○□1,2</sup>.

1. Division of Integrative Systems Medicine and Digestive Disease, Department of Surgery and Cancer, Faculty of Medicine, Imperial College London, London, UK.
2. Departments of Gastroenterology and Hepatology, St Mary's Hospital, Imperial College Healthcare NHS Trust, Paddington, London, UK.
3. Department of Gastroenterology, Queen Elizabeth Hospital Birmingham, University Hospitals Birmingham NHS Foundation Trust, Birmingham, UK.
4. Inflammatory Bowel Disease Unit, St Mark's Hospital, Harrow, London, UK.
5. Public Health England, Public Health Laboratory Birmingham, Birmingham, UK.
6. Institute of Microbiology and Infection, University of Birmingham, Birmingham, UK.
7. Department of Microbiology, Royal Devon and Exeter NHS Foundation Trust, Exeter, UK.
8. Healthcare Infection Society, London, UK.
9. Institute of Applied Health Research, University of Birmingham, Birmingham, UK.
10. Institute of Translational Medicine, University of Birmingham, Edgbaston, Birmingham, UK.
11. Department of Microbiology, City Hospitals Sunderland NHS Foundation Trust, Sunderland, UK.
12. C diff Support, UK.
13. Centre for Clinical Infection and Diagnostics Research, King's College London, London, UK.
14. Department of Microbiology, Guy's and St Thomas' NHS Foundation Trust, London UK.

\*Joint first authors.

○Joint senior authors.

□Corresponding author.

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32 Contact via: Dr Horace Williams

33 Department of Gastroenterology

34 3<sup>rd</sup> Floor, Salton House

35 St Mary's Hospital, Imperial College Healthcare NHS Trust

36 London, W2 1NY

37 United Kingdom

38 Email: h.williams@imperial.ac.uk

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40 Keywords: microbiota; faecal transplant; *Clostridium difficile*; inflammatory bowel disease

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42 Word count: 16301

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44 Abbreviations: FMT faecal microbiota transplant

45 CDI *Clostridium difficile* infection

46 EBV Epstein-Barr virus

47 CMV cytomegalovirus

48 BMI body mass index

49 GI gastrointestinal

50 RCT randomised controlled trial

51 NAAT nucleic acid amplification test

52 GDH glutamate dehydrogenase

53 EIA enzymes immunoassay

54 PCR polymerase chain reaction

55 IBD inflammatory bowel disease

56 IBS irritable bowel syndrome

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57	HIV	human immunodeficiency virus
58	AIDS	acquired immune deficiency syndrome
59	CPE	carbapenemase-producing <i>Enterobacteriaceae</i>
60	ESBL	extended-spectrum beta-lactamase
61	VRE	vancomycin-resistant <i>Enterococci</i>
62	MRSA	methicillin-resistant <i>Staphylococcus aureus</i>
63	PPI	proton pump inhibitor
64	UC	ulcerative colitis
65	HE	hepatic encephalopathy
66	MELD	Model for End-Stage Liver Disease

## 1. **Abstract:**

Interest in the therapeutic potential of faecal microbiota transplant (FMT) has been increasing globally in recent years, particularly as a result of randomised studies in which it has been used as an intervention. The main focus of these studies has been the treatment of recurrent or refractory *Clostridium difficile* infection (CDI), but there is also an emerging evidence base regarding potential applications in non-CDI settings. The key clinical stakeholders for the provision and governance of FMT services in the United Kingdom (UK) have tended to be in two major specialty areas: gastroenterology and microbiology/infectious diseases. Whilst the National Institute for Health and Care Excellence (NICE) guidance (2014) for use of FMT for recurrent or refractory CDI has become accepted in the UK, clear evidence-based UK guidelines for FMT have been lacking. This resulted in discussions between the British Society of Gastroenterology (BSG) and Healthcare Infection Society (HIS), and a joint BSG/HIS FMT working group was established. This guideline document is the culmination of that joint dialogue.

## 2. **Executive summary:**

### 2.1. **Overview:**

The remit of the British Society of Gastroenterology (BSG)/ Healthcare Infection Society (HIS) working group was to provide recommendations as to best practice in the provision of a faecal microbiota transplant (FMT) service. This guideline considers the use of FMT for the treatment of *Clostridium difficile* infection (CDI) – as well as for potential non-CDI indications – in adults. The working group have primarily targeted their report at clinicians involved in the use and provision of FMT services, but have also aimed it to be of interest to patients and their relatives.

### 2.2. **Summary of recommendations:**

#### 2.2.1. **Which patients with *Clostridium difficile* infection should be considered for faecal microbiota transplant, and how should they be followed up after treatment?**

##### 2.2.1.1. **Prior to faecal microbiota transplant. Patient selection:**

##### 2.2.1.1.1. **Recurrent *Clostridium difficile* infection:**

We recommend that FMT should be offered to patients with recurrent CDI who have had at least two recurrences, or those who have had one recurrence and have risk factors for further episodes, including severe and severe-complicated CDI (*GRADE of evidence: high; strength of recommendation: strong*).



**2.2.1.1.2. Refractory *Clostridium difficile* infection:**

We recommend that FMT should be considered in cases of refractory CDI (*GRADE of evidence: moderate; strength of recommendation: strong*).

**2.2.1.1.3. FMT as initial therapy for *Clostridium difficile* infection:**

We recommend that FMT should not be administered as initial treatment for CDI (*GRADE of evidence: low; strength of recommendation: strong*).

**2.2.1.1.4. Antimicrobial/ antitoxin therapy prior to considering FMT for patients with CDI:**

- i. We recommend that FMT for recurrent CDI should only be considered after recurrence of symptoms following resolution of an episode of CDI that was treated with appropriate antimicrobials for at least 10 days (*GRADE of evidence: low; strength of recommendation: strong*).
- ii. We recommend consideration of treatment with extended/ pulsed vancomycin and/or fidaxomicin before considering FMT as treatment for recurrent CDI (*GRADE of evidence: low; strength of recommendation: strong*).
- iii. For those with severe or complicated CDI, which appears to be associated with reduced cure rates, we recommend that consideration should be given to offering patients treatment with medications which are associated with reduced risk of recurrence (e.g. fidaxomicin and bezlotoxumab), before offering FMT (*GRADE of evidence: low; strength of recommendation: strong*).

**2.2.1.2. Post-FMT follow-up, outcomes and adverse events:**

**2.2.1.2.1. Management of FMT failure:**

We recommend that FMT should be offered after initial FMT failure (*GRADE of evidence: high; strength of recommendation: strong*).

**2.2.1.2.2. General approach to follow-up post-FMT:**

HIS/ BSG FMT Guideline: Main Document, Gut version.

143 We recommend that all FMT recipients should routinely receive follow-up. Clinicians should  
144 follow-up FMT recipients for long enough to fully establish efficacy/adverse events, and for  
145 at least eight weeks in total (*GRADE of evidence: low; strength of recommendation: strong*).

#### 147 **2.2.1.2.3. Management of the FMT recipient:**

148 *i.* We recommend that immediate management after endoscopic administration of  
149 FMT should be as per endoscopy unit protocol (*GRADE of evidence: very low:*  
150 *strength of recommendation: strong*).

151 *ii.* We recommend that patients should be warned about short term adverse events, in  
152 particular the possibility of self-limiting GI symptoms. They should be advised that  
153 serious adverse events are rare (*GRADE of evidence: very low; strength of*  
154 *recommendation: strong*).

155 *iii.* After enteral tube administration, we recommend that patients may have the tube  
156 removed and oral water given from 30 minutes post-administration (*GRADE of*  
157 *evidence: very low; strength of recommendation: strong*).

#### 159 **2.2.1.2.4. Definition of cure post-FMT for CDI:**

160 We recommend that a decision regarding cure/remission from CDI should be recorded  
161 during follow-up. However, this has no uniformly-agreed definition, and should be decided  
162 on a case-by-case basis (*GRADE of evidence: very low; strength of recommendation: strong*).

#### 164 **2.2.1.2.5. Definition of treatment failure post-FMT for CDI:**

165 We recommend that treatment failure/recurrence should be defined on a case-by-case  
166 basis. Routine testing for *C. difficile* toxin after FMT is not recommended, but it is  
167 appropriate to consider in the case of persistent CDI symptoms/suspected relapse (*GRADE*  
168 *of evidence: low; strength of recommendation: strong*).

#### 170 **2.2.2. What recipient factors influence the outcome of faecal microbiota transplant when** 171 **treating people with *Clostridium difficile* infection?**

##### 172 **2.2.2.1. General approach to co-morbidities and FMT:**

- i. We recommend that FMT should be avoided in those with anaphylactic food allergy (GRADE of evidence: very low; strength of recommendation: strong).
- ii. We suggest that FMT should be offered with caution to patients with CDI and decompensated chronic liver disease (GRADE of evidence: very low; strength of recommendation: weak).

**2.2.2.2. Immunosuppression and FMT:**

- i. We recommend that FMT should be offered with caution to immunosuppressed patients, in whom FMT appears efficacious without significant additional adverse effects (GRADE of evidence: moderate; strength of recommendation: strong).
- ii. We recommend that immunosuppressed FMT recipients at risk of severe infection if exposed to EBV or CMV should only receive FMT from donors negative for EBV and CMV (GRADE of evidence: very low; strength of recommendation: strong).

**2.2.2.3. Other comorbidities and FMT:**

- i. We recommend that FMT should be offered to those with recurrent CDI and inflammatory bowel disease, but patients should be counselled about a small but recognised risk of exacerbation of IBD (GRADE of evidence: moderate; strength of recommendation: strong).
- ii. We recommend that FMT should be considered for appropriate patients with recurrent CDI regardless of other comorbidities (GRADE of evidence: moderate; strength of recommendation: strong).

**2.2.3. What donor factors influence the outcome of faecal microbiota transplant when treating people with Clostridium difficile infection?**

**2.2.3.1. General approach to donor selection:**

We recommend that related or unrelated donors should both be considered acceptable. However, where possible, FMT is best sourced from a centralised stool bank, from a healthy unrelated donor (GRADE of evidence: low; strength of recommendation: strong).

**2.2.3.2. Age and BMI restrictions for potential donors:**

HIS/ BSG FMT Guideline: Main Document, Gut version.

204 We suggest that people should only be considered as potential FMT donors if they are  $\geq 18$   
205 and  $\leq 60$  years old, and have a BMI of  $\geq 18$  and  $\leq 30$  kg/m<sup>2</sup> (*GRADE of evidence: low; strength*  
206 *of recommendation: weak*).

### 208 **2.2.3.3. General approach to the donor screening assessment:**

209 It is mandatory to screen potential donors by questionnaire and personal interview, to  
210 establish risk factors for transmissible diseases and factors influencing the gut microbiota  
211 (**Table 1**) (*GRADE of evidence: low; strength of recommendation: strong*).

### 213 **2.2.3.4. Laboratory screening of potential donors:**

214 Blood and stool screening of donors is mandatory (**Tables 2 and 3**) (*GRADE of evidence: low;*  
215 *strength of recommendation: strong*).

### 217 **2.2.3.5. Repeat donor checks, and donation pathway:**

- 218 i. In centres using frozen FMT, before FMT may be used clinically, we recommend that  
219 donors should have successfully completed a donor health questionnaire and laboratory  
220 screening assays both before and after the period of stool donation. This is the  
221 preferred means of donor screening (*GRADE of evidence: low; strength of*  
222 *recommendation: strong*).
- 223 ii. In centres using fresh FMT, we recommend that a repeat health questionnaire should be  
224 assessed at the time of each stool donation. To ensure ongoing suitability for inclusion  
225 as a donor, the donor health questionnaire and laboratory screening should be repeated  
226 regularly (*GRADE of evidence: low; strength of recommendation: strong*).

### 228 **2.2.4. What factors related to the preparation of the transplant influence the outcome of** 229 **faecal microbiota transplant when treating people with *Clostridium difficile*** 230 **infection?**

#### 231 **2.2.4.1. General principles of FMT preparation:**

- i. We recommend that stool collection should follow a standard protocol (GRADE of evidence: low; strength of recommendation: strong).*
- ii. We recommend that donor stool should be processed within 6 hours of defaecation (GRADE of evidence: low; strength of recommendation: strong).*
- iii. We recommend that both aerobically and anaerobically prepared FMT treatments should be considered suitable when preparing FMT for the treatment of recurrent CDI (GRADE of evidence: moderate; strength of recommendation: strong).*
- iv. We recommend that sterile 0.9% saline should be considered as an appropriate diluent for FMT production, and cryoprotectant such as glycerol should be added for frozen FMT (GRADE of evidence: moderate: strength of recommendation: strong).*
- v. We recommend using ≥50g of stool in each FMT preparation (GRADE of evidence: moderate: strength of recommendation: strong).*
- vi. We suggest that stool should be mixed 1:5 with diluent to make the initial faecal emulsion (GRADE of evidence: low; strength of recommendation: weak).*
- vii. We suggest that homogenisation and filtration of FMT should be undertaken in a closed disposable system (GRADE of evidence: low; strength of recommendation: weak).*

**2.2.4.2. Fresh vs frozen FMT:**

*We recommend that the use of banked frozen FMT material should be considered preferable to fresh preparations for CDI (GRADE of evidence: high; strength of recommendation: strong).*

**2.2.4.3. Use of frozen FMT:**

- i. We recommend that FMT material stored frozen at -80°C should be regarded as having a maximum shelf life of six months from preparation (GRADE of evidence: low; strength of recommendation: strong).*
- ii. We suggest consideration of thawing frozen FMT at ambient temperature, and using within six hours of thawing (GRADE of evidence: low; strength of recommendation: weak).*

iii. We suggest not thawing FMT in warm water baths, due to the risks of cross contamination with *Pseudomonas* (and other contaminants) and reduced bacterial viability (*GRADE of evidence: very low; strength of recommendation: weak*).

### **2.2.5. What factors related to administration of the transplant influence the outcome of faecal microbiota transplant when treating people with *Clostridium difficile* infection?**

#### **2.2.5.1. Use of specific medications in the period around FMT administration:**

##### **2.2.5.1.1. General principles of FMT administration:**

- i. We recommended that bowel lavage should be administered prior to FMT via the lower GI route, and that bowel lavage should be considered prior to FMT via the upper GI route; polyethylene glycol preparation is preferred (*GRADE of evidence: low; strength of recommendation: strong*).
- ii. For upper GI FMT administration, we suggest that a proton pump inhibitor should be considered, e.g. the evening before and morning of delivery (*GRADE of evidence: low; strength of recommendation: weak*).
- iii. We suggest that a single dose of loperamide (or other anti-motility drugs) should be considered following lower GI FMT delivery (*GRADE of evidence: low; strength of recommendation: weak*).
- iv. We suggest that prokinetics (such as metoclopramide) should be considered prior to FMT via the upper GI route (*GRADE of evidence: low; strength of recommendation: weak*).
- v. We recommend that best practice for prevention of further transmission of CDI should be applied throughout when administering FMT to patients with CDI (nursing with enteric precautions, sporicidal treatment of endoscope, etc) (*GRADE of evidence: high; strength of recommendation: strong*).

##### **2.2.5.1.2. Additional antibiotics pre-FMT:**

We recommend the administration of further antimicrobial treatment for CDI for at least 72 hours prior to FMT (*GRADE of evidence: low; strength of recommendation: strong*).

**2.2.5.1.3. Washout period between antibiotic use and FMT:**

- i. To minimise any deleterious effect of antimicrobials on the FMT material, we recommend that there should be a minimum washout period of 24 hours between the last dose of antibiotic and treatment with FMT (*GRADE of evidence: low; strength of recommendation: strong*).
- ii. We suggest considering consultation with infectious disease specialists or medical microbiologists for advice whenever FMT recipients also have an indication for long-term antibiotics, or have an indication for non-CDI antibiotics within eight weeks of FMT (*GRADE of evidence: very low; strength of recommendation: weak*).

**2.2.5.2. Route of FMT delivery:**

**2.2.5.2.1. Upper gastrointestinal tract administration of FMT:**

- i. We recommend that upper GI administration of FMT as treatment for recurrent or refractory CDI should be used where clinically appropriate (*GRADE of evidence: high; strength of recommendation: strong*).
- ii. Where upper GI administration is considered most appropriate, we recommend that FMT administration should be via nasogastric, nasoduodenal, or nasojejunal tube, or alternatively via upper GI endoscopy. Administration via a permanent feeding tube is also appropriate (*GRADE of evidence: high; strength of recommendation: strong*).
- iii. We recommend that no more than 100ml of FMT is administered to the upper GI tract (*GRADE of evidence: low; strength of recommendation: strong*).
- iv. We recommend that upper GI administration of FMT should be used with caution in those at risk of regurgitation and/ or those with swallowing disorders (*GRADE of evidence: low; strength of recommendation: strong*).

**2.2.5.2.2. Lower gastrointestinal tract administration of FMT:**

- i. We recommend that colonoscopic administration of FMT as treatment for recurrent or refractory CDI should be used where appropriate (*GRADE of evidence: high; strength of recommendation: strong*).



ii. Where colonoscopic administration is used, we suggest considering preferential delivery to the caecum or terminal ileum, as this appears to give the highest efficacy rate (*GRADE of evidence: low; strength of recommendation: weak*).

iii. We recommend that FMT via enema should be used as a lower GI option when delivery using colonoscopy or flexible sigmoidoscopy is not possible (*GRADE of evidence: high; strength of recommendation: strong*).

#### 2.2.5.2.3. Capsulised FMT:

Capsulised FMT holds promise as a treatment option for recurrent CDI and we recommend that this should be offered to patients as a potential treatment modality where available. Capsule preparations should follow a standard protocol. Further evidence regarding optimal dosing and formulation is required (*GRADE of evidence: high; strength of recommendation: strong*).

#### 2.2.6. What is the clinical effectiveness of FMT in treating conditions other than *Clostridium difficile* infection?

We do not currently recommended FMT as treatment for inflammatory bowel disease. Apart from CDI, there is insufficient evidence to recommend FMT for any other gastrointestinal or non-gastrointestinal disease (*GRADE of evidence: moderate; strength of recommendation: strong*).

#### 2.2.7. Basic requirements for implementing a FMT service:

##### 2.2.7.1. General considerations:

i. The development of FMT centres should be encouraged (*GRADE of evidence: very low; strength of recommendation: strong*).

ii. We suggest that FMT centres should work to raise awareness about FMT as a treatment option amongst clinicians caring for patients with CDI, and provide training to relevant healthcare professionals on the practicalities of delivering an FMT service (*GRADE of evidence: very low; strength of recommendation: weak*).

**2.2.7.2. Legal aspects and clinical governance:**

In the UK, FMT must be manufactured in accordance with MHRA guidance for human medicines regulation. When FMT is supplied on a named patient basis, within a single organisation, a pharmacy exemption may be used, subject to ensuring proper governance and traceability. All centres that are processing and distributing FMT should seek guidance from the MHRA and where necessary obtain appropriate licenses prior to establishing an FMT service. This is a legal requirement. In countries other than the UK, FMT should only be manufactured following appropriate approval from the national authority of that country (*GRADE of evidence: very low; strength of recommendation: strong*).

**2.2.7.3. Multidisciplinary teams:**

We recommend that a multidisciplinary team should be formed to deliver FMT services (*GRADE of evidence: very low; strength of recommendation: strong*).

**2.2.7.4. Infrastructure:**

We recommend utilisation of suitable laboratory facilities and infrastructure for FMT production (*GRADE of evidence: very low; strength of recommendation: strong*).

**2.2.7.5. FMT manufacturing:**

We recommend ensuring the traceability of supply (*GRADE of evidence: very low; strength of recommendation: strong*).

**2.2.7.6. FMT production quality control:**

We recommend monitoring, notification and investigation of all adverse events and reactions related to FMT (*GRADE of evidence: very low; strength of recommendation: strong*).

**2.2.7.7. Donor screening governance:**

We recommend ensuring the clinical governance of donor screening (*GRADE of evidence: very low; strength of recommendation: strong*).

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**3. Introduction:**

The aim of the BSG/ HIS FMT working group was to establish a guideline that defined best practice in all aspects of a FMT service, by providing evidence-based recommendations wherever possible, and consensus multi-disciplinary expert opinion where specific published evidence is currently lacking. This included the evaluation of the use of FMT in the treatment of *Clostridium difficile* infection (CDI; also referred to as *Clostridioides difficile*<sup>1</sup>), and also in potential non-CDI indications. Relevant guidance published to date includes the interventional procedure guidance from the National Institute for Health and Care Excellence (NICE)<sup>2</sup>, UK, European and US microbiological guidelines on the treatment of *Clostridium difficile* infection (CDI)<sup>3–5</sup>, and recent expert consensus documents on FMT in clinical practice<sup>6,7</sup>. Furthermore, there have also been national recommendations regarding FMT produced by working groups in several different countries<sup>8–10</sup>. Principally as a result of randomised studies that have been published in recent years<sup>11–18</sup>, FMT has become an accepted treatment for recurrent/refractory CDI.

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The unique remit and objectives of this guideline when commissioned by the BSG and HIS was:

i. To review the rapidly-growing body of randomised trial evidence for the efficacy of FMT in the treatment of adults (≥18 years), both in CDI and in other clinical conditions, much of which has been published after the publication of current CDI treatment algorithms<sup>3,4</sup>.

ii. To provide specific guidance about best practice for an FMT service within the context of the regulatory framework for the intervention as it currently exists in the UK<sup>19,20</sup>.

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The elucidation of the mechanisms underlying the efficacy of FMT in treating CDI remains an active area of global research, with the aim of rationalising FMT from its current crude form to a more targeted, refined therapeutic modality<sup>21</sup>. Previous research has demonstrated that commensal bacteria cultured from the stool of healthy donors<sup>22</sup>, sterile faecal filtrate<sup>23</sup>, and/ or spores of *Firmicutes* derived from ethanol-treated stool from healthy donors<sup>24</sup>, may have similar efficacy to conventional FMT in treating CDI, although results of the latter approach produced disappointing outcome data when extended to a Phase II clinical trial<sup>25</sup>. For the purposes of this guideline, the BSG/HIS working group considered only studies that used the administration of manipulated whole stool (including encapsulated faeces). They deemed studies using cultured microorganisms (or their

proteins, metabolites or other components), or microbiota suspensions, to be in the pre-clinical research stage, without firm evidence.

FMT has been shown to be very acceptable to patients, both in the setting of CDI<sup>11,26</sup> and in non-CDI settings, e.g. ulcerative colitis<sup>27</sup>. However, the absence of appropriate protocols<sup>28–31</sup> specifically taking into account UK clinical practice and regulation of FMT has been perceived as a barrier to the use of FMT in the UK and Ireland; these guidelines seek to rectify this problem.

**4. Guideline development:**

**4.1. Guideline development team**

BSG and HIS commissioned the authors to undertake the Working Party Report. The authors represent the membership of both societies. The working group included gastroenterologists, infectious diseases/microbiology clinicians, a clinical scientist, a systematic reviewer, and patient representatives. The views expressed in this publication are those of the authors, and have been endorsed by BSG and HIS following consultation.

**4.2. Scope of the guidelines**

The main scope of the guidelines is to provide guidance for the optimal provision of an effective and safe FMT service, principally for recurrent or refractory CDI, but non-CDI indications are also considered. These guidelines only apply to adult patients (≥18 years); the working party did not consider the role of FMT in the treatment of either CDI or non-CDI indications in children or young people. The guidelines were written with a focus upon UK practice, but also with consideration of more global practice as it applied. The diagnosis and management of *Clostridium difficile* infection in general are outside the remit of these guidelines.

**4.3. Evidence appraisal**

Questions for review were derived from the Working Party Group, which included patient representatives in accordance with the PICO process<sup>32</sup>. To prepare these recommendations, the working group collectively reviewed relevant peer-reviewed research.

#### 4.4. Data sources and search strategy

A systematic literature search was undertaken using MEDLINE, EMBASE databases and Cochrane Library for relevant articles published from 1<sup>st</sup> January 1980 to 1<sup>st</sup> January 2018. The MEDLINE and EMBASE strategy are shown in **Supplementary Material 1, Appendix 2ii**. Free text and MESH/ index terms for faecal microbial transplant and *Clostridium difficile* or other diseases of interest were combined. In addition, conference proceedings from microbiology, infectious disease, and gastroenterology conferences were also searched to identify additional studies.

#### 4.5. Study eligibility and selection criteria

The members of the guideline group determined criteria for study inclusion. Two reviewers (BHM, MNQ) screened the titles and abstracts of each article for relevance independently; any disagreements were resolved by discussion with a third reviewer (JPS). Copies of relevant articles were obtained and assessed for inclusion as evidence in the guideline by all three reviewers. The reason for not selecting studies was recorded. Only articles published in English and human clinical studies were included. For evidence on FMT for CDI, both randomised studies (including randomised controlled trials (RCTs)) and case series with at least 10 patients were selected. Only randomised trials were included as evidence for FMT for non-CDI indications. Conference abstracts were only included for CDI and non-CDI indications if they reported a randomised trial; where abstracts were available reporting data from a randomised trial that was subsequently published, only the published paper was reviewed.

#### 4.6. Data extraction and quality assessment

The initial search identified 2658 publications, and of these, 802 duplicates were excluded. 1856 studies were subsequently screened, from which 78 studies were assessed by reviewing the full text for eligibility (see **Supplementary Material 1, Appendix 2iii** and **Supplementary Material 2, Additional Appendix D**). Of these 78 studies, 58 studies were included as the basis of evidence for writing this guideline. In total, 39 were case studies in CDI including at least 10 patients (see **Supplementary Material 2, Additional Appendix C.1**), and ten were randomised studies in CDI (see **Supplementary Material 2, Additional Appendix C.2**). Nine were randomised trials for non-CDI indications (see **Supplementary Material 2, Additional Appendix C.3**). Data were extracted for patient demographics, disease characteristics, donor screening characteristics, stool preparation and administration, clinical outcomes and adverse events. The quality of randomised studies was

assessed with the Cochrane Collaboration’s risk of bias tool. Case series were assessed using the Centre for Reviews and Dissemination guidance.

**4.7. Rating of evidence and recommendations**

The BSG version of these guidelines was prepared in keeping with the BSG Clinical Services & Standards Committee (CSSC) advice document on the writing of clinical guidelines<sup>33</sup>. Evidence tables were presented and discussed by the working group, and guidelines were prepared according to the nature and applicability of the evidence regarding efficacy and patient preference and acceptability. For the BSG version of this guideline, the GRADE system (Grades of Recommendation Assessment, Development and Evaluation)<sup>34</sup> was used to assess the strength of evidence (high/ moderate/ low/ very low) and strength of recommendation (strong/ weak) (**Table 4**). The section entitled ‘Basic requirements for implementing an FMT service’ (**Supplementary Material 3**) was based on expert opinion, since this was a key area of the working party’s remit but not one amenable to evaluation by the PICO process. Face-to-face meetings and group teleconferences were held to agree on recommendations. Any disagreements on recommendations or the strength of recommendation were resolved by discussion and, where necessary, voting by the members of the working group, with consensus achieved when >80% were in agreement.

**4.8. Consultation process**

Feedback on draft guidelines was received from the Scientific Development Committee (SDC) of HIS, and changes made. These guidelines were then opened to consultation with relevant stakeholders (see **Supplementary Material 1, Appendix 3** of this document). The draft report was available on the HIS website for one month. Views were invited on format, content, local applicability, patient acceptability, and recommendations. The working group reviewed stakeholder comments, and collectively agreed revisions. Final changes were made after repeat reviews from HIS (Chair of the SDC and HIS Council) and BSG (BSG CSSC and BSG Council), and after further external peer review.

**4.9. Guideline accreditation and scheduled review**

The guidelines will be reviewed at least every four years and updated if change(s) in the evidence are sufficient to require a change in practice.

#### 4.0. Additional information:

Additional information related to this guideline (including a lay summary, background on the working party report, and information on the implementation of these guidelines) is contained within **Supplementary Material 1, Section 1**.

### 5. Rationale for recommendations:

#### 5.1. Which patients with *Clostridium difficile* infection should be considered for faecal microbiota transplant, and how should they be followed up after treatment?

##### 5.1.1. Prior to faecal microbiota transplant. Patient selection:

##### 5.1.1.1. Recurrent *Clostridium difficile* infection:

As already described, there is widespread consensus that FMT is an efficacious treatment for recurrent CDI. In defining recurrent CDI, some studies have relied on a minimum threshold of return of clinical symptoms (e.g. at least three unformed bowel movements within 24 hours, for at least two consecutive days)<sup>12,18</sup> following previous successful CDI treatment; most studies have also included a requirement for a positive microbiological test<sup>12,14,18,35–45</sup>. Other studies explicitly state that a positive test was not required<sup>46</sup>. Recommendations for CDI testing are beyond the scope of this guideline, and there are already well-established evidence-based guidelines<sup>47</sup>. These recommend testing with either a nucleic acid amplification test (NAAT) or GDH assay, followed by detection of free toxin (either by toxin A/B enzyme immunoassay (EIA) or cytotoxin neutralisation assay), which allows differentiation of patients with active disease as well as those who are likely colonised<sup>47</sup>. However, the working group discussed the importance of the accurate diagnosis of true recurrent CDI prior to consideration of FMT; in particular, they noted a study which observed that of 117 patients with presumed recurrent CDI referred for work-up for FMT, 25% ( $n=29/117$ ) were determined to have a non-CDI diagnosis, with irritable bowel syndrome ( $n=18$ ) and inflammatory bowel disease ( $n=3$ ) being the most common alternative diagnoses, and younger patients more likely to be misdiagnosed<sup>48</sup>.

All of the reviewed studies have included patients with recurrent CDI, however some studies offered FMT to patients at the first recurrence (second episode)<sup>12,15,16,18,35,37,42,43,46,49</sup>, whereas others offered FMT after the second recurrence (third episode)<sup>13,14,39,41,44,45,50,51</sup>. Some protocols offered FMT after three or more recurrences<sup>52</sup>, whilst others did not define the point at which it was administered<sup>40,53</sup>.



The severity of infection has been used as a parameter to decide at which stage FMT is offered. Youngster *et al.* offered FMT to patients with at least three episodes of mild to moderate CDI, or at least two episodes of severe CDI resulting in hospitalisation and associated with significant morbidity<sup>17</sup>. Another study selected patients for FMT using four categories of severity, which also accounted for prior anti-CDI therapy and requirement for hospitalisation<sup>54</sup>.

None of the studies directly compared the efficacy of FMT according to the stage at which it was offered (i.e. first recurrence vs.  $\geq$  two recurrences). A small number of studies<sup>55-57</sup> included patients with severe CDI (defined as hypoalbuminaemia with increased peripheral white cell count and/or abdominal tenderness) or complicated CDI (defined as admission to Intensive Care, altered mental status, hypotension, fever, ileus, white blood cell count  $> 30 \times 10^9/l$ , lactate  $> 2.2\text{mmol/l}$ , or evidence of end organ damage). A single study described an apparent lower rate of treatment success when FMT was used to treat patients with recurrent CDI with disease caused by ribotype 027<sup>43</sup>, but this is the case for all anti-CDI treatment modalities for this ribotype in comparison to others. The working group agreed that there was insufficient evidence to suggest that *C. difficile* ribotype should influence whether or not FMT is offered.

A lower primary cure rate was reported for complicated CDI (66%) compared with recurrent CDI (82%) and severe CDI (91%) in one study<sup>55</sup>; in a case series of 17 patients who all had severe and/or complicated CDI, a primary cure rate of 88% was described<sup>57</sup>. A cohort of 328 patients was analysed to determine which factors were associated with failure of FMT<sup>58</sup>. Higher early (one month) failure rates were found in patients with severe (72%,  $n=19/25$ ) or severe-complicated (52.9%,  $n=9/17$ ) CDI than for recurrent CDI (11.9%,  $n=34/286$ ). This study also identified that patients who were treated with FMT as an inpatient were nearly four times more likely to fail as those who had FMT as an outpatient; however, the working group noted that the authors of this study themselves identified that inpatient status is likely a proxy of severity of CDI and/or co-morbidities. A further similar study, including 64 patients treated with FMT as treatment for recurrent CDI, also identified severe CDI as the strongest independent risk factor for FMT failure on multivariate analysis<sup>59</sup>.

The working group discussed their experience of treating patients with CDI whose disease fitted an intermediate pattern to the typical descriptions given of recurrent or refractory CDI, e.g. patients with CDI who have some (but incomplete) symptomatic improvement with anti-CDI antibiotics and worsening of disease when these are stopped. The experience of the working group was that such

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571 patients experienced excellent responses to FMT, and that these patients should be considered for  
572 FMT.

573

574 As FMT is currently an unlicensed medicine with poorly-studied long term sequelae, the working  
575 group considered that it should generally be reserved for patients who have had three or more  
576 episodes of infection. There are no studies directly comparing its effectiveness with some of the  
577 newer agents such as fidaxomicin or bezlotoxumab, hence this recommendation is made on the  
578 basis of safety. However, the working group agreed that it may be reasonable in certain patient  
579 groups with ongoing risk factors for further recurrence to offer FMT after the second episode.

580

581 **Recommendation:**

582 **We recommend that FMT should be offered to patients with recurrent CDI who have had**  
583 **at least two recurrences, or those who have had one recurrence and have risk factors for**  
584 **further episodes, including severe and severe-complicated CDI (GRADE of evidence: high;**  
585 **strength of recommendation: strong).**

586

587 **5.1.1.2. Refractory *Clostridium difficile* infection:**

588 Two randomised trials allowed the recruitment of patients with refractory CDI. The first defined this  
589 as at least three weeks of ongoing severe symptoms despite standard antimicrobial therapy for  
590 CDI<sup>17</sup>. The second required persistent or worsening diarrhoea and one of the following: ongoing  
591 abdominal pain, fever > 38°C, or white blood cell count > 15x 10<sup>9</sup>/l despite oral vancomycin at a dose  
592 of 500mg four times daily for at least five days<sup>16</sup>. Both studies included only small numbers of  
593 patients with refractory CDI ( $n=4/20$  (20%) and  $n=15/219$  (6.8%), respectively). There did not appear  
594 to be any significant difference in primary outcome measure (clinical cure) in patients with recurrent  
595 or refractory CDI, although neither study was designed to assess this difference. There are also a  
596 number of case series in which FMT was given to patients with refractory CDI; however, outcome  
597 measures were not reported for these groups individually in these studies<sup>37,38,54,60</sup>.

598

599 Overall, the working group concluded that there is little consensus on the definition of refractory  
600 CDI, with some studies using the terms 'refractory' and 'recurrent' interchangeably (as well as other  
601 terms, e.g. 'salvage therapy'). Consequently, the quality of evidence for the utility of FMT in

refractory cases of CDI is lower than for recurrent CDI. The standardisation of definitions will allow more robust comparison between patient cohorts.

**Recommendation:**

**We recommend that FMT should be considered in cases of refractory CDI (GRADE of evidence: moderate; strength of recommendation: strong).**

**5.1.1.3. FMT as initial therapy for *Clostridium difficile* infection:**

Experience of the use of FMT as initial therapy for CDI is very limited. In a case series of patients with CDI with ribotype 027, use of anti-CDI antibiotics together with nasogastric FMT within a week of diagnosis during an initial episode of CDI was associated with reduced mortality when compared to using FMT only after the failure of three courses of antibiotics (mortality of 18.75% ( $n=3/16$  patients) vs 64.4% ( $n=29/45$  patients))<sup>61</sup>. However, 37.5% ( $n=6/16$ ) of the patients treated with FMT within a week of CDI diagnosis required further antibiotics and a second FMT within one month of the first FMT because of relapse<sup>61</sup>. In a small pilot randomised trial, patients were randomised to either vancomycin or multi-donor FMT (administered either via upper or lower GI routes) as initial therapy for CDI; CDI resolution occurred in 88.9% ( $n=8/9$ ) patients with vancomycin, compared to 57.1% of patients ( $n=4/7$ ) patients with one FMT, and 71.4% of patients ( $n=5/7$ ) after two FMTs<sup>62</sup>. Given the small size of these studies and equivocal results, the working group concluded that the reviewed studies did not support FMT as initial therapy for CDI.

**Recommendation:**

**We recommend that FMT should not be administered as initial treatment for CDI (GRADE of evidence: low; strength of recommendation: strong).**

**5.1.1.4. Antimicrobial/ antitoxin therapy prior to considering FMT for patients with CDI:**

There are now at least two licensed agents (fidaxomicin and bezlotoxumab) which have been shown to significantly reduce the risk of recurrence compared with vancomycin<sup>63,64</sup>. There is also some evidence that pulsed/tapered dosing of vancomycin and fidaxomicin (including pulsed fidaxomicin<sup>65</sup>) results in fewer recurrences than with standard dosing of these agents<sup>66,67</sup> (although this finding has

not been replicated in all studies<sup>68</sup>). Pre-planned subgroup analysis of patients with severe CDI in a randomised trial demonstrated a significantly lower recurrence rate when treated with fidaxomicin (13.0%,  $n=12/92$ ) than when treated with vancomycin (26.6%,  $n=29/209$ )<sup>63</sup>; this finding was replicated in another randomised controlled trial, with 8.3% ( $n=4/48$ ) and 32.6% ( $n=14/43$ ) experiencing a recurrence respectively<sup>69</sup>. In a further randomised trial, bezlotoxumab (together with standard of care antibiotics) was shown to reduce recurrence of severe CDI compared to standard of care antibiotics alone (10.9% ( $n=6/55$ ) vs 20% ( $n=13/65$ ) respectively)<sup>64</sup>.

As discussed above, the working group noted that there are no studies comparing FMT to fidaxomicin or bezlotoxumab, and only one study comparing a vancomycin taper to FMT<sup>12</sup>. The working group agreed that in the absence of this evidence, on the balance of safety and potential risks, consideration should be given to using antimicrobial/antitoxin therapy associated with reduced CDI recurrence prior to considering the use of FMT.

Several studies specify that patients should be treated with anti-*C. difficile* antibiotics for a minimum period of 10 days before diagnosing recurrent CDI and offering FMT<sup>12,15,16,18</sup>.

#### **Recommendations:**

- i. **We recommend that FMT for recurrent CDI should only be considered after recurrence of symptoms following resolution of an episode of CDI that was treated with appropriate antimicrobials for at least 10 days (GRADE of evidence: low; strength of recommendation: strong).**
- ii. **We recommend consideration of treatment with extended/ pulsed vancomycin and/or fidaxomicin before considering FMT as treatment for recurrent CDI (GRADE of evidence: low; strength of recommendation: strong).**
- iii. **For those with severe or complicated CDI, which appears to be associated with reduced cure rates, we recommend that consideration should be given to offering patients treatment with medications which are associated with reduced risk of recurrence (e.g. fidaxomicin and bezlotoxumab), before offering FMT (GRADE of evidence: low; strength of recommendation: strong).**

#### **5.1.2. Post-FMT follow-up, outcomes and adverse events:**

##### **5.1.2.1. Management of FMT failure:**

Where patients were deemed not to have responded to an initial FMT, many studies have offered repeat FMT and success rates have been excellent even in patients with modest response to a first FMT<sup>14,15,17,18,35,43,46,51,54,70,71</sup>. The success of a second FMT appears to be high whether treatment failure represents non-response to the first FMT, or a late failure (i.e. further relapse of CDI after an initial response); however, these terms have been defined variably between different studies (also see **Section 5.1.2.5**). Second FMTs have been offered as soon as 24-72 hours after an initial FMT for presumed non-response<sup>37,72,73</sup>. For FMT failure in patients with pseudomembranous colitis, repeat FMT every three days until resolution of pseudomembranes has been a successful approach<sup>18</sup>. Good outcomes in pseudomembranous disease have also been achieved through a protocol that routinely restarted five days of vancomycin if FMT failed, before offering another FMT<sup>73</sup>. Other studies have demonstrated potential success in treating initial FMT failure with further antibiotics, including repeat FMT with vancomycin between procedures<sup>42</sup>, or anti-CDI antibiotics alone<sup>35,42,43,45,51,70,71</sup>. Patients unresponsive to two FMTs have been offered further FMT or antibiotic therapy<sup>16</sup>, or even the administration of intravenous immunoglobulin<sup>35</sup>. Whilst the working group collectively agreed that there was strong evidence to recommend repeat FMT after initial FMT failure, they were not able to recommend a specific protocol for administering repeat FMT and/ or maximum number of FMTs, given the wide heterogeneity of approach described within the reviewed literature.

**Recommendation:**

**We recommend that FMT should be offered after initial FMT failure (GRADE of evidence: high; strength of recommendation: strong).**

**5.1.2.2. General approach to follow-up post-FMT:**

Follow-up post-FMT (in terms of duration, modality and regimen for follow-up) varies considerably between studies, and is largely dependent upon study design. Follow-up regimens vary not only between studies but within them too, reflecting the retrospective nature of many early FMT studies in CDI, where follow-up mostly reflected pragmatic routine clinical care.

Modalities of follow-up have included outpatient review<sup>14,43,58,71,74-76</sup>, telephone interview<sup>17,39,43,46,58,71,74</sup> and case note/ database review<sup>35,39,70,71,74,40,42,43,45,46,49,51,54</sup>. Follow-up duration has varied from 60 days<sup>45</sup> to 8 years<sup>36</sup>, with very different durations used in each study. Once again, however, this variability in follow-up largely reflects the retrospective analysis of case

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series rather than being justified by any specific methodology. The working group decided by consensus that at least eight weeks of follow-up was appropriate post-FMT to fully assess efficacy and potential adverse events; this figure was also influenced by discussions regarding the timepoint after FMT at which a decision could be made regarding cure/ remission of CDI (see **Section 5.1.2.4**).

**Recommendation:**

**We recommend that all FMT recipients should routinely receive follow-up. Clinicians should follow-up FMT recipients for long enough to fully establish efficacy/adverse events, and for at least eight weeks in total (GRADE of evidence: low; strength of recommendation: strong).**

**5.1.2.3. Management of the FMT recipient:**

Procedural adverse events during administration of FMT have predominantly occurred with colonoscopic administration of FMT. These have included mild nausea and vomiting attributed to sedation for the colonoscopy, minor mucosal tears during colonoscopy<sup>49,60</sup>, and microperforation following biopsy of an area of presumed ischaemic small bowel injury in a patient with chronically dilated small bowel (which resolved with conservative management<sup>46</sup>). One death occurred due to witnessed aspiration at the time of colonoscopy<sup>60</sup>. Faecal regurgitation and vomiting with temporal association to upper GI FMT administration has also been described (discussed further in **Section 5.5.2.2**)<sup>77</sup>.

The predominant short term adverse events post-FMT for CDI are mild: self-limiting GI symptoms have been the most frequently reported adverse events. These may be related to the route of administration and include belching<sup>15</sup>, nausea<sup>15,16,49,60</sup>, abdominal cramps/ discomfort/ bloating/ pain<sup>15,18,49,60,72</sup>, and diarrhoea<sup>15,16,18,60</sup>. One patient with a history of autonomic dysfunction experienced dizziness with diarrhoea after FMT<sup>15</sup>. These symptoms are typically short-lived, resolving in hours to days<sup>15,16,18,49,72</sup>. Minor subsequent adverse events have included a range of GI side effects including self-limiting abdominal discomfort<sup>14,17,57,76</sup>, nausea<sup>14,49,70</sup>, flatulence<sup>14,16,17,41,42,49,57</sup>, self-limiting irregular bowel movements<sup>41</sup>, *C. difficile*-toxin negative diarrhoea<sup>52,55</sup>, constipation<sup>14,15,42,55,70</sup> and constitutional symptoms/ temperature disturbance<sup>14,17</sup>.

As such, immediately post-endoscopic administration of FMT, most FMT centres typically manage patients using standard protocols for an endoscopic procedure<sup>41,49</sup>, without any specific adaptations (apart from to reiterate advice about the possibility of self-limiting GI side effects, and the use of departmental infection control protocols). There is often a relatively short period of post-procedural observation<sup>15,18</sup>. Most studies allow patients to leave the administration site after the period of observation, although overnight observation was the protocol used for a cohort of very elderly patients with multiple comorbidities<sup>51</sup>. Where enteral tube administration is used, post-procedure management has ranged between removal of the tube after 30 minutes (following nasoenteral administration of 500ml of FMT<sup>15</sup>) to prompt post-procedure removal and oral water administration (after nasogastric administration of 90ml of FMT<sup>72</sup>), with no direct adverse outcomes in either case. The working group felt that removal of the tube at 30 minutes, with administration of water at this point, was a pragmatic approach.

The definition of post-FMT serious adverse events has varied between studies, but has included significant morbidity necessitating hospital admission and death in the follow up period. Many of these events are described as not directly caused by the FMT, including the scenario of post-FMT severe CDI recurrences<sup>72</sup> and probable or certain CDI-related deaths<sup>16,60,70</sup> occurring in the context of FMT failure, or deaths related to patient comorbidities<sup>17,55</sup>. One patient was admitted to hospital with self-limiting abdominal pain post-FMT<sup>60</sup>, and four patients with flares of inflammatory bowel disease<sup>60</sup>. Three patients underwent colectomy during the post-FMT follow-up period, with all related to ulcerative colitis and not believed to be due to CDI<sup>60</sup>. Other reported serious adverse events include recurrent urinary tract infection<sup>15</sup>, fever during haemodialysis<sup>15</sup> and upper gastrointestinal haemorrhage after nasogastric FMT (in a patient taking NSAIDs<sup>51</sup>), none of which were thought to be strongly linked to FMT. There have also been a number of new onset autoimmune, inflammatory and metabolic conditions described post-FMT, although these have been described from single centres only, with these findings not replicated elsewhere. Such conditions include microscopic colitis, Sjögren's syndrome, follicular lymphoma, peripheral neuropathy, immune thrombocytopenia and rheumatoid arthritis<sup>53,55</sup>.

Significant adverse events are therefore rare but well-described. Furthermore, the procedure is relatively novel, and longer-term follow-up data regarding safety are required. Therefore, the working group opined that formal follow-up post-FMT to assess outcome and possible adverse events is essential.



The use of questionnaires to compare symptoms pre- and post-FMT is common. Specifically, data collected have included clinical response to symptom severity<sup>55</sup>, stool frequency<sup>15,17,46,55,57,72</sup>, stool consistency<sup>14,15,72</sup>, abdominal pain or tenderness<sup>55,57</sup>, rating of gastrointestinal symptoms<sup>72</sup>, general well-being<sup>55,72</sup>, days to improvement post-FMT<sup>57</sup>, weight change<sup>72</sup>, functional status<sup>55</sup>, and changes in medication/use of antibiotics<sup>57,72</sup>. Additionally, certain patients have been given specific advice post-FMT to contact their clinical team if there is recurrence of diarrhoea or symptoms<sup>14,35,41,43</sup>. Where patients underwent outpatient clinical evaluation, this was generally undertaken relatively early post-FMT<sup>39,52,76</sup>. In one study, patients were additionally given instructions for cleaning and disinfection at home, with the aim of reducing the possibility of *C. difficile* reinfection<sup>43</sup>, and counselling on the risk of recurrent CDI with future antibiotic courses<sup>76</sup>.

#### **Recommendations:**

- i. **We recommend that immediate management after endoscopic administration of FMT should be as per endoscopy unit protocol (GRADE of evidence: very low; strength of recommendation: strong).**
- ii. **We recommend that patients should be warned about short term adverse events, in particular the possibility of self-limiting GI symptoms. They should be advised that serious adverse events are rare (GRADE of evidence: very low; strength of recommendation: strong).**
- iii. **After enteral tube administration, we recommend that patients may have the tube removed and oral water given from 30 minutes post-administration (GRADE of evidence: very low; strength of recommendation: strong).**

#### **5.1.2.4. Definition of cure post-FMT for CDI:**

It is recognised that symptoms of CDI resolve relatively promptly post-successful FMT, although this has been variably described (within hours in some studies<sup>52</sup>, at an average of 4-5 days in others<sup>57,71</sup>). Treatment success post-FMT for CDI has no uniformly-agreed definition, with the time point at which cure/ remission is defined on clinical grounds varying between 3-5 days<sup>36</sup> up to six months<sup>42</sup>. A consensus document from the USA recommends 'resolution of symptoms as a primary end point; absence within eight weeks of FMT as a secondary end point'<sup>78</sup>. The working group recommended that this definition should be made on a case-by-case basis; however, they agreed that an

assessment for cure/ remission of CDI within eight weeks post-FMT was reasonable in most cases, and therefore that this was also a reasonable minimum length of time to undertake follow-up post-FMT (see **Section 5.1.2.2**).

**Recommendation:**

**We recommend that a decision regarding cure/remission from CDI should be recorded during follow-up. However, this has no uniformly-agreed definition, and should be decided on a case-by-case basis (GRADE of evidence: very low; strength of recommendation: strong).**

**5.1.2.5. Definition of treatment failure post-FMT for CDI:**

There is no uniformly-agreed definition of treatment failure/recurrence post-FMT for CDI, with varied definitions used in studies. The use of *C. difficile* toxin as a marker of treatment success or failure is variable, with some studies opting not to test for CDT unless symptoms consistent with CDI recurred<sup>49,52–54,60,72,74</sup>. Some studies have routinely performed CDT testing without specifying any action taken after a positive result<sup>14,15,18,36,39,41</sup>, whilst others have tested for *C. difficile* PCR but relied on clinical criteria (even if PCR was positive) post-FMT for evaluating FMT efficacy<sup>14</sup>. A recent prospective study from the USA identified that only 3% (3/129) of patients who were asymptomatic at four weeks post-FMT for recurrent CDI had positive *C. difficile* PCR, again emphasising that symptoms rather than laboratory assays are more useful contributors to establishing FMT success<sup>79</sup>.

**Recommendation:**

**We recommend that treatment failure/recurrence should be defined on a case-by-case basis. Routine testing for C. difficile toxin after FMT is not recommended, but it is appropriate to consider in the case of persistent CDI symptoms/suspected relapse (GRADE of evidence: low; strength of recommendation: strong).**

**5.2. What recipient factors influence the outcome of faecal microbiota transplant when treating people with Clostridium difficile infection?**

**5.2.1. General approach to co-morbidities and FMT:**

Most published studies had a core set of general recipient exclusions which included: significant/ anaphylactic food allergy<sup>14,17</sup>, pregnancy<sup>12–15,17,18</sup>, breastfeeding<sup>14</sup>, admission to Intensive Care or the

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requirement for vasopressors<sup>12,15,18</sup>, chronic diarrhoea or other infectious cause of diarrhoea<sup>12,14,18,50</sup>, inflammatory bowel disease (IBD) and irritable bowel syndrome (IBS)<sup>14,36</sup>, immunodeficiency due to recent chemotherapy and/ or neutropenia<sup>12,14–18,50</sup>, HIV/AIDS<sup>14,17,18</sup>, prolonged use of corticosteroids<sup>15,17,18</sup>, graft versus host disease<sup>12</sup>, and decompensated cirrhosis<sup>14,15,17,18</sup>.

The working group discussed the reported practice of several centres of treating patients with recurrent CDI and food allergies through the use of FMT prepared from a patient-directed donor instructed to avoid trigger foods before stool donation. They agreed that this seemed reasonable for patients with true adverse immunological reactions to defined food groups (e.g. gluten-free diet donor for a recipient with coeliac disease). However, the working group noted that food allergies are often poorly-defined clinically, and also expressed concerns that there was no means to verify how closely a donor had followed an exclusion diet; as such, they felt unable to make any specific recommendation about FMT in patients with food allergies in general. In contrast, whilst the working group were unaware of any reports in the literature of anaphylaxis attributable to FMT, they felt that the theoretical risk of a serious adverse outcome in patients with anaphylactic food allergy merited a specific recommendation that such individuals should not be offered FMT. Similarly, the working group expressed concern about the theoretical risk of adverse outcomes when administering FMT to patients with advanced decompensated chronic liver disease (including translocation of microbial material from the intestinal tract into the portal and systemic circulations, and theoretical risk of sepsis), and felt that FMT should be used with caution in this patient group.

#### **Recommendations:**

- i. **We recommend that FMT should be avoided in those with anaphylactic food allergy (GRADE of evidence: very low; strength of recommendation: strong).**
- ii. **We suggest that FMT should be offered with caution to patients with CDI and decompensated chronic liver disease (GRADE of evidence: very low; strength of recommendation: weak).**

#### **5.2.2. Immunosuppression and FMT:**

One randomised study<sup>16</sup> included patients with immunodeficiency (treatment with immunosuppressive therapy (azathioprine, ciclosporin, infliximab, methotrexate alone, or in combination with corticosteroids) ( $n=18$ ), renal transplant ( $n=5$ ), chronic haemodialysis ( $n=5$ ), solid organ tumours ( $n=3$ ) and haematological malignancy ( $n=4$ )) at the time of FMT. Clinical resolution

rates after up to two FMTs were high: 27/29 (93%) for immunocompromised individuals, 5/6 (83%) for patients with IBD.

There are also limited data from case series and single case reports describing the use of FMT in patients with immunocompromise. Agrawal and colleagues<sup>55</sup> included 46/146 (32%) patients with a history of cancer, and an additional 15/146 (10%) patients with non-cancer-related immunologic dysfunction, although primary outcome measures were not specifically reported for these groups. Overall cure at 12 weeks in a case series of 80 patients with immunocompromise was reported in 71 (89%) of patients<sup>60</sup>. Adverse events occurred in 12 (15%) immunocompromised patients; this included two deaths (one due to respiratory failure and another due to pneumonia resulting from aspiration at the time of FMT administration)<sup>60</sup>; however, such adverse events have also been reported in non-immunocompromised patient populations<sup>80</sup>. Hefazi and coauthors described high efficacy rates in a case series of FMT for recurrent CDI and a range of haematological or solid organ malignancies (remission after one FMT in 11/12 with haematological patients, and 8/10 in solid organ malignancy patients). No significant FMT-related complications were reported<sup>81</sup>. A further case series<sup>45</sup> reported FMT treatment for 75 patients with recurrent CDI and found no significant difference in primary cure rates for patients with diabetes mellitus, malignancy, or steroid use in the preceding three months.

The working group discussed the potential impact of donor EBV and CMV status for the immunocompromised FMT recipient at risk of severe infection if exposed to these viruses. Their opinion was that such recipients should only receive FMT from donors with negative EBV and CMV status.

**Recommendations:**

- i. **We recommend that FMT should be offered with caution to immunosuppressed patients, in whom FMT appears efficacious without significant additional adverse effects (GRADE of evidence: moderate; strength of recommendation: strong).**
- ii. **We recommend that immunocompromised FMT recipients at risk of severe infection if exposed to EBV or CMV should only receive FMT from donors negative for EBV and CMV (GRADE of evidence: very low; strength of recommendation: strong).**

**5.2.3. Other comorbidities and FMT:**

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Only a limited number of cited studies included specific detail about the presence of comorbidities in patients receiving FMT. However, several studies reported median Charlson comorbidity scores<sup>12,14,15,18,50</sup>. One randomised study reported the presence of IBD in 10/17 (59%) FMT recipients<sup>16</sup>, and there did not appear to be any significant difference in primary outcome measures in this group. Another randomised trial included 14/72 (33%) patients with IBD and reported clinical cure of CDI in 12/14 (86%) of these patients<sup>13</sup>. This study also included 64/72 (89%) patients with cardiac, respiratory, renal, central nervous system or multi-organ system comorbidities<sup>13</sup>; however outcomes were not stratified according to co-morbidity. Kelly and coauthors<sup>60</sup> reported an overall cure rate of 94% in a subset of CDI patients with IBD. A meta-analysis of studies in which patients with IBD received FMT (either primarily as treatment for concurrent recurrent CDI, or with the aim of treating IBD) noted a small risk of exacerbation of IBD in association with the use of FMT<sup>82</sup>. The working group noted the complexity of the relationship between IBD and CDI, given that IBD is itself a risk factor for CDI.

Other exclusions have been more directly related to the mode of administration. For upper gastrointestinal delivery, exclusion criteria have included delayed gastric emptying, chronic aspiration, 'swallow dysfunction', and dysphagia<sup>17,50</sup>. Exclusions for lower GI administration have included colostomy/ileostomy<sup>16,50</sup>, significant bleeding disorders<sup>12</sup>, untreated colorectal cancer<sup>14,36,54</sup>, and ileus/small bowel obstruction<sup>50</sup>.

In summary, the working group noted that co-morbidities amongst patients with recurrent CDI are common. Most studies did not analyse primary outcome measures according to co-morbidity; however, a small number of studies have analysed primary outcome measures (clinical cure) for patients with IBD receiving FMT for recurrent CDI and have found no significant difference compared to those without IBD, along with no overall significant worsening of IBD activity.

#### **Recommendations:**

- i. ***We recommend that FMT should be offered to those with recurrent CDI and inflammatory bowel disease, but patients should be counselled about a small but recognised risk of exacerbation of IBD (GRADE of evidence: moderate; strength of recommendation: strong).***

ii. **We recommend that FMT should be considered for appropriate patients with recurrent CDI regardless of other comorbidities (GRADE of evidence: moderate; strength of recommendation: strong).**

**5.3. What donor factors influence the outcome of faecal microbiota transplant when treating people with *Clostridium difficile* infection?**

**5.3.1. General approach to donor selection:**

Excellent efficacy has been shown in treating recurrent CDI using FMT derived from both related<sup>14,36,54,57,59,61,83,38,40,41,43,45,46,49,53</sup> and unrelated<sup>14,15,57,59,61,72,74,83–87,16,17,35,37,38,41,43,53</sup> donors. To date, there have been no randomised studies comparing differences in efficacy. Case series have tended to rely more on donation of stool from healthy family members. In randomised studies using FMT, all donors were healthy unrelated individuals<sup>12–18,88</sup>. Three case series used donor stool from healthcare professionals<sup>39,61,85</sup>; no randomised studies have used stool from this cohort. However, the working group noted that there were clear advantages to using FMT from a screened anonymous donor, in particular with regards to monitoring and traceability, as discussed further later.

**Recommendation:**

**We recommend that related or unrelated donors should both be considered acceptable. However, where possible, FMT is best sourced from a centralised stool bank, from a healthy unrelated donor (GRADE of evidence: low; strength of recommendation: strong).**

**5.3.2. Age and BMI restrictions for potential donors:**

There are no well-defined age restrictions on donors. Randomised studies have used donors of  $\geq 18$ <sup>12,72</sup> and  $\leq 60$  years old<sup>15,17,18</sup> with satisfactory outcomes. Two of the case series defined age limitations for donors as  $\geq 18$  and  $\leq 50$  years<sup>72,89</sup>. A recent study demonstrated that *Bacteroides: Firmicutes* ratio and microbial diversity was similar for donors above and below 60 years, and their stool donations had similar clinical efficacy as FMT; however, there were loss of the phylum *Actinobacteria* and family *Bifidobacteriaceae* from donors older than 60 years<sup>90</sup>. On balance, the working group agreed that an age range of 18 – 60 years was appropriate for donors.

A widely-reported case study noted apparent weight gain in a recipient of FMT for treatment of CDI when an overweight donor was used<sup>91</sup>, but any association between a donor with a raised BMI and weight gain post-FMT has not been replicated elsewhere in the literature<sup>92</sup>. Whereas most randomised studies did not report donor-specific BMIs, some have excluded those without a 'normal' BMI<sup>13,17</sup>. The working group considered an acceptable BMI for donors as between  $\geq 18$  to  $\leq 30$  kg/m<sup>2</sup>.

**Recommendation:**

**We suggest that people should only be considered as potential FMT donors if they are  $\geq 18$  and  $\leq 60$  years old, and have a BMI of  $\geq 18$  and  $\leq 30$  kg/m<sup>2</sup> (GRADE of evidence: low; strength of recommendation: weak).**

**5.3.3. General approach to the donor screening assessment:**

There is a clear theoretical risk of the transmission of infection by FMT; furthermore, given the large number of conditions in which perturbation of the gut microbiota has been described<sup>93</sup>, there is a concern regarding a risk of transmission of microbiota associated with vulnerability to disease. Whilst FMT is efficacious for recurrent CDI, adverse events may be associated with its use (discussed further later), and long-term safety follow-up is lacking. The aim of a donor screening questionnaire and interview is to minimise post-FMT adverse events by excluding potential donors from whom FMT may be associated with risk to recipients. Randomised studies performed to date used various pre-screening questionnaires, including self-screening questionnaires which focused on high risk behaviours for blood-borne infections<sup>12-16</sup>, questionnaires that focused on previous potential transferable medical conditions<sup>18</sup>, and adaptations from the American Association of Blood Banks Donor Questionnaire<sup>14,17</sup>. One randomised study used the OpenBiome questionnaire as a screening questionnaire<sup>94</sup>. Some studies have suggested excluding potential donors who have recently travelled to defined regions (typically tropical areas), varying between 3-6 months prior to donation<sup>38,39,49,52,55,59,74,87</sup>; this is also the protocol employed in randomised studies<sup>14,16,18</sup>. Another important point for assessment is recent use of medications by potential donors. In particular, given the profound effects of antimicrobials on the gut microbiota<sup>95-98</sup> (along with the theoretical concern that recent antimicrobials might precipitate gut colonisation with antimicrobial-resistant bacteria that could be transferred during FMT), studies advocate either a three month<sup>14,46,53-55,57,61,74</sup> or six month<sup>16-18,35,38,39,43,49,85,99,100</sup> period without antimicrobial use prior to FMT donation.



986

987 The working group agreed that, given the growing evidence for the contribution of the gut  
988 microbiota to the aetiopathogenesis of colorectal carcinoma, patients with a significant personal or  
989 family history of (or risk factors for) this condition should be excluded as donors (**Table 1**). However,  
990 the working group noted an added complexity, in that their recommendation was that potential  
991 donors may be up to 60 years of age, but bowel scope screening for colorectal carcinoma currently  
992 begins within the UK at 55 years of age, and formal NHS bowel cancer screening starts at the age of  
993 60 years<sup>101</sup>. The working group agreed that potential donors living in countries with bowel cancer  
994 screening programmes that start before the age of 60 years should have therefore completed  
995 appropriate screening with negative/ normal tests before they are considered further as donors.

996

997 The working group was of the opinion that a screening process is mandatory; any positive responses  
998 should usually result in exclusion from donation, although this will depend upon the particular  
999 circumstances/ answers given. A donor screening questionnaire should be performed both prior to  
1000 considering a person as a donor, and also at a further point in time (discussed further in **Section**  
1001 **5.3.5**).

1002

1003 **Recommendation:**

1004 ***It is mandatory to screen potential donors by questionnaire and personal interview, to***  
1005 ***establish risk factors for transmissible diseases and factors influencing the gut microbiota***  
1006 ***(Table 1) (GRADE of evidence: low; strength of recommendation: strong).***

1007

1008 **5.3.4. Laboratory screening of potential donors:**

1009 Currently, there are no known confirmed cases of blood-borne pathogens being transmitted by FMT,  
1010 but strict preventative measures are important, as the potential risk of transmission is unknown.  
1011 Many of the suggestions are extended from established blood screening guidelines<sup>102</sup>. Case series  
1012 almost universally screen for HIV, hepatitis B and hepatitis C as a minimum<sup>35,36,52–</sup>  
1013 <sup>55,59,61,72,74,84,86,37,87,103,39–43,46,49</sup>; other studies (including the randomised trials) have a more thorough  
1014 blood screening process<sup>14–18</sup>. Many studies have also included a ‘metabolic/general blood screen’, to  
1015 select out donors with hitherto undiagnosed chronic illness. **Table 2** shows the suggested blood  
1016 screening protocol of the BSG/HIS working group.

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1017

1018 The working group specifically discussed the role of screening donors for their EBV and CMV status;  
1019 the importance of the rationale for this is discussed in **Section 5.2.2**. They agreed that EBV and CMV  
1020 testing was only required where there is the potential that the FMT prepared from that donor would  
1021 be administered to immunosuppressed patients at risk of severe infection if exposed to CMV and  
1022 EBV.

1023

1024 The primary aim of stool screening of potential donors is to minimise the risk of transmission of  
1025 pathogens; again, the relative novelty of FMT for CDI means that these risks are not currently well-  
1026 defined. Stool screening protocols are universal amongst published studies, though widely-variable  
1027 protocols have been used. **Table 3** displays the suggested stool screening protocol of the working  
1028 group. The working group discussed stool screening for multi-drug resistant bacteria carriage, and  
1029 agreed that carbapenemase-producing *Enterobacteriaceae* (CPE) should be screened for. Although  
1030 these bacteria are carried only by a minority of the UK population, transfer into debilitated patients  
1031 with CDI is clearly undesirable given that CPE are potentially so difficult to treat. They also agreed  
1032 that extended-spectrum beta-lactamase (ESBL)-producing organisms could also potentially cause  
1033 severe disease (with limited antimicrobial options) if transplanted into patients with CDI, and so  
1034 should also be screened for. Whilst vancomycin-resistant *Enterococci* (VRE) carriage is relatively  
1035 common in the community (probably related to food consumption)<sup>104</sup>, community strains of VRE are  
1036 genetically distinct from (and generally of much lower pathogenicity than) those found  
1037 nosocomially<sup>105</sup>; as such, the working group thought that routine screening was not justified. The  
1038 working group also noted that methicillin-resistant *Staphylococcus aureus* (MRSA) carriage is very  
1039 rare in healthy adults in non-healthcare settings (with significant intestinal carriage even rarer), so  
1040 did not justify routine screening. However, the working group acknowledged that the potential  
1041 infection risk from VRE and MRSA would vary regionally dependent upon local prevalence and  
1042 pathogenicity, and as such recommended that a risk assessment is performed to assess whether  
1043 screening for these organisms should be considered.

1044

1045 A donor laboratory screening should be performed both prior to considering a person as a donor,  
1046 and also at a further point in time (discussed further in **Section 5.3.5**).

1047

1048 **Recommendation:**

**Blood and stool screening of donors is mandatory (Tables 2 and 3) (GRADE of evidence: low; strength of recommendation: strong).**

**5.3.5. Repeat donor checks, and donation pathway:**

Almost all reviewed studies have repeated at least some elements of the initial donor screening process either at the time of donation of each stool sample used to prepare FMT, or at the end of a period of donation to assess ongoing suitability for inclusion. However, protocols have differed widely between studies.

The opinion of the working group was that when a donor had met criteria for donation (both with an acceptable health questionnaire and satisfactory laboratory tests), they were suitable to begin donation of stool that may be prepared into FMT. Repeat donor screening was also deemed necessary. In centres where frozen FMT is being prepared, stool may be collected and processed immediately after the first donor screen is successfully completed, but should be stored in ‘quarantine’ pending further donor screening, rather than used immediately for clinical use. At the end of the locally-defined period of donation, potential donors should undergo repeat testing, with a further health questionnaire and laboratory screening. If the donor’s health questionnaire remains acceptable and repeat laboratory screening is negative at this point, then the frozen FMT may be released from ‘quarantine’, and used. The working group thought that donor screening both before and after donation was the safest route possible, and that this represented the preferred scenario. A proposed summary pathway for donor screening in this scenario is provided in **Figure 1**.

In centres using fresh FMT, the working group agreed that a repeat health questionnaire should be completed at the time of donation of each stool sample used to prepare FMT. Formal repetition of both the personal interview/ health questionnaire and laboratory screening tests should occur at regular intervals to ensure ongoing suitability for inclusion as a donor. The working group’s opinion was that this repetition of the screening process should occur at least once every four months.

**Recommendations:**

- i. In centres using frozen FMT, before FMT may be used clinically, we recommend that donors should have successfully completed a donor health questionnaire and***

*laboratory screening assays both before and after the period of stool donation. This is the preferred means of donor screening (GRADE of evidence: low; strength of recommendation: strong).*

*ii. In centres using fresh FMT, we recommend that a repeat health questionnaire should be assessed at the time of each stool donation. To ensure ongoing suitability for inclusion as a donor, the donor health questionnaire and laboratory screening should be repeated regularly (GRADE of evidence: low; strength of recommendation: strong).*

**5.4. What factors related to the preparation of the transplant influence the outcome of faecal microbiota transplant when treating people with *Clostridium difficile* infection?**

**5.4.1. General principles of FMT preparation:**

There is very little evidence or guidance on the collection of donor stool. Critical steps during this process centre on the reduction of environmental cross-contamination risk, so the use of clean collection devices and clean collection procedures is advocated. To promote standardised practice and a safe and effective product, clear instructions should be provided to the donor for stool collection (Table 5).

Regardless of the methods used to prepare FMT, stool donations should be processed within six hours of defaecation. The period of six hours has been generally applied across many successful studies of FMT treatment in CDI<sup>14,18,35,39,43,52</sup>, although no formal comparative study has been undertaken. This strategy aims to minimise sample degradation and alteration over time, which may occur due to the complex metabolic and environmental requirements of the faecal microbiota.

There are no comparative trials of anaerobically versus aerobically prepared FMT in the treatment of recurrent CDI. With the exception of small observational studies<sup>41,74</sup>, the vast majority of FMT preparation has been undertaken aerobically for the treatment of CDI and has proved highly efficacious. There appears to be no clear need to process anaerobically, a method which introduces complexity and cost for the treatment of CDI.

1110 The reviewed randomised studies reported variable amounts of stool used in the preparation of  
1111 each FMT aliquot, and the lack of comparative data means that it is not possible to link stool mass to  
1112 outcome from these studies. However, a previous systematic review of case series using FMT as  
1113 treatment for recurrent CDI reported similar rates of treatment efficacy, but an approximate  
1114 fourfold increase in recurrence rates, if <50g of stool was used compared to ≥50g<sup>106</sup>. Similarly, the  
1115 initial volume of diluent used to create the faecal emulsion is variable between studies, although the  
1116 most common practice appears to be creation of a stool: diluent ratio of approximately 1:5. The  
1117 overwhelming majority of the reviewed studies used stool from only a single donor per FMT (rather  
1118 than stool pooled from a mixture of donors), and there are no comparative studies of outcomes of  
1119 CDI from single donor vs pooled donor FMT; as such, the working group found no justification to  
1120 recommend donor stool pooling for FMT for CDI.

1122 The majority of studies have used preservative-free sterile 0.9% saline as the diluent for FMT  
1123 production, although there have been a handful of reports of other diluents including potable  
1124 water<sup>16,35,43</sup>. There have been no comparative studies of FMT diluent. In cases where frozen FMT is  
1125 prepared, an appropriate cryoprotective substance should be added prior to freezing. Most studies  
1126 use glycerol at a final concentration of ~10%<sup>16,41</sup>. It has been demonstrated that storing stool at -  
1127 80°C for up to six months in saline without glycerol decreases viable aerobic and anaerobic bacterial  
1128 counts; the reduction was statistically significant in all bacterial groups with the exception of *E. coli*  
1129 and total anaerobes. When stored with glycerol, no significant reduction in viable counts was  
1130 observed<sup>74</sup>.

1132 A variety of homogenisation and open filtration systems have been used, with no apparent major  
1133 variation in efficacy. Open filtration systems such as gauze<sup>16,37,40,55</sup>, filter paper<sup>39</sup> and strainers/  
1134 sieves<sup>17,41</sup> are unpleasant to use and pose a risk of external contamination. In order to best comply  
1135 with GMP standards, a sterile, single-use closed homogenisation and filtration system is  
1136 recommended. An example of such a system includes the use of sterile filter bags inside a  
1137 laboratory paddle homogeniser.

1139 **Recommendations:**

- 1140 ***i. We recommend that donor stool collection should follow a standard protocol***  
1141 ***(GRADE of evidence: low; strength of recommendation: strong).***

- 1142 **ii. We recommend that donor stool should be processed within 6 hours of defaecation**  
 1143 **(GRADE of evidence: low; strength of recommendation: strong).**
- 1144 **iii. We recommend that both aerobically and anaerobically prepared FMT treatments**  
 1145 **should be considered suitable when preparing FMT for the treatment of recurrent**  
 1146 **CDI (GRADE of evidence: moderate; strength of recommendation: strong).**
- 1147 **iv. We recommend that sterile 0.9% saline should be considered as an appropriate**  
 1148 **diluent for FMT production, and cryoprotectant such as glycerol should be added**  
 1149 **for frozen FMT (GRADE of evidence: moderate: strength of recommendation:**  
 1150 **strong).**
- 1151 **v. We recommend using ≥50g of stool in each FMT preparation (GRADE of evidence:**  
 1152 **moderate: strength of recommendation: strong).**
- 1153 **vi. We suggest that stool should be mixed 1:5 with diluent to make the initial faecal**  
 1154 **emulsion (GRADE of evidence: low; strength of recommendation: weak).**
- 1155 **vii. We suggest that homogenisation and filtration of FMT should be undertaken in a**  
 1156 **closed disposable system (GRADE of evidence: low; strength of recommendation:**  
 1157 **weak).**

#### 1159 **5.4.2. Fresh vs frozen FMT:**

1160 Two randomised studies have examined this area. One double-blind randomised study concluded  
 1161 that enema frozen FMT ( $n=91$ ) was non-inferior for clinical resolution of diarrhoea to fresh FMT  
 1162 ( $n=87$ ) for the treatment of recurrent or refractory CDI<sup>16</sup> (with frozen FMT in this study stored at -  
 1163 20°C for up to 30 days). A further randomised study demonstrated statistically comparable  
 1164 remission rates for recurrent CDI with fresh or frozen FMT delivered colonoscopically ( $n=25/25$  vs  
 1165 20/24 respectively,  $p=0.233$ ) (using frozen FMT stored at -80°C for up to six months)<sup>13</sup>. These data  
 1166 support the findings of earlier small observational studies<sup>35,41</sup>. Frozen FMT is preferable to fresh FMT  
 1167 on logistical and cost grounds<sup>16</sup>. Banked frozen FMT also enables the window period for donor  
 1168 screening to be minimised, allowing centres to more closely to meet regulatory requirements (also  
 1169 see **Section 5.3.5**).

#### 1171 **Recommendation:**

**We recommend that the use of banked frozen FMT material should be considered preferable to fresh preparations for CDI (GRADE of evidence: high; strength of recommendation: strong).**

**5.4.3. Use of frozen FMT:**

Frozen FMT has been used up to six months after storage at  $-80^{\circ}\text{C}^{17,41,74}$ , with high efficacy rates (>70%) observed in the cases treated. However, there have been no comparative trials investigating storage durations. A trend towards decrease in the viability of certain gut microbiota taxa was noted when faecal aliquots were frozen in 10% glycerol for six months<sup>74</sup>, and as such, the working group agreed that six months was the acceptable limit for freezing of an FMT in glycerol. Storage at  $-80^{\circ}\text{C}$  is recommended rather than  $-20^{\circ}\text{C}$  to minimise sample degradation.

Warm water baths have been recommended to speed thawing<sup>6</sup>; however, the working group thought that this should be strongly discouraged, as this may introduce risks of cross contamination by *Pseudomonas* species (and other contaminants) from the water bath<sup>107,108</sup>, and may reduce bacterial viability in the FMT. Repetitive freeze thawing of FMT samples should be avoided as bacterial numbers will be reduced during this process<sup>109</sup>.

**Recommendations:**

- i. We recommend that FMT material stored frozen at  $-80^{\circ}\text{C}$  should be regarded as having a maximum shelf life of six months from preparation (GRADE of evidence: low; strength of recommendation: strong).**
- ii. We suggest consideration of thawing frozen FMT at ambient temperature, and using within six hours of thawing (GRADE of evidence: low; strength of recommendation: weak).**
- iii. We suggest not thawing FMT in warm water baths, due to the risks of cross contamination with *Pseudomonas* (and other contaminants) and reduced bacterial viability (GRADE of evidence: very low; strength of recommendation: weak).**



**5.5. What factors related to administration of the transplant influence the outcome of faecal microbiota transplant when treating people with *Clostridium difficile* infection?**

**5.5.1. Use of specific medications in the period around FMT administration:**

**5.5.1.1. General principles of FMT administration:**

Bowel purgatives have been proposed pre-FMT as a means of removing residual antibiotics that may affect engraftment of transplanted microorganisms, and as a means of removing any residual *C. difficile* toxin, spores and vegetative cells<sup>110–114</sup>. Furthermore, bowel purgatives pre-colonoscopy FMT delivery facilitate safe endoscopy. Various bowel purgatives have been used in colonoscopic FMT studies, including polyethylene glycol (PEG) (often 4 litres)<sup>14,17,115–117,35,41,43,46,54–56,100</sup>, Moviprep<sup>®35,41</sup>, and macrogol<sup>13,15,18,59</sup>. In those studies that used an upper GI route for FMT, PEG<sup>54,55,84</sup> and Klean-Prep<sup>®15,61</sup> were used. FMT without bowel preparation has also been used as treatment for recurrent CDI without any apparent reduction in efficacy, including in randomised studies<sup>16</sup>.

The rationale for the use of proton pump inhibitors (PPI) prior to upper GI FMT is to minimise acidity which may impair engraftment of transplanted microorganisms; however, PPIs have been shown to alter the gut microbiota<sup>118,119</sup>, and have also been associated with primary and recurrent CDI<sup>120,121</sup>. Some studies advocate the use of PPI prior to receiving FMT via the upper GI route<sup>37,39,45,84,85,122,123</sup>, but there appears to be comparable efficacy data in studies where it has not been used. Certain studies have also given recipients PPI prior to receiving colonoscopic FMT<sup>17,87</sup>.

The use of prokinetics (such as metoclopramide) has been described prior to FMT delivery via the upper GI tract route, but only in a very small number of studies<sup>85</sup>. Given the potential risk of regurgitation/aspiration associated with upper GI administration of FMT, the working group felt that its use should be considered where appropriate.

A single dose/ short course of loperamide has been used following FMT (predominantly for lower GI administration) in an attempt to prolong the exposure of the FMT to the mucosa, and to aid retention of the FMT within the GI tract<sup>13,46,49,55,84,123</sup>. One study utilised diphenoxylate with atropine<sup>54</sup> instead. However, no studies have compared FMT with and without anti-motility drugs.

The working group also discussed infection control aspects as they apply to FMT administration. Specifically, they agreed that recipients should ideally be cared for in a single room with en-suite bathroom facilities and, where appropriate, be placed at the end of an endoscopy list, to facilitate enhanced environmental decontamination and prevention of transmission of *C. difficile* spores. Protocols for decontamination of endoscopes should follow national guidance<sup>124,125</sup>, using a sporicidal agent. Best practice for prevention of transmission of healthcare-associated infections, as described in national guidelines<sup>126</sup>, should also be applied throughout.

**Recommendations:**

- i. **We recommend that bowel lavage should be administered prior to FMT via the lower GI route, and bowel lavage should be considered prior to FMT via the upper GI route; polyethylene glycol preparation is preferred (GRADE of evidence: low; strength of recommendation: strong).**
- ii. **For upper GI FMT administration, we suggest that a proton pump inhibitor should be considered, e.g. the evening before and morning of delivery (GRADE of evidence: low; strength of recommendation: weak).**
- iii. **We suggest that a single dose of loperamide (or other anti-motility drugs) should be considered following lower GI FMT delivery (GRADE of evidence: low; strength of recommendation: weak).**
- iv. **We suggest that prokinetics (such as metoclopramide) should be considered prior to FMT via the upper GI route (GRADE of evidence: low; strength of recommendation: weak).**
- v. **We recommend that best practice for prevention of further transmission of CDI should be applied throughout when administering FMT to patients with CDI (nursing with enteric precautions, sporicidal treatment of endoscope, etc) (GRADE of evidence: high; strength of recommendation: strong).**

**5.5.1.2. Additional antibiotics pre-FMT:**

Many studies have given further courses of conventional antimicrobial *C. difficile* treatment prior to FMT. Regimens have included vancomycin alone<sup>12,14,18,35,39,55,59,86,117</sup>, metronidazole or

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vancomycin<sup>40,41,43,122</sup>, or alternatively vancomycin, fidaxomicin or metronidazole<sup>56</sup>, with one study using a range of regimens which included rifaximin<sup>123</sup>. The length of treatment was also variable, ranging from 24 hours<sup>54</sup> up to four days prior to receiving FMT<sup>39,45</sup>; however, comparative studies have not been undertaken.

1267

#### 1268 ***Recommendation:***

1269 **We recommend the administration of further antimicrobial treatment for CDI for at least**  
1270 **72 hours prior to FMT (GRADE of evidence: low; strength of recommendation: strong).**

1271

#### 1272 **5.5.1.3. Washout period between antibiotic use and FMT:**

1273 Nearly all studies specified a washout period after completing anti-CDI antibiotics and before  
1274 administration of FMT. However, this time period appeared to be arbitrarily selected and varied  
1275 from as little as four<sup>46</sup> or 12 hours<sup>51</sup>, up to 72 hours<sup>36</sup>. The majority of studies specified either 24  
1276 hours<sup>15,37,39,40,45,54,127</sup> or 48 hours<sup>41,42,49,60</sup>, however some allowed a range from 1-3 days<sup>16,44,52,53,55</sup>.  
1277 One study appeared to allow co-administration of vancomycin with bowel preparation, without a  
1278 washout period<sup>18</sup>.

1279

1280 The working group discussed the challenging scenario of providing FMT to patients with recurrent  
1281 CDI, but who also had a strong indication for long-term non-anti-CDI antibiotics (e.g. splenectomy,  
1282 osteomyelitis, or infective endocarditis), or patients who develop an indication for antibiotics for a  
1283 reason other than CDI shortly after receiving FMT. The concern in this instance is that the use of  
1284 antibiotics may limit engraftment of microbial communities derived from the FMT, and therefore  
1285 reduce its effectiveness. The working group discussed a recent retrospective study demonstrating  
1286 that exposure to non-anti-CDI antimicrobials within eight weeks of FMT is associated with an  
1287 approximate threefold risk of FMT failure ( $n=8/29$  failures with antibiotic exposure vs  $36/320$  failures  
1288 without antibiotic exposure)<sup>128</sup>. Similarly, the experience of the large pan-Netherlands stool bank<sup>129</sup>  
1289 was that ~50% of their failures of FMT in the treatment of recurrent CDI occurred in patients who  
1290 had received antibiotics within one month of their FMT. For patients requiring long-term antibiotics,  
1291 the working group's expert opinion was that such patients should still be eligible for FMT, but that  
1292 the regimen for the use of non-anti-CDI antibiotics should be decided on a case-by-case basis, based  
1293 on factors including response to FMT and/or strength of indication of antibiotics. Both in this  
1294 scenario, and the scenario in which antibiotics are required shortly after receiving FMT, the working

1295 party agreed that infectious diseases specialists/medical microbiologists should be involved in  
1296 making decisions regarding the choice of agents used.

1298 **Recommendations:**

1299 *iii. To minimise any deleterious effect of antimicrobials on the FMT material, we*  
1300 *recommend that there should be a minimum washout period of 24 hours between the*  
1301 *last dose of antibiotic and treatment with FMT (GRADE of evidence: low; strength of*  
1302 *recommendation: strong).*

1303 *iv. We suggest considering consultation with infectious disease specialists or medical*  
1304 *microbiologists for advice whenever FMT recipients also have an indication for long-*  
1305 *term antibiotics, or have an indication for non-CDI antibiotics within eight weeks of*  
1306 *FMT (GRADE of evidence: very low; strength of recommendation: weak).*

1308 **5.5.2. Route of FMT delivery:**

1309 **5.5.2.1. Introduction:**

1310 FMT can be delivered via the lower GI route (retention enema, colonoscopy), upper GI route  
1311 (endoscopically, or via nasogastric tube, nasoduodenal or nasojejunal tube), or via capsules  
1312 (containing either frozen FMT or lyophilised faecal material). Systematic reviews with meta-analysis  
1313 suggest that FMT for recurrent CDI via colonoscopy may have slightly higher efficacy compared to  
1314 upper GI administration<sup>127,130–132</sup> with similar safety profiles, but also note the trend towards using  
1315 larger amounts of stool or ‘higher concentration’ FMT in lower GI administration. One systematic  
1316 review (reviewing principally case series, and including only one randomised study) compared  
1317 remission rates for CDI using FMT delivered to different areas of the GI tract, and reported that for  
1318 FMT infused into the stomach, duodenum/jejunum, caecum/ascending colon, and rectum the rates  
1319 of cure rate were 81%, 86%, 93%, and 84%, respectively<sup>131</sup>.

1321 In the only randomised study that directly compared upper and lower GI administration, there was  
1322 no significant difference in overall cure rate ( $p = 0.53$ )<sup>17</sup>.

1324 **5.5.2.2. Upper gastrointestinal tract administration of FMT:**

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FMT has been shown to be safe and efficacious in the treatment of *C. difficile* when administered via nasogastric tube<sup>37,39,45,61,83,123</sup>, nasoduodenal tube<sup>15,84,85</sup>, enteroscopy<sup>122,123</sup>, or via the infusion channel on a gastroscope<sup>40,45</sup>. In a randomised trial, nasoduodenal donor FMT has been shown to be more efficacious than vancomycin in treating recurrent CDI<sup>15</sup>. Furthermore, it has been shown that FMT can also be safely and effectively delivered via a percutaneous endoscopic gastrectomy tube<sup>45,83</sup>. The working group noted that upper GI administration of FMT may be particularly suitable for certain patient groups, such as those in whom there are contraindications or who would find it difficult to tolerate lower GI endoscopy, and/ or patients unlikely to be unable to retain enemas.

1333

Typically, smaller volumes of faecal suspension are administered to the upper GI tract compared to lower GI administration, with quoted volumes ranging from 25ml<sup>39</sup> up to 150ml<sup>84</sup>- 250ml<sup>37,85</sup>. Up to 500ml of suspension has been given safely and effectively via the upper GI route<sup>15,77</sup>. However, the working group expressed concerns regarding the risk of regurgitation and aspiration if large volumes of FMT are administered to the upper GI tract, and also discussed cases in which this has been described with adverse outcomes<sup>80</sup>. This included a reported death from aspiration, after 100-150ml of FMT was delivered by enteroscope into the distal duodenum under general anaesthetic as attempted treatment for recurrent CDI<sup>133</sup>. A further reported case described a case of fatal aspiration pneumonitis likely related to a 500ml FMT via nasoduodenal tube; this patient had a swallowing disorder following oropharyngeal radiation after surgical removal of a maxillary carcinoma two years previously<sup>77</sup>. Based on their expert opinion, the working group recommended that upper GI FMT should be used with caution in those at risk of regurgitation (e.g. known large hiatus hernia, severe gastro-oesophageal reflux disease, etc) and/ or with swallowing disorders (although administration via a gastrostomy tube would be acceptable). They also recommended that no more than 100ml of FMT should be administered to the upper GI tract to minimise these risks.

1350

#### 1351 **Recommendations:**

- 1352 **i. We recommend that upper GI administration of FMT as treatment for recurrent or**
- 1353 **refractory CDI should be used where clinically appropriate (GRADE of evidence:**
- 1354 **high; strength of recommendation: strong).**
- 1355 **ii. Where upper GI administration is considered most appropriate, we recommend**
- 1356 **that FMT administration should be via nasogastric, nasoduodenal, or nasojejunal**

*tube, or alternatively via upper GI endoscopy. Administration via a permanent feeding tube is also appropriate (GRADE of evidence: high; strength of recommendation: strong).*

**v. We recommend that no more than 100ml of FMT is administered to the upper GI tract (GRADE of evidence: low; strength of recommendation: strong).**

**vi. We recommend that upper GI administration of FMT should be used with caution in those at risk of regurgitation and/ or those with swallowing disorders (GRADE of evidence: low; strength of recommendation: strong).**

**5.5.2.3. Lower gastrointestinal tract administration of FMT:**

**FMT via enema:** Successful treatment of *C. difficile* with FMT enema has been demonstrated<sup>16,38,42,53,55,83,86</sup> but enema appears to have a lower efficacy than other routes of FMT administration. Specifically, in a randomised study primarily comparing the efficacy of fresh and frozen FMT in the treatment of recurrent CDI, only 52.8% of patients in the ‘frozen’ arm and 50.5% of patients in the ‘fresh’ arm of the study ( $n=57/108$  and  $56/111$  respectively) experienced resolution of symptoms after a single enema, by modified intention to treat analysis<sup>16</sup>. However, resolution rates in both arms only reached >80% after at least three enemas<sup>16</sup>. A recent randomised study demonstrated similar rates of recurrence of CDI in patients with recurrent CDI treated with either a single FMT enema or a six week vancomycin taper ( $n=9/16$  patients with recurrence vs  $5/12$  respectively)<sup>12</sup>. Notwithstanding this, enemas do have specific advantages, such as being a treatment option where full colonoscopy is contraindicated. It is also possible to give multiple infusions relatively easily and outside a hospital setting.

**FMT via colonoscopy:** Randomised study evidence has demonstrated that colonoscopic FMT has higher efficacy in treating recurrent CDI than vancomycin<sup>18</sup>. Efficacy is similar whether FMT is fresh or frozen, but modestly reduced when using a lyophilised FMT product<sup>13</sup>. Colonoscopic delivery of donor FMT into the ileum or caecum was associated with a 91% cure rate for recurrent CDI<sup>14</sup>. Observational studies highlighted similar success, describing cure rates of 88% ( $n=14/16$ )<sup>74</sup> and 91%<sup>46</sup> ( $n=21/23$ ) in response to infusion of donor FMT into the caecum or terminal ileum. A further advantage of using colonoscopy to administer FMT has been to allow assessment for the presence of pseudomembranes; in certain reviewed studies, the presence or absence of pseudomembranes has influenced the FMT regimen used<sup>18,73</sup>. However, the working group noted that that many patients

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with CDI are frail and elderly, and as such it will not always be safe or feasible to undertake colonoscopy in this particular group of patients. Flexible sigmoidoscopy appears to be an feasible option where full colonoscopy cannot be performed e.g. unable to tolerate colonoscopy, severity of colitis<sup>56,60</sup>.

The amount of faecal suspension via enema has varied between 150-500mls<sup>16,38,42,55,86</sup>. The amount of faecal suspension delivered via colonoscopy has been similarly variable, with some studies suggesting as little as 100ml can be used with success rates of 94%<sup>43</sup>. 250ml-400ml had a success rate of 100%<sup>36</sup>, whereas infusions of up to 500-700ml were associated with cure rates of 92%<sup>46</sup>. However, the working group noted that it is difficult to compare 'concentration' of FMT in different studies as different protocols used varied starting amounts of faecal material. Currently, there are no randomised studies that compare concentration/ volume of colonoscopic or enema FMT. As such, no recommendation was made to this regard.

#### **Recommendations:**

- i. **We recommend that colonoscopic administration of FMT as treatment for recurrent or refractory CDI should be used where appropriate (GRADE of evidence: high; strength of recommendation: strong).**
- ii. **Where colonoscopic administration is used, we suggest considering preferential delivery to the caecum or terminal ileum, as this appears to give the highest efficacy rate (GRADE of evidence: low; strength of recommendation: weak).**
- iii. **We recommend that FMT via enema should be used as a lower GI option when delivery using colonoscopy or flexible sigmoidoscopy is not possible (GRADE of evidence: high; strength of recommendation: strong).**

#### **5.5.2.4. Capsulised FMT:**

Capsulised FMT aims to remove some of the concerns regarding conventional FMT, such as the invasive means of administration and palatability. The largest case series describing the use of capsules as treatment for recurrent CDI<sup>72,89</sup> noted clinical resolution at eight weeks off antibiotics for CDI in 82% of cases ( $n=147/180$ ) after one course of capsules, and 91% ( $n=164/180$ ) after two courses. The capsules contained frozen FMT prepared in a diluent of saline with 10% glycerol; 15



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capsules were administered each day for two consecutive days (equating to a mean 48g of original crude stool). Other smaller case series have demonstrated comparable results<sup>87,123,134</sup>, including when lyophilised stool is used instead of frozen whole FMT<sup>134</sup>.

The working group reviewed two randomised studies which have examined the efficacy of capsulised FMT in treating recurrent CDI. In one study, published in abstract form<sup>94</sup>, a 'high dose' regimen of frozen FMT capsules (30 capsules each day for two days) was compared to 'low dose' (30 capsules in one day). CDI resolution was comparably high in both arms with one treatment course (77% ( $n=7/9$ ) in the 'high dose' arm vs 70% ( $n=7/10$ ) in the 'low dose arm'). 4/5 initial non-responders entered remission after a second capsule course with the 'high dose' regimen<sup>94</sup>. In a recent large randomised trial, patients with recurrent CDI were randomised to receive either thawed frozen FMT either via colonoscopy or via capsules (one treatment of 40 capsules)<sup>11</sup>. On per protocol analysis, remission at 12 weeks after a single treatment occurred in 96% in both arms ( $n=51/53$  by capsule,  $n=50/52$  by colonoscopy).

The working group discussed certain unresolved issues regarding capsules. Specifically, capsules are often large, and swallowing 30 capsules in a single day may be a significant undertaking for patients with CDI, such as the frail elderly with an existing high pill burden. They also noted that follow-up data post-capsule administration is relatively short compared to other modalities of FMT.

**Recommendation:**

***Capsulised FMT holds promise as a treatment option for recurrent CDI and we recommend that this should be offered to patients as a potential treatment modality where available.***

***Capsule preparations should follow a standard protocol. Further evidence regarding optimal dosing and formulation is required (GRADE of evidence: high; strength of recommendation: strong).***

**5.6. What is the clinical effectiveness of FMT in treating conditions other than *Clostridium difficile* infection?**

**5.6.1. Introduction:**

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In current clinical practice, FMT is used predominantly in the treatment of recurrent CDI. Its success has led to exploration of its efficacy in other GI diseases, primarily ulcerative colitis (UC), where perturbation of the gut microbiota has been observed and implicated in disease pathogenesis<sup>135</sup>. Due to variability of the quality, methodology and cohorts of patients recruited in trials of FMT for non-CDI indications, and in order to control for significant confounding factors, the working group only included randomised trials involving patients with well-defined conditions and in which there was a primary clinical outcome. To date, there have been a total of 71 such studies investigating the role of FMT in IBD; of these, only four are prospective randomised controlled trials, limited to patients with ulcerative colitis<sup>136–139</sup>. Five other reviewed randomised studies investigated the use of FMT in irritable bowel syndrome<sup>140</sup>, slow transit constipation<sup>141</sup>, hepatic encephalopathy<sup>142</sup> and metabolic syndrome<sup>143,144</sup>.

## **5.6.2. Use of FMT for ulcerative colitis:**

### **5.6.2.1. Efficacy:**

All four RCTs, with a total of 277 subjects, included patients with mild to moderate UC (Mayo score 3-11 and endoscopic sub-score of at least 1). Participants were aged between 27 and 56 years and largely included patients on stable immunosuppressive therapy (only one study excluded patients using biologic treatments and methotrexate within the preceding two months)<sup>136</sup>. Three studies included patients on oral corticosteroids at the time of FMT, however only two required a mandatory wean of these to meet eligibility. Studies generally included patients with all disease distributions found in UC. Time to evaluation of response to FMT in these studies varied between seven and twelve weeks. Two studies used autologous FMT as placebo<sup>136,139</sup>. Three of the four studies demonstrated that patients receiving donor FMT were significantly more likely to achieve clinical and endoscopic remission compared to placebo<sup>137–139</sup>. The pooled rate of combined clinical and endoscopic remission was 27.9% for donor FMT and 9.5% for placebo. A pooled risk ratio for failure of FMT to achieve these combined outcomes was 0.8 (95% CI: 0.7-0.9). Deep remission (histological) was only reported in one RCT: 18.4% of patients receiving FMT achieved this outcome compared to 2.7% of those receiving placebo<sup>137</sup>.

### **5.6.2.2. Characteristics of FMT preparation and delivery:**

The four RCTs varied in their FMT preparation and delivery methodology. Two RCTs delivered frozen FMT, one fresh FMT, and one used a combination. Three RCTs with a positive outcome delivered the FMT via the lower GI route; these studies used a high intensity protocol ranging from a total of three

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1483 infusions in one week to 40 FMTs over an eight week period<sup>137–139</sup>. The other RCT (that failed to  
1484 show efficacy of FMT for UC) had adopted a low intensity protocol of two nasoduodenal infusions  
1485 given three weeks apart<sup>136</sup>. Interestingly, the only RCT that prepared stool in anaerobic conditions  
1486 demonstrated the highest rate of steroid-free clinical remission and steroid-free clinical response  
1487 with donor FMT<sup>139</sup>. A further interesting observation in one study was a trend towards higher rates  
1488 of remission with one particular donor<sup>137</sup>.

1489

#### 1490 **5.6.2.3. Adverse events:**

1491 Short-lived GI symptoms such as abdominal bloating, cramps, diarrhoea and fever were reported in  
1492 patients receiving FMT for UC. There were no significant differences in serious adverse events  
1493 between patients receiving FMT compared to placebo (10 vs 7 respectively). Most of the serious  
1494 adverse events were a consequence of worsening colitis: one patient who received FMT required a  
1495 colectomy<sup>136</sup>. In addition, one patient developed concurrent CDI<sup>137</sup>. No deaths were reported in any  
1496 of the studies.

1497

#### 1498 **5.6.3. Use of FMT in functional bowel disorders:**

1499 Two RCTs have investigated the role of FMT in functional bowel disorders. In a double-blind placebo  
1500 controlled RCT that recruited 90 patients with IBS with diarrhoea or with diarrhoea and  
1501 constipation<sup>140</sup>, the primary endpoint only just reached statistical significance in inducing symptom  
1502 relief (as assessed by 75 point reduction in IBS-severity scoring system at three months following a  
1503 single infusion FMT by colonoscopy) ( $p=0.049$ ). The second RCT randomised 60 patients with slow  
1504 transit constipation to either six consecutive days of nasogastric-delivered FMT or conventional  
1505 treatment<sup>141</sup>. This demonstrated that a significant proportion of patients achieved the primary  
1506 endpoint of a mean of at least three complete spontaneous bowel movements per week (53.3% vs.  
1507 20.0%,  $p=0.009$ ) along with improvement in stool consistency score and colonic transit time.  
1508 However, the intervention group had more treatment-related adverse events than did the control  
1509 group (total of 50 vs 4 cases).

1510

#### 1511 **5.6.4. Use of FMT in hepatic encephalopathy:**

1512 One small study has investigated the role of FMT in the management of hepatic encephalopathy  
1513 (HE)<sup>142</sup>. This RCT randomised 20 male patients with cirrhosis with refractory HE to receive either five  
1514 days of broad-spectrum antibiotic pre-treatment followed by a single FMT enema or standard of

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care. Patients in the FMT arm had a significantly lower incidence of serious adverse events and improved cognition. The Model for End-Stage Liver Disease (MELD) score, however, transiently worsened post-antibiotics in the FMT arm. The study was potentially confounded as patients in the FMT arm continued to receive lactulose and/or rifaximin for treatment of their HE.

#### **5.6.5. Use of FMT for metabolic syndrome:**

Two randomised studies<sup>143,144</sup>, with a combined total of 56 patients, demonstrated an improvement in peripheral (but not hepatic) insulin sensitivity in Caucasian male obese patients with metabolic syndrome following one or two infusions via nasoduodenal tube of FMT obtained from lean donors. This improvement was observed at six weeks post-FMT, but was no longer present by 18 weeks. No improvement in insulin sensitivity was identified in patients transplanted with autologous FMT (i.e. patients transplanted with their own collected faeces). The improvement in peripheral insulin sensitivity in the lean donor FMT group was accompanied by a small but significant improvement in HbA1c at six weeks<sup>144</sup>, but no improvements in other metabolic parameters, such as weight. Whilst these data are of interest, the working group felt that the limited, transient nature of the benefits seen and small size of the studies meant that FMT could not be recommended as treatment for metabolic syndrome.

#### **5.6.6. Future directions for randomised trials of FMT for non-CDI indications:**

Currently there are a large number of randomised trials (including RCTs) being undertaken globally, to evaluate the potential role of FMT as treatment for a wide range of conditions. The working group concluded that until there are more reliable data to inform decision-making, the best practice principles described in this document for the governance of an FMT service for recurrent CDI should also be applied to FMT clinical trials for other conditions. However, specific adaptations may be considered depending on the condition being studied, e.g. consideration of using anaerobic conditions for the preparation of FMT in trials for the treatment of UC, as described above.

In conclusion, FMT has the potential to be an effective treatment option for mild to moderate ulcerative colitis, and appears to be safe despite the use of immunosuppressive therapy. FMT may also have a potential role in the treatment of functional bowel disorders. However, recommendations for clinical use for both these indications cannot be made until there is clearer evidence of the most appropriate patient characteristics, preparation methodology, route of delivery

1547 and intensity of administration of FMT. The evidence for the use of FMT in hepatic encephalopathy  
1548 and metabolic syndrome is currently limited, and further well-designed RCTs are needed to evaluate  
1549 its potential role here.

1551 ***Recommendation:***

1552 ***We do not currently recommend FMT as treatment for inflammatory bowel disease.***  
1553 ***Apart from CDI, there is insufficient evidence to recommend FMT for any other***  
1554 ***gastrointestinal or non-gastrointestinal disease (GRADE of evidence: moderate; strength***  
1555 ***of recommendation: strong).***

1557 **6. Basic requirements for implementing a FMT service:**

1558 As discussed above, there is an absence of published studies to support the recommendations in this  
1559 section (although the experience of setting up a nationwide stool bank has recently been reported  
1560 from the Netherlands<sup>129</sup>). This section is therefore based on the working group's expert opinion and  
1561 experience of developing FMT services. The working group considered best practice in this area as it  
1562 applied to legal and clinical governance aspects, the relevant professionals required to establish an  
1563 FMT service, the infrastructure of a service, and appropriate practices for FMT manufacturing and  
1564 quality control monitoring where relevant. The full text of this section is in **Supplementary Material**  
1565 **3.**

1567 **7. Key performance indicators:**

- 1568 • All donors to have completed initial screening questionnaires and blood and stool screening  
1569 results, as well as final health check prior to each stool donation processed to FMT. Results from  
1570 each subsequent serial round of screening also to be documented.
- 1571 • All FMT recipients to have clear documentation of details of their disease course and  
1572 preparation prior to FMT, including whether recurrent or refractory disease, previous  
1573 antimicrobial courses, and use of bowel purgatives/other preparatory medications pre-FMT.
- 1574 • All FMT recipients to have sufficient documentation to allow clear traceability of the exact FMT  
1575 aliquot transfused. Records should include identification of the donor, as well as a frozen FMT  
1576 aliquot (and original faecal sample) - as well as serum - from that donor (see **Supplementary**  
1577 **Material 3**).

- All FMT recipients for recurrent or refractory CDI to have documentation during follow-up of treatment success or failure (and subsequent treatment plan if failure), together with clear documentation of any adverse events that may be attributable to FMT.

## **8. Further research:**

- As described within this guideline, many aspects of the terminology of CDI are used variably between studies, and end-points in FMT trials are inconsistent. The working group noted the need to standardise this terminology to allow more robust comparisons between studies.
- Given the relative novelty of FMT as a procedure, any potential long-term adverse events associated with its use are poorly-defined. The establishment of formal FMT registries should be considered. Whilst this would primarily act as an important tool for defining the safety and efficacy of FMT, it would also be a valuable database for researchers within the field. Standardisation of other key documentation related to FMT administration (e.g. establishment of a proforma for assessing eligibility for FMT and/or follow-up after FMT) would also be advantageous for the same reasons.
- The working group noted the lack of consistency in definitions related to the severity of CDI disease and to response or failure to FMT. This limited interpretation of the published studies. As such, the working group thought that standardisation of these definitions would allow more accurate delineation of the factors influencing the efficacy of FMT for CDI. The working group also noted that only one reviewed study had reported the relationship between *C difficile* ribotype and FMT outcome, and that recording of this information should be encouraged better to evaluate its influence.
- Further well-designed clinical trials (in particular, RCTs) are required to identify the optimal means of administration of FMT as treatment for recurrent and/or refractory CDI.
- The working group noted that even capsulised FMT may be associated with potential drawbacks. They also noted that there are many patients with recurrent CDI for whom FMT (or any form of 'bacteriotherapy') may be inappropriate, including those with very marked immunosuppression, and/or multi-organ disease. Despite high levels of efficacy, there is a small but appreciable FMT failure rate and it is not currently understood whether this is due to underlying donor or recipient factors. Therefore, a research priority should be in basic and translational studies better to define the mechanisms underlying the efficacy of FMT in CDI. This includes comparing the structure and function of the microbiota of donors to patients pre-FMT and post-FMT, via techniques including next-generation microbial sequencing, metabolic profiling, and

immunological assays. This would allow the refinement of FMT from its current state to a more targeted therapy, removing the concerns associated with FMT.

- The working group identified a need for further well-designed RCTs to investigate the potential role of FMT for non-CDI indications.

**9. Conclusions:**

FMT has become an accepted, efficacious treatment for recurrent and/or refractory CDI. In developing this guideline, the evidence for the technique has been reviewed in the context of other available treatments. Specific guidance for best practice for an FMT service is provided.

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**11. Competing interests:**

- THI: Acted as consultant, advisor or speaker for Pharmacosmos and Shield Therapeutics.
- ALH: Acted as consultant, advisory board member or speaker for AbbVie, Atlantic, Bristol-Myers Squibb, Celltrion, Falk, Ferring, Janssen, MSD, Napp Pharmaceuticals, Pfizer, Pharmacosmos, Shire and Takeda. ALH also serves on the Global Steering Committee for Genentech.
- SDG: Received consultancy fees, speaker fees and research grant support from Astellas between 2015-2017; received consultancy fees and speaker fees from MSD between 2015-2017; and received consultancy fees in 2017 from Pfizer.
- All other authors declared no conflict of interest.

**12. Provenance and peer review:**

Commissioned. Peer review through stakeholder consultation, HIS (SDC and Council), BSG (CSSC and Council) and externally.



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### **14.References:**

1. Lawson PA, Citron DM, Tyrrell KL, Finegold SM. Reclassification of *Clostridium difficile* as *Clostridioides difficile* (Hall and O'Toole 1935) Prévot 1938. *Anaerobe*. 2016;40:95-99. doi:10.1016/j.anaerobe.2016.06.008.
2. Faecal microbiota transplant for recurrent *Clostridium difficile* infection | Guidance and guidelines | NICE. <https://www.nice.org.uk/guidance/ipg485>. Accessed October 2, 2017.

HIS/ BSG FMT Guideline: Main Document, *Gut* version.

- 1665 3. Health England P. Updated guidance on the management and treatment of *Clostridium*
- 1666 *difficile* infection. 2013. <http://www.gov.uk/phe>. Accessed March 20, 2017.
- 1667 4. Debast SB, Bauer MP, Kuijper EJ. European Society of Clinical Microbiology and Infectious
- 1668 Diseases: Update of the Treatment Guidance Document for *Clostridium difficile* Infection. *Clin*
- 1669 *Microbiol Infect*. 2014;20(s2):1-26. doi:10.1111/1469-0691.12418.
- 1670 5. McDonald LC, Gerding DN, Johnson S, et al. Clinical Practice Guidelines for *Clostridium*
- 1671 *difficile* Infection in Adults and Children: 2017 Update by the Infectious Diseases Society of
- 1672 America (IDSA) and Society for Healthcare Epidemiology of America (SHEA). *Clin Infect Dis*.
- 1673 February 2018. doi:10.1093/cid/cix1085.
- 1674 6. Cammarota G, Ianiro G, Tilg H, et al. European consensus conference on faecal microbiota
- 1675 transplantation in clinical practice. *Gut*. 2017;66(4):569-580. doi:10.1136/gutjnl-2016-
- 1676 313017.
- 1677 7. König J, Siebenhaar A, Högenauer C, et al. Consensus report: faecal microbiota transfer -
- 1678 clinical applications and procedures. *Aliment Pharmacol Ther*. 2017;45(2):222-239.
- 1679 doi:10.1111/apt.13868.
- 1680 8. Treating *Clostridium difficile* Infection With Fecal Microbiota Transplantation. *Clin*
- 1681 *Gastroenterol Hepatol*. 2011;9(12):1044-1049. doi:10.1016/J.CGH.2011.08.014.
- 1682 9. Kump P, Krause R, Steininger C, et al. Empfehlungen zur Anwendung der fäkalen
- 1683 Mikrobiotatransplantation „Stuhltransplantation“: Konsensus der Österreichischen
- 1684 Gesellschaft für Gastroenterologie und Hepatologie (ÖGGH) in Zusammenarbeit mit der
- 1685 Österreichischen Gesellschaft für Infektiologie und. *Z Gastroenterol*. 2014;52(12):1485-1492.
- 1686 doi:10.1055/s-0034-1385562.
- 1687 10. Faecal microbiota transplantation in recurrent *Clostridium difficile* infection:
- 1688 Recommendations from the French Group of Faecal microbiota Transplantation. *Dig Liver Dis*.
- 1689 2016;48(3):242-247. doi:10.1016/J.DLD.2015.08.017.
- 1690 11. Kao D, Roach B, Silva M, et al. Effect of Oral Capsule— vs Colonoscopy-Delivered Fecal
- 1691 Microbiota Transplantation on Recurrent *Clostridium difficile* Infection. *JAMA*.
- 1692 2017;318(20):1985. doi:10.1001/jama.2017.17077.
- 1693 12. Hota SS, Sales V, Tomlinson G, et al. Oral Vancomycin Followed by Fecal Transplantation
- 1694 Versus Tapering Oral Vancomycin Treatment for Recurrent *Clostridium difficile* Infection: An
- 1695 Open-Label, Randomized Controlled Trial. *Clin Infect Dis*. 2017;64(3):265-271.

HIS/ BSG FMT Guideline: Main Document, *Gut* version.

- doi:10.1093/cid/ciw731.
- 1697 13. Jiang ZD, Ajami NJ, Petrosino JF, et al. Randomised clinical trial: faecal microbiota  
1698 transplantation for recurrent *Clostridium difficile* infection – fresh, or frozen, or lyophilised  
1699 microbiota from a small pool of healthy donors delivered by colonoscopy. *Aliment Pharmacol*  
1700 *Ther.* 2017;45(7):899-908. doi:10.1111/apt.13969.
  - 1701 14. Kelly CR, Khoruts A, Staley C, et al. Effect of Fecal Microbiota Transplantation on Recurrence  
1702 in Multiply Recurrent *Clostridium difficile* Infection. *Ann Intern Med.* 2016;165(9):609.  
1703 doi:10.7326/M16-0271.
  - 1704 15. van Nood E, Vrieze A, Nieuwdorp M, et al. Duodenal Infusion of Donor Feces for Recurrent  
1705 *Clostridium difficile*. *N Engl J Med.* 2013;368(5):407-415. doi:10.1056/NEJMoa1205037.
  - 1706 16. Lee CH, Steiner T, Petrof EO, et al. Frozen vs Fresh Fecal Microbiota Transplantation and  
1707 Clinical Resolution of Diarrhea in Patients With Recurrent *Clostridium difficile* Infection.  
1708 *JAMA.* 2016;315(2):142. doi:10.1001/jama.2015.18098.
  - 1709 17. Youngster I, Sauk J, Pindar C, et al. Fecal Microbiota Transplant for Relapsing *Clostridium*  
1710 *difficile* Infection Using a Frozen Inoculum From Unrelated Donors: A Randomized, Open-  
1711 Label, Controlled Pilot Study. *Clin Infect Dis.* 2014;58(11):1515-1522. doi:10.1093/cid/ciu135.
  - 1712 18. Cammarota G, Masucci L, Ianiro G, et al. Randomised clinical trial: faecal microbiota  
1713 transplantation by colonoscopy vs. vancomycin for the treatment of recurrent *Clostridium*  
1714 *difficile* infection. *Aliment Pharmacol Ther.* 2015;41(9):835-843. doi:10.1111/apt.13144.
  - 1715 19. Faecal Microbiota Transplantation (FMT) MHRA's position.  
1716 [http://www.bsg.org.uk/images/stories/docs/clinical/guidance/fmt\\_mhra\\_position\\_june2015.](http://www.bsg.org.uk/images/stories/docs/clinical/guidance/fmt_mhra_position_june2015.pdf)  
1717 pdf. Accessed October 3, 2017.
  - 1718 20. Thomas A. HTA Policy on the Regulation of Faecal Microbiota Transplant. 2015.  
1719 [http://www.bsg.org.uk/images/stories/docs/clinical/guidance/hta\\_pol\\_030\\_policy\\_regulation\\_of\\_fmt.pdf](http://www.bsg.org.uk/images/stories/docs/clinical/guidance/hta_pol_030_policy_regulation_of_fmt.pdf). Accessed October 3, 2017.
  - 1721 21. Khoruts A, Sadowsky MJ. Understanding the mechanisms of faecal microbiota  
1722 transplantation. *Nat Rev Gastroenterol Hepatol.* 2016;13(9):508-516.  
1723 doi:10.1038/nrgastro.2016.98.
  - 1724 22. Petrof EO, Gloor GB, Vanner SJ, et al. Stool substitute transplant therapy for the eradication  
1725 of *Clostridium difficile* infection: 'RePOOPulating' the gut. *Microbiome.* 2013;1(1):3.  
1726 doi:10.1186/2049-2618-1-3.

1  
2  
3 1727 23. Ott SJ, Waetzig GH, Rehman A, et al. Efficacy of Sterile Fecal Filtrate Transfer for Treating  
4 1728 Patients With Clostridium difficile Infection. *Gastroenterology*. 2017;152(4):799-811.e7.  
5 1729 doi:10.1053/j.gastro.2016.11.010.  
6  
7  
8 1730 24. Khanna S, Pardi DS, Kelly CR, et al. A Novel Microbiome Therapeutic Increases Gut Microbial  
9 1731 Diversity and Prevents Recurrent *Clostridium difficile* Infection. *J Infect Dis*. 2016;214(2):173-  
10 1732 181. doi:10.1093/infdis/jiv766.  
11  
12  
13 1733 25. Martin J, Wilcox M. New and emerging therapies for Clostridium difficile infection. *Curr Opin*  
14 1734 *Infect Dis*. 2016;29(6):546-554. doi:10.1097/QCO.0000000000000320.  
15  
16  
17 1735 26. Zipursky JS, Sidorsky TI, Freedman CA, Sidorsky MN, Kirkland KB. Patient Attitudes Toward the  
18 1736 Use of Fecal Microbiota Transplantation in the Treatment of Recurrent Clostridium difficile  
19 1737 Infection. *Clin Infect Dis*. 2012;55(12):1652-1658. doi:10.1093/cid/cis809.  
20  
21  
22 1738 27. Kahn SA, Vachon A, Rodriguez D, et al. Patient perceptions of fecal microbiota  
23 1739 transplantation for ulcerative colitis. *Inflamm Bowel Dis*. 2013;19(7):1506-1513.  
24 1740 doi:10.1097/MIB.0b013e318281f520.  
25  
26  
27 1741 28. Quraishi MN, Segal J, Mullish B, et al. National survey of practice of faecal microbiota  
28 1742 transplantation for Clostridium difficile infection in the UK. *J Hosp Infect*. 2016.  
29 1743 doi:10.1016/j.jhin.2016.10.023.  
30  
31  
32 1744 29. Porter RJ, Fogg C. Faecal microbiota transplantation for Clostridium difficile infection in the  
33 1745 United Kingdom. *Clin Microbiol Infect*. 2015;21(6):578-582. doi:10.1016/j.cmi.2015.01.020.  
34  
35  
36 1746 30. Ding NS, Mullish BH, McLaughlin J, Hart A, Marchesi JR. Meeting update: faecal microbiota  
37 1747 transplantation—bench, bedside, courtroom? *Frontline Gastroenterol*. November  
38 1748 2016:flgastro-2016-100752. doi:10.1136/flgastro-2016-100752.  
39  
40  
41 1749 31. Prior AR, Kevans D, McDowell L, Cudmore S, Fitzpatrick F. Treatment of Clostridium difficile  
42 1750 infection: a national survey of clinician recommendations and the use of faecal microbiota  
43 1751 transplantation. *J Hosp Infect*. 2017;95(4):438-441. doi:10.1016/j.jhin.2016.10.004.  
44  
45  
46 1752 32. 1995 - The well-built clinical question: a key to evidence-based decisions (Editorial) | 1995  
47 1753 Nov-Dec : Volume 123, Number 3, Page A12 | ACP Journal Club Archives.  
48 1754 <https://acpjc.acponline.org/Content/123/3/issue/ACPJC-1995-123-3-A12.htm>. Accessed  
49 1755 October 18, 2017.  
50  
51  
52 1756 33. British Society of Gastroenterology CS and SC. Guideline Development Within the BSG Clinical  
53 1757 Services and Standards Committee Policies. <https://www.bsg.org.uk/resource/guideline->

HIS/ BSG FMT Guideline: Main Document, *Gut* version.

- development-within-the-bsg-clinical-services-and-standards-committee-policies.html.
- Accessed April 25, 2018.
34. Guyatt GH, Oxman AD, Vist GE, et al. GRADE: an emerging consensus on rating quality of evidence and strength of recommendations. *BMJ*. 2008;336(7650):924-926. doi:10.1136/bmj.39489.470347.AD.
35. Satokari R, Mattila E, Kainulainen V, Arkkila PET. Simple faecal preparation and efficacy of frozen inoculum in faecal microbiota transplantation for recurrent *Clostridium difficile* infection - an observational cohort study. *Aliment Pharmacol Ther*. 2015;41(1):46-53. doi:10.1111/apt.13009.
36. Yoon SS, Brandt LJ. Treatment of Refractory/Recurrent *C. difficile*-associated Disease by Donated Stool Transplanted Via Colonoscopy. *J Clin Gastroenterol*. 2010;44(8):562-566. doi:10.1097/MCG.0b013e3181dac035.
37. Zainah H, Hassan M, Shiekh-Sroujeh L, Hassan S, Alangaden G, Ramesh M. Intestinal microbiota transplantation, a simple and effective treatment for severe and refractory *Clostridium difficile* infection. *Dig Dis Sci*. 2014;60(1):181-185. doi:10.1007/s10620-014-3296-y.
38. Kassam Z. Fecal Transplant via Retention Enema for Refractory or Recurrent *Clostridium difficile* Infection. *Arch Intern Med*. 2012;172(2):191. doi:10.1001/archinte.172.2.191.
39. Aas J, Gessert CE, Bakken JS. Recurrent *Clostridium difficile* Colitis: Case Series Involving 18 Patients Treated with Donor Stool Administered via a Nasogastric Tube. *Clin Infect Dis*. 2003;36(5):580-585. doi:10.1086/367657.
40. Garborg K, Waagsbø B, Stallemo A, Matre J, Sundøy A. Results of faecal donor instillation therapy for recurrent *Clostridium difficile*-associated diarrhoea. *Scand J Infect Dis*. 2010;42(11-12):857-861. doi:10.3109/00365548.2010.499541.
41. Hamilton MJ, Weingarden AR, Sadowsky MJ, Khoruts A. Standardized Frozen Preparation for Transplantation of Fecal Microbiota for Recurrent *Clostridium difficile* Infection. *Am J Gastroenterol*. 2012;107(5):761-767. doi:10.1038/ajg.2011.482.
42. Lee CH, Belanger JE, Kassam Z, et al. The outcome and long-term follow-up of 94 patients with recurrent and refractory *Clostridium difficile* infection using single to multiple fecal microbiota transplantation via retention enema. *Eur J Clin Microbiol Infect Dis*.

1789 2014;33(8):1425-1428. doi:10.1007/s10096-014-2088-9.

1790 43. Mattila E, Uusitalo-Seppälä R, Wuorela M, et al. Fecal transplantation, through colonoscopy,  
1791 is effective therapy for recurrent Clostridium difficile infection. *Gastroenterology*.  
1792 2012;142(3):490-496. doi:10.1053/j.gastro.2011.11.037.

1793 44. Rohlke F, Surawicz CM, Stollman N. Fecal Flora Reconstitution for Recurrent Clostridium  
1794 difficile Infection: Results and Methodology. *J Clin Gastroenterol*. 2010;44(8):567-570.  
1795 doi:10.1097/MCG.0b013e3181dadab10.

1796 45. Rubin TA, Gessert CE, Aas J, Bakken JS. Fecal microbiome transplantation for recurrent  
1797 Clostridium difficile infection: Report on a case series. *Anaerobe*. 2013;19:22-26.  
1798 doi:10.1016/j.anaerobe.2012.11.004.

1799 46. Patel NC, Griesbach CL, DiBaise JK, Orenstein R. Fecal Microbiota Transplant for Recurrent  
1800 Clostridium difficile Infection: Mayo Clinic in Arizona Experience. *Mayo Clin Proc*.  
1801 2013;88(8):799-805. doi:10.1016/j.mayocp.2013.04.022.

1802 47. Crobach MJT, Planche T, Eckert C, et al. European Society of Clinical Microbiology and  
1803 Infectious Diseases: update of the diagnostic guidance document for Clostridium difficile  
1804 infection. *Clin Microbiol Infect*. 2016;22 Suppl 4:S63-81. doi:10.1016/j.cmi.2016.03.010.

1805 48. Jackson M, Olefson S, Machan JT, Kelly CR. A High Rate of Alternative Diagnoses in Patients  
1806 Referred for Presumed Clostridium difficile Infection. *J Clin Gastroenterol*. 2016;50(9):742-  
1807 746. doi:10.1097/MCG.0000000000000447.

1808 49. Ray A, Smith R, Breaux J. Fecal Microbiota Transplantation for Clostridium difficile Infection:  
1809 The Ochsner Experience. *Ochsner J*. 2014;14(4):538-544.  
1810 <http://www.ncbi.nlm.nih.gov/pubmed/25598718>. Accessed October 9, 2017.

1811 50. Kao, D., Roach B., Hotte, N., Silva, M., Madsen, K., Beck, P., Louie T, Canadian Association for  
1812 the Study of the Liver. A Prospective, Dual Center, Randomized Trial Comparing Colonoscopy  
1813 versus Capsule Delivered Fecal Microbiota Transplantation (FMT) in the Management of  
1814 Recurrent Clostridium Difficile Infection (RCDI). In: *Canadian Journal of Gastroenterology and*  
1815 *Hepatology*. Vol 2016. Hindawi; 2016:1-204. doi:10.1155/2016/4792898.

1816 51. MacConnachie AA, Fox R, Kennedy DR, Seaton RA. Faecal transplant for recurrent Clostridium  
1817 difficile-associated diarrhoea: a UK case series. *QJM*. 2009;102(11):781-784.  
1818 doi:10.1093/qjmed/hcp118.

1819 52. Kelly CR, de Leon L, Jasutkar N. Fecal Microbiota Transplantation for Relapsing Clostridium

HIS/ BSG FMT Guideline: Main Document, Gut version.

- 1820 difficile Infection in 26 Patients. *J Clin Gastroenterol*. 2012;46(2):145-149.
- 1821 doi:10.1097/MCG.0b013e318234570b.
- 1822 53. Brandt LJ, Aroniadis OC, Mellow M, et al. Long-Term Follow-Up of Colonoscopic Fecal
- 1823 Microbiota Transplant for Recurrent Clostridium difficile Infection. *Am J Gastroenterol*.
- 1824 2012;107(7):1079-1087. doi:10.1038/ajg.2012.60.
- 1825 54. Pathak R, Enuh HA, Patel A, Wickremesinghe P. Treatment of relapsing Clostridium difficile
- 1826 infection using fecal microbiota transplantation. *Clin Exp Gastroenterol*. 2013;7:1-6.
- 1827 doi:10.2147/CEG.S53410.
- 1828 55. Agrawal M, Aroniadis OC, Brandt LJ, et al. The Long-term Efficacy and Safety of Fecal
- 1829 Microbiota Transplant for Recurrent, Severe, and Complicated Clostridium difficile Infection
- 1830 in 146 Elderly Individuals. *J Clin Gastroenterol*. 2015;50(5):1.
- 1831 doi:10.1097/MCG.0000000000000410.
- 1832 56. Fischer M, Kao D, Kelly C, et al. Fecal Microbiota Transplantation is Safe and Efficacious for
- 1833 Recurrent or Refractory Clostridium difficile Infection in Patients with Inflammatory Bowel
- 1834 Disease. *Inflamm Bowel Dis*. 2016;22(10):2402-2409. doi:10.1097/MIB.0000000000000908.
- 1835 57. Aroniadis OC, Brandt LJ, Greenberg A, et al. Long-term Follow-up Study of Fecal Microbiota
- 1836 Transplantation for Severe and/or Complicated Clostridium difficile Infection. *J Clin*
- 1837 *Gastroenterol*. 2015;50(5):1. doi:10.1097/MCG.0000000000000374.
- 1838 58. Fischer M, Kao D, Mehta SR, et al. Predictors of Early Failure After Fecal Microbiota
- 1839 Transplantation for the Therapy of Clostridium Difficile Infection: A Multicenter Study. *Am J*
- 1840 *Gastroenterol*. 2016;111(7):1024-1031. doi:10.1038/ajg.2016.180.
- 1841 59. Ianiro G, Valerio L, Masucci L, et al. Predictors of failure after single faecal microbiota
- 1842 transplantation in patients with recurrent Clostridium difficile infection: results from a 3-year,
- 1843 single-centre cohort study. *Clin Microbiol Infect*. 2017;23(5):337.e1-337.e3.
- 1844 doi:10.1016/j.cmi.2016.12.025.
- 1845 60. Kelly CR, Ihunnah C, Fischer M, et al. Fecal microbiota transplant for treatment of Clostridium
- 1846 difficile infection in immunocompromised patients. *Am J Gastroenterol*. 2014;109(7):1065-
- 1847 1071. doi:10.1038/ajg.2014.133.
- 1848 61. Lagier J-C, Delord M, Million M, et al. Dramatic reduction in Clostridium difficile ribotype 027-
- 1849 associated mortality with early fecal transplantation by the nasogastric route: a preliminary
- 1850 report. *Eur J Clin Microbiol Infect Dis*. 2015;34(8):1597-1601. doi:10.1007/s10096-015-2394-x.



HIS/ BSG FMT Guideline: Main Document, *Gut* version.

- 1851 62. Camacho-Ortiz A, Gutiérrez-Delgado EM, Garcia-Mazcorro JF, et al. Randomized clinical trial  
1852 to evaluate the effect of fecal microbiota transplant for initial *Clostridium difficile* infection in  
1853 intestinal microbiome. Green J, ed. *PLoS One*. 2017;12(12):e0189768.  
1854 doi:10.1371/journal.pone.0189768.
- 1855 63. Louie TJ, Miller MA, Mullane KM, et al. Fidaxomicin versus Vancomycin for *Clostridium*  
1856 *difficile* Infection. *N Engl J Med*. 2011;364(5):422-431. doi:10.1056/NEJMoa0910812.
- 1857 64. Wilcox MH, Gerding DN, Poxton IR, et al. Bezlotoxumab for Prevention of Recurrent  
1858 *Clostridium difficile* Infection. *N Engl J Med*. 2017;376(4):305-317.  
1859 doi:10.1056/NEJMoa1602615.
- 1860 65. Guery B, Menichetti F, Anttila V-J, et al. Extended-pulsed fidaxomicin versus vancomycin for  
1861 *Clostridium difficile* infection in patients 60 years and older (EXTEND): a randomised,  
1862 controlled, open-label, phase 3b/4 trial. *Lancet Infect Dis*. December 2017.  
1863 doi:10.1016/S1473-3099(17)30751-X.
- 1864 66. McFarland L V., Elmer GW, Surawicz CM. Breaking the cycle: treatment strategies for 163  
1865 cases of recurrent *Clostridium difficile* disease. *Am J Gastroenterol*. 2002;97(7):1769-1775.  
1866 doi:10.1111/j.1572-0241.2002.05839.x.
- 1867 67. Sirbu BD, Soriano MM, Manzo C, Lum J, Gerding DN, Johnson S. Vancomycin Taper and Pulse  
1868 Regimen With Careful Follow-up for Patients With Recurrent *Clostridium difficile* Infection.  
1869 *Clin Infect Dis*. 2017;65(8):1396-1399. doi:10.1093/cid/cix529.
- 1870 68. Gentry CA, Giancola SE, Thind S, Kurdgelashvili G, Skrepnek GH, Williams RJ. A Propensity-  
1871 Matched Analysis Between Standard Versus Tapered Oral Vancomycin Courses for the  
1872 Management of Recurrent *Clostridium difficile* Infection. *Open Forum Infect Dis*.  
1873 2017;4(4):ofx235. doi:10.1093/ofid/ofx235.
- 1874 69. Cornely OA, Crook DW, Esposito R, et al. Fidaxomicin versus vancomycin for infection with  
1875 *Clostridium difficile* in Europe, Canada, and the USA: a double-blind, non-inferiority,  
1876 randomised controlled trial. *Lancet Infect Dis*. 2012;12(4):281-289. doi:10.1016/S1473-  
1877 3099(11)70374-7.
- 1878 70. Tauxe WM, Haydek JP, Rebolledo PA, et al. Fecal microbiota transplant for *Clostridium*  
1879 *difficile* infection in older adults. *Therap Adv Gastroenterol*. 2016;9(3):273-281.  
1880 doi:10.1177/1756283X15622600.
- 1881 71. Khan MA, Sofi AA, Ahmad U, et al. Efficacy and safety of, and patient satisfaction with,

HIS/ BSG FMT Guideline: Main Document, *Gut* version.

- 1882 colonoscopic-administered fecal microbiota transplantation in relapsing and refractory
- 1883 community- and hospital-acquired *Clostridium difficile* infection. *Can J Gastroenterol Hepatol*.
- 1884 2014;28(8):434-438. <http://www.ncbi.nlm.nih.gov/pubmed/25014180>. Accessed October 9,
- 1885 2017.
- 1886 72. Youngster I, Russell GH, Pindar C, Ziv-Baran T, Sauk J, Hohmann EL. Oral, Capsulized, Frozen
- 1887 Fecal Microbiota Transplantation for Relapsing *Clostridium difficile* Infection. *JAMA*.
- 1888 2014;312(17):1772. doi:10.1001/jama.2014.13875.
- 1889 73. Fischer M, Sipe BW, Rogers NA, et al. Faecal microbiota transplantation plus selected use of
- 1890 vancomycin for severe-complicated *Clostridium difficile* infection: description of a protocol
- 1891 with high success rate. *Aliment Pharmacol Ther*. 2015;42(4):470-476. doi:10.1111/apt.13290.
- 1892 74. Costello SP, Conlon MA, Vuaran MS, Roberts-Thomson IC, Andrews JM. Faecal microbiota
- 1893 transplant for recurrent *Clostridium difficile* infection using long-term frozen stool is
- 1894 effective: Clinical efficacy and bacterial viability data. *Aliment Pharmacol Ther*.
- 1895 2015;42(8):1011-1018. doi:10.1111/apt.13366.
- 1896 75. Hui J, Kench JG, Chitturi S, et al. Long-Term outcomes of cirrhosis in nonalcoholic
- 1897 steatohepatitis compared with hepatitis C. *Hepatology*. 2003;38(2):420-427.
- 1898 doi:10.1053/jhep.2003.50320.
- 1899 76. Allegretti JR, Korzenik JR, Hamilton MJ. Fecal microbiota transplantation via colonoscopy for
- 1900 recurrent *C. difficile* Infection. *J Vis Exp*. 2014;(94). doi:10.3791/52154.
- 1901 77. van Beurden YH, de Groot PF, van Nood E, Nieuwdorp M, Keller JJ, Goorhuis A. Complications,
- 1902 effectiveness, and long term follow-up of fecal microbiota transfer by nasoduodenal tube for
- 1903 treatment of recurrent *Clostridium difficile* infection. *United Eur Gastroenterol J*.
- 1904 2017;5(6):868-879. doi:10.1177/2050640616678099.
- 1905 78. Bakken JS, Borody T, Brandt LJ, et al. Treating *Clostridium difficile* Infection With Fecal
- 1906 Microbiota Transplantation. *Clin Gastroenterol Hepatol*. 2011;9(12):1044-1049.
- 1907 doi:10.1016/j.cgh.2011.08.014.
- 1908 79. Allegretti JR, Allegretti AS, Phelps E, Xu H, Kassam Z, Fischer M. Asymptomatic *Clostridium*
- 1909 *difficile* carriage rate post-fecal microbiota transplant is low: a prospective clinical and stool
- 1910 assessment. *Clin Microbiol Infect*. November 2017. doi:10.1016/J.CMI.2017.10.022.
- 1911 80. Baxter M, Colville A. Adverse events in faecal microbiota transplant: a review of the
- 1912 literature. *J Hosp Infect*. 2016;92(2):117-127. doi:10.1016/j.jhin.2015.10.024.

1  
2  
3 1913 81. Hefazi M, Patnaik MM, Hogan WJ, Litzow MR, Pardi DS, Khanna S. Safety and Efficacy of Fecal  
4 1914 Microbiota Transplant for Recurrent Clostridium difficile Infection in Patients With Cancer  
5 1915 Treated With Cytotoxic Chemotherapy: A Single-Institution Retrospective Case Series. *Mayo*  
6 1916 *Clin Proc.* 2017;92(11):1617-1624. doi:10.1016/j.mayocp.2017.08.016.  
7  
8  
9  
10 1917 82. Qazi T, Amaratunga T, Barnes EL, Fischer M, Kassam Z, Allegretti JR. The risk of inflammatory  
11 1918 bowel disease flares after fecal microbiota transplantation: Systematic review and meta-  
12 1919 analysis. *Gut Microbes.* July 2017:1-15. doi:10.1080/19490976.2017.1353848.  
13  
14  
15 1920 83. Meighani A, Hart BR, Mittal C, Miller N, John A, Ramesh M. Predictors of fecal transplant  
16 1921 failure. *Eur J Gastroenterol Hepatol.* 28:826-830. doi:10.1097/MEG.0000000000000614.  
17  
18  
19 1922 84. Alrabaa S, Jariwala R, Zeitler K, Montero J. Fecal microbiota transplantation outcomes in  
20 1923 immunocompetent and immunocompromised patients: A single-center experience. *Transpl*  
21 1924 *Infect Dis.* 2017;19(4):e12726. doi:10.1111/tid.12726.  
22  
23  
24 1925 85. Cohen NA, Livovsky DM, Yaakobovitch S, et al. A Retrospective Comparison of Fecal Microbial  
25 1926 Transplantation Methods for Recurrent Clostridium Difficile Infection. *Isr Med Assoc J.*  
26 1927 2016;18(10):594-599.  
27  
28  
29 1928 86. Orenstein R, Dubberke E, Hardi R, et al. Safety and Durability of RBX2660 (Microbiota  
30 1929 Suspension) for Recurrent *Clostridium difficile* Infection: Results of the PUNCH CD Study. *Clin*  
31 1930 *Infect Dis.* 2016;62(5):596-602. doi:10.1093/cid/civ938.  
32  
33  
34 1931 87. Hirsch BE, Saraiya N, Poeth K, Schwartz RM, Epstein ME, Honig G. Effectiveness of fecal-  
35 1932 derived microbiota transfer using orally administered capsules for recurrent Clostridium  
36 1933 difficile infection. *BMC Infect Dis.* 2015;15(1):191. doi:10.1186/s12879-015-0930-z.  
37  
38  
39 1934 88. Kao D, Roach B, Beck P, Hotte N, Madsen K, Louie T. A dual center, randomized trial  
40 1935 comparing colonoscopy and oral capsule delivered fecal microbiota transplantation in the  
41 1936 treatment of recurrent clostridium difficile infection: Preliminary results. *Am J Gastroenterol.*  
42 1937 2015;110:S553. doi:http://dx.doi.org/10.1038/ajg.2015.294.  
43  
44  
45 1938 89. Youngster I, Mahabamunuge J, Systrom HK, et al. Oral, frozen fecal microbiota transplant  
46 1939 (FMT) capsules for recurrent Clostridium difficile infection. *BMC Med.* 2016;14(1):134.  
47 1940 doi:10.1186/s12916-016-0680-9.  
48  
49  
50 1941 90. Anand R, Song Y, Garg S, et al. Effect of Aging on the Composition of Fecal Microbiota in  
51 1942 Donors for FMT and Its Impact on Clinical Outcomes. *Dig Dis Sci.* 2017;62(4):1002-1008.  
52 1943 doi:10.1007/s10620-017-4449-6.  
53  
54  
55  
56  
57  
58  
59  
60

HIS/ BSG FMT Guideline: Main Document, *Gut* version.

- 1944 91. Alang N, Kelly CR. Weight gain after fecal microbiota transplantation. *Open forum Infect Dis.*  
1945 2015;2(1):ofv004. doi:10.1093/ofid/ofv004.
- 1946 92. Fischer M, Kao D, Kassam Z, et al. Stool Donor Body Mass Index Does Not Affect Recipient  
1947 Weight After a Single Fecal Microbiota Transplantation for *C. difficile* Infection. *Clin*  
1948 *Gastroenterol Hepatol.* December 2017. doi:10.1016/J.CGH.2017.12.007.
- 1949 93. Marchesi JR, Adams DH, Fava F, et al. The gut microbiota and host health: a new clinical  
1950 frontier. *Gut.* 2016;65(2):330-339. doi:10.1136/gutjnl-2015-309990.
- 1951 94. Allegretti JR, Fischer M, Papa E, et al. Su1738 Fecal Microbiota Transplantation Delivered via  
1952 Oral Capsules Achieves Microbial Engraftment Similar to Traditional Delivery Modalities:  
1953 Safety, Efficacy and Engraftment Results From a Multi-Center Cluster Randomized Dose-  
1954 Finding Study. *Gastroenterology.* 2016;150(4):S540. doi:10.1016/S0016-5085(16)31855-8.
- 1955 95. Langdon A, Crook N, Dantas G. The effects of antibiotics on the microbiome throughout  
1956 development and alternative approaches for therapeutic modulation. *Genome Med.*  
1957 2016;8(1):39. doi:10.1186/s13073-016-0294-z.
- 1958 96. Lange K, Buerger M, Stallmach A, Bruns T. Effects of Antibiotics on Gut Microbiota. *Dig Dis.*  
1959 2016;34(3):260-268. doi:10.1159/000443360.
- 1960 97. Becattini S, Taur Y, Pamer EG. Antibiotic-Induced Changes in the Intestinal Microbiota and  
1961 Disease. *Trends Mol Med.* 2016;22(6):458-478. doi:10.1016/j.molmed.2016.04.003.
- 1962 98. Ianiro G, Tilg H, Gasbarrini A. Antibiotics as deep modulators of gut microbiota: between  
1963 good and evil. *Gut.* 2016;65(11):1906-1915. doi:10.1136/gutjnl-2016-312297.
- 1964 99. Boostrom SY, Mathis KL, Pendlimari R, et al. Burden of *Clostridium difficile* on the healthcare  
1965 system. *Inflamm Bowel Dis.* 2012;18(1):994-1002. doi:10.1126/science.1241214.
- 1966 100. J.R. A, J.R. K, M.J. H. Intestinal microbiome restoration for recurrent *clostridium difficile*  
1967 infection in patients with concurrent inflammatory bowel disease. *Gastroenterology.*  
1968 2015;148(4 SUPPL. 1):S869.
- 1969 101. National Health Service. Population screening programmes: NHS bowel cancer screening  
1970 (BCSP) programme - GOV.UK. [https://www.gov.uk/topic/population-screening-](https://www.gov.uk/topic/population-screening-programmes/bowel)  
1971 [programmes/bowel](https://www.gov.uk/topic/population-screening-programmes/bowel). Accessed June 10, 2018.
- 1972 102. London: TSO Guidelines for the Blood Transfusion Services in the United Kingdom 7 th Edition  
1973 2005 TSO Accredited Agents Web Access.

1  
2  
3 1974 103. Emanuelsson F, Claesson BEB, Ljungström L, Tvede M, Ung K-A. Faecal microbiota  
4 1975 transplantation and bacteriotherapy for recurrent Clostridium difficile infection: A  
5 1976 retrospective evaluation of 31 patients. *Scand J Infect Dis*. 2014;46(2):89-97.  
6 1977 doi:10.3109/00365548.2013.858181.  
7  
8  
9  
10 1978 104. Endtz HP, van den Braak N, van Belkum A, et al. Fecal carriage of vancomycin-resistant  
11 1979 enterococci in hospitalized patients and those living in the community in The Netherlands. *J*  
12 1980 *Clin Microbiol*. 1997;35(12):3026-3031. <http://www.ncbi.nlm.nih.gov/pubmed/9399488>.  
13 1981 Accessed February 15, 2018.  
14  
15  
16 1982 105. Willems RJL, Top J, van Santen M, et al. Global spread of vancomycin-resistant Enterococcus  
17 1983 faecium from distinct nosocomial genetic complex. *Emerg Infect Dis*. 2005;11(6):821-828.  
18 1984 doi:10.3201/eid1106.041204.  
19  
20  
21  
22 1985 106. Gough E, Shaikh H, Manges AR. Systematic Review of Intestinal Microbiota Transplantation  
23 1986 (Fecal Bacteriotherapy) for Recurrent Clostridium difficile Infection. *Clin Infect Dis*.  
24 1987 2011;53(10):994-1002. doi:10.1093/cid/cir632.  
25  
26  
27 1988 107. Casewell MW, Slater NGP, Cooper JE. Operating theatre water-baths as a cause of  
28 1989 pseudomonas septicaemia. *J Hosp Infect*. 1981;2:237-240. doi:10.1016/0195-6701(81)90043-  
29 1990 8.  
30  
31  
32  
33 1991 108. Muyltermans G, de Smet F, Pierard D, et al. Neonatal infections with Pseudomonas  
34 1992 aeruginosa associated with a water-bath used to thaw fresh frozen plasma. *J Hosp Infect*.  
35 1993 1998;39(4):309-314. <http://www.ncbi.nlm.nih.gov/pubmed/9749402>. Accessed June 11,  
36 1994 2018.  
37  
38  
39  
40 1995 109. Sleight SC, Wigginton NS, Lenski RE. Increased susceptibility to repeated freeze-thaw cycles in  
41 1996 Escherichia coli following long-term evolution in a benign environment. *BMC Evol Biol*.  
42 1997 2006;6(1):104. doi:10.1186/1471-2148-6-104.  
43  
44  
45 1998 110. O'Brien CL, Allison GE, Grimpen F, Pavli P. Impact of colonoscopy bowel preparation on  
46 1999 intestinal microbiota. *PLoS One*. 2013;8(5):e62815. doi:10.1371/journal.pone.0062815.  
47  
48  
49 2000 111. Mai V, Stine OC. Bowel preparation for colonoscopy: relevant for the gut's microbiota? *Gut*.  
50 2001 2015;64(10):1504-1505. doi:10.1136/gutjnl-2014-308937.  
51  
52  
53 2002 112. Jalanka J, Salonen A, Salojärvi J, et al. Effects of bowel cleansing on the intestinal microbiota.  
54 2003 *Gut*. 2015;64(10):1562-1568. doi:10.1136/gutjnl-2014-307240.  
55  
56 2004 113. Mai V, Greenwald B, Glenn Morris J, Raufman J-P, Stine OC. Effect of bowel preparation and  
57  
58  
59  
60

HIS/ BSG FMT Guideline: Main Document, *Gut* version.

- 2005 colonoscopy on post-procedure intestinal microbiota composition. *Gut*. 2006;55(12):1822-
- 2006 1823. doi:10.1136/gut.2006.108266.
- 2007 114. Harrell L, Wang Y, Antonopoulos D, et al. Standard Colonic Lavage Alters the Natural State of
- 2008 Mucosal-Associated Microbiota in the Human Colon. Singh SR, ed. *PLoS One*.
- 2009 2012;7(2):e32545. doi:10.1371/journal.pone.0032545.
- 2010 115. Chin SM, Sauk J, Mahabamunuge J, Kaplan JL, Hohmann EL, Khalili H. Fecal Microbiota
- 2011 Transplantation for Recurrent *Clostridium difficile* Infection in Patients With Inflammatory
- 2012 Bowel Disease: A Single-Center Experience. *Clin Gastroenterol Hepatol*. 2017;15(4):597-599.
- 2013 doi:10.1016/j.cgh.2016.11.028.
- 2014 116. Wang S, Xu M, Wang W, et al. Systematic Review: Adverse Events of Fecal Microbiota
- 2015 Transplantation. *PLoS One*. 2016;11(8):e0161174. doi:10.1371/journal.pone.0161174.
- 2016 117. Khoruts A, Rank KM, Newman KM, et al. Inflammatory Bowel Disease Affects the Outcome of
- 2017 Fecal Microbiota Transplantation for Recurrent *Clostridium difficile* Infection. *Clin*
- 2018 *Gastroenterol Hepatol*. 2016;14(10):1433-1438. doi:10.1016/j.cgh.2016.02.018.
- 2019 118. de Jager CPC, Wever PC, Gemen EFA, et al. Proton pump inhibitor therapy predisposes to
- 2020 community-acquired *Streptococcus pneumoniae* pneumonia. *Aliment Pharmacol Ther*.
- 2021 2012;36(10):941-949. doi:10.1111/apt.12069.
- 2022 119. Imhann F, Bonder MJ, Vich Vila A, et al. Proton pump inhibitors affect the gut microbiome.
- 2023 *Gut*. 2016;65(5):740-748. doi:10.1136/gutjnl-2015-310376.
- 2024 120. McDonald EG, Milligan J, Frenette C, Lee TC. Continuous Proton Pump Inhibitor Therapy and
- 2025 the Associated Risk of Recurrent *Clostridium difficile* Infection. *JAMA Intern Med*.
- 2026 2015;175(5):784. doi:10.1001/jamainternmed.2015.42.
- 2027 121. Janarthanan S, Ditah I, Adler DG, Ehrinpreis MN. *Clostridium difficile*-Associated Diarrhea and
- 2028 Proton Pump Inhibitor Therapy: A Meta-Analysis. *Am J Gastroenterol*. 2012;107(7):1001-
- 2029 1010. doi:10.1038/ajg.2012.179.
- 2030 122. Girotra M, Garg S, Anand R, Song Y, Dutta SK. Fecal Microbiota Transplantation for Recurrent
- 2031 *Clostridium difficile* Infection in the Elderly: Long-Term Outcomes and Microbiota Changes.
- 2032 *Dig Dis Sci*. 2016;61(10):3007-3015. doi:10.1007/s10620-016-4229-8.
- 2033 123. Hagel S, Fischer A, Ehlermann P, et al. Fecal Microbiota Transplant in Patients With Recurrent
- 2034 *Clostridium Difficile* Infection. *Dtsch Arztebl Int*. 2016;113(35-36):583-589.
- 2035 doi:10.3238/arztebl.2016.0583.



HIS/ BSG FMT Guideline: Main Document, *Gut* version.

- 2036 124. Department of Health (UK). Management and decontamination of flexible endoscopes (HTM  
2037 01-06) - GOV.UK. [https://www.gov.uk/government/publications/management-and-](https://www.gov.uk/government/publications/management-and-decontamination-of-flexible-endoscopes)  
2038 decontamination-of-flexible-endoscopes. Accessed December 19, 2017.
- 2039 125. British Society of Gastroenterology. Guidance on Decontamination of Equipment for  
2040 Gastrointestinal Endoscopy: 2017 Edition. [https://www.bsg.org.uk/resource/guidance-on-](https://www.bsg.org.uk/resource/guidance-on-decontamination-of-equipment-for-gastrointestinal-endoscopy-2017-edition.html)  
2041 decontamination-of-equipment-for-gastrointestinal-endoscopy-2017-edition.html. Accessed  
2042 December 19, 2017.
- 2043 126. Loveday HP, Wilson JA, Pratt RJ, et al. epic3: National Evidence-Based Guidelines for  
2044 Preventing Healthcare-Associated Infections in NHS Hospitals in England. *J Hosp Infect*.  
2045 2014;86(1):S1-S70. doi:10.1016/S0195-6701(13)60012-2.
- 2046 127. Kassam Z, Lee CH, Yuan Y, Hunt RH. Fecal Microbiota Transplantation for *Clostridium difficile*  
2047 Infection: Systematic Review and Meta-Analysis. *Am J Gastroenterol*. 2013;108(4):500-508.  
2048 doi:10.1038/ajg.2013.59.
- 2049 128. Allegretti JR, Kao D, Sitko J, Fischer M, Kassam Z. Early antibiotic use post-fecal microbiota  
2050 transplantation increases the risk of treatment failure. *Clin Infect Dis*. August 2017.  
2051 doi:10.1093/cid/cix684.
- 2052 129. Terveer EM, van Beurden YH, Goorhuis A, et al. How to: Establish and run a stool bank. *Clin*  
2053 *Microbiol Infect*. 2017;23(12):924-930. doi:10.1016/j.cmi.2017.05.015.
- 2054 130. Quraishi MN, Widlak M, Bhala N, et al. Systematic review with meta-analysis: the efficacy of  
2055 faecal microbiota transplantation for the treatment of recurrent and refractory *Clostridium*  
2056 *difficile* infection. *Aliment Pharmacol Ther*. 2017;46(5):479-493. doi:10.1111/apt.14201.
- 2057 131. Cammarota G, Ianiro G, Gasbarrini A. Fecal microbiota transplantation for the treatment of  
2058 *Clostridium difficile* infection: a systematic review. *J Clin Gastroenterol*. 2014;48(8):693-702.  
2059 doi:10.1097/MCG.000000000000046.
- 2060 132. D. D, J. R, S. G, et al. Fecal microbiota transplantation for *clostridium difficile* infection a  
2061 systematic review. *Ann Intern Med*. 2015;162(9):630-638.
- 2062 133. Baxter M, Ahmad T, Colville A, Sheridan R. Fatal Aspiration Pneumonia as a Complication of  
2063 Fecal Microbiota Transplant. *Clin Infect Dis*. 2015;61(1):136-137. doi:10.1093/cid/civ247.
- 2064 134. Hecker MT, Obrenovich ME, Cadnum JL, et al. Fecal Microbiota Transplantation by Freeze-  
2065 Dried Oral Capsules for Recurrent *Clostridium difficile* Infection. *Open forum Infect Dis*.  
2066 2016;3(2):ofw091. doi:10.1093/ofid/ofw091.



HIS/ BSG FMT Guideline: Main Document, *Gut* version.

- 2067 135. Ni J, Wu GD, Albenberg L, Tomov VT. Gut microbiota and IBD: causation or correlation? *Nat*  
2068 *Rev Gastroenterol Hepatol.* 2017;14(10):573-584. doi:10.1038/nrgastro.2017.88.
- 2069 136. Rossen NG, Fuentes S, van der Spek MJ, et al. Findings From a Randomized Controlled Trial of  
2070 Fecal Transplantation for Patients With Ulcerative Colitis. *Gastroenterology.* 2015;149(1):110-  
2071 118.e4. doi:10.1053/j.gastro.2015.03.045.
- 2072 137. Moayyedi P, Surette MG, Kim PT, et al. Fecal Microbiota Transplantation Induces Remission in  
2073 Patients With Active Ulcerative Colitis in a Randomized Controlled Trial. *Gastroenterology.*  
2074 2015;149(1):102-109.e6. doi:10.1053/j.gastro.2015.04.001.
- 2075 138. Paramsothy S, Kamm MA, Kaakoush NO, et al. Multidonor intensive faecal microbiota  
2076 transplantation for active ulcerative colitis: a randomised placebo-controlled trial. *Lancet*  
2077 *(London, England).* 2017;389(10075):1218-1228. doi:10.1016/S0140-6736(17)30182-4.
- 2078 139. Costello S, Waters O, Bryant R. Short Duration, Low Intensity, Pooled Fecal Microbiota  
2079 Transplantation Induces Remission in Patients with Mild-Moderately Active Ulcerative Colitis:  
2080 A Randomised Controlled Trial. (Abstract). *Gastroenterology.* 152(5):S198-S199.
- 2081 140. Johnsen PH, Hilpüsch F, Cavanagh JP, et al. Faecal microbiota transplantation versus placebo  
2082 for moderate-to-severe irritable bowel syndrome: a double-blind, randomised, placebo-  
2083 controlled, parallel-group, single-centre trial. *lancet Gastroenterol Hepatol.* 2018;3(1):17-24.  
2084 doi:10.1016/S2468-1253(17)30338-2.
- 2085 141. Tian H, Ge X, Nie Y, et al. Fecal microbiota transplantation in patients with slow-transit  
2086 constipation: A randomized, clinical trial. Green J, ed. *PLoS One.* 2017;12(2):e0171308.  
2087 doi:10.1371/journal.pone.0171308.
- 2088 142. Bajaj JS, Kassam Z, Fagan A, et al. Fecal Microbiota Transplant from a Rational Stool Donor  
2089 Improves Hepatic Encephalopathy: A Randomized Clinical Trial. *Hepatology.* June 2017.  
2090 doi:10.1002/hep.29306.
- 2091 143. Vrieze A, Van Nood E, Holleman F, et al. Transfer of Intestinal Microbiota From Lean Donors  
2092 Increases Insulin Sensitivity in Individuals With Metabolic Syndrome. *Gastroenterology.*  
2093 2012;143(4):913-916.e7. doi:10.1053/j.gastro.2012.06.031.
- 2094 144. Kootte RS, Levin E, Salojärvi J, et al. Improvement of Insulin Sensitivity after Lean Donor Feces  
2095 in Metabolic Syndrome Is Driven by Baseline Intestinal Microbiota Composition. *Cell Metab.*  
2096 2017;26(4):611-619.e6. doi:10.1016/j.cmet.2017.09.008.
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**15. Figure legends and tables:**

**Figure 1: Proposed summary pathway for donor screening for centres preparing frozen FMT from recurring donors.**

**Table 1: Recommended donor history/ questionnaire:** A positive response to any of these questions would usually result in exclusion from further consideration as a donor, although this would depend upon the particular circumstances/ answers given.

- |      |   |
|------|---|
| 2125 | 1. Receipt of antimicrobials within the past three months.  |
| 2126 | 2. Known prior exposure to HIV and/ or viral hepatitis, and known previous or latent tuberculosis.  |
| 2127 |   |
| 2128 | 3. Risk factors for blood-borne viruses - including high risk sexual behaviours, use of illicit drugs, any tattoo/ body piercing/ needlestick injury/ blood transfusion/ acupuncture, all within the previous six months.   |
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| 2131 | 4. Receipt of a live attenuated virus within the past six months.   |
| 2132 | 5. Underlying gastrointestinal conditions/ symptoms (e.g. history of IBD, IBS, chronic diarrhoea, chronic constipation, coeliac disease, bowel resection or bariatric surgery) - also including acute diarrhoea/ gastrointestinal symptoms within the past two weeks. |
| 2133 |   |
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| 2135 | 6. Family history of any significant gastrointestinal conditions (e.g. family history of IBD, or colorectal cancer).  |
| 2136 |   |
| 2137 | 7. History of atopy (e.g. asthma, eosinophilic disorders).  |
| 2138 | 8. Any systemic autoimmune conditions.  |
| 2139 | 9. Any metabolic conditions, including diabetes and obesity.  |
| 2140 | 10. Any neurological or psychiatric conditions, or known risk of prion disease.   |
| 2141 | 11. History of chronic pain syndromes, including chronic fatigue syndrome and fibromyalgia.   |
| 2142 | 12. History of any malignancy.  |
| 2143 | 13. Taking particular regular medications, or such medications within the past three months, i.e. antimicrobials, proton pump inhibitors, immunosuppression, chemotherapy   |
| 2144 |   |
| 2145 | 14. History of receiving growth hormone, insulin from cows, or clotting factor concentrates.  |
| 2146 | 15. History of receiving an experimental medicine or vaccine within the past six months.  |
| 2147 | 16. History of travel to tropical countries within the past six months.   |

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2150 **Table 2: Recommended blood screening for stool donors:** \*EBV and CMV testing is only  
 2151 recommended where there is the potential that the FMT prepared from that donor will be  
 2152 administered to immunosuppressed patients at risk of severe infection if exposed to CMV and EBV.

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3	<i>Pathogen screening:</i>
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5	• Hepatitis A IgM
6	• Hepatitis B (HBsAg and HBcAb)
7	• Hepatitis C antibody
8	• Hepatitis E IgM
9	• HIV -1 and -2 antibodies
10	• HTLV-1 and -2 antibodies
11	• <i>Treponema pallidum</i> antibodies (TPHA, VDRL)
12	• Epstein-Barr virus IgM and IgG*
13	• Cytomegalovirus IgM and IgG*
14	• <i>Strongyloides stercoralis</i> IgG
15	• <i>Entamoeba histolytica</i> serology
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17	<i>General/ metabolic screening:</i>
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19	• Full blood count with differential.
20	• Creatinine and electrolytes
21	• Liver enzymes (including albumin, bilirubin, aminotransferases, gamma-glutamyltransferase
22	and alkaline phosphatase).
23	• C-reactive protein
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**Table 3: Recommended stool screening for stool donors:** \*Whilst CPE and ESBL are the only multi-drug resistant bacteria that are recommended to be screened for universally, consider testing for other resistant organisms (including vancomycin-resistant *Enterococci* (VRE) and/ or methicillin-resistant *Staphylococcus aureus* (MRSA)) based upon risk assessment and local prevalence.

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- *Clostridium difficile* PCR
- *Campylobacter*, *Salmonella*, and *Shigella* by standard stool culture and/ or PCR
- Shiga toxin-producing *Escherichia coli* by PCR.
- Multi-drug resistant bacteria, at least carbapenemase-producing *Enterobacteriaceae* (CPE) and extended-spectrum beta-lactamases (ESBL)\*.
- Stool ova, cysts and parasite analysis, including for *Microsporidia*.
- Faecal antigen for *Cryptosporidium* and *Giardia*.
- Acid fast stain for *Cyclospora* and *Isospora*.
- *Helicobacter pylori* faecal antigen.
- Norovirus, Rotavirus PCR.

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2179 **Table 4: A summary of the GRADE system:**

<b>GRADE - strength of evidence:</b>	<b>GRADE - strength of recommendation:</b>
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<i>High quality:</i> Further research is very unlikely to change our confidence in the estimate of effect.	<i>The trade-offs:</i> Taking into account the estimate size of the effect for main outcomes, the confidence limits around those estimates and the relative value placed on each outcome.
<i>Moderate quality:</i> Further research is likely to have an important impact on our confidence in the estimate of effect and may change the estimate.	<i>The quality of the evidence.</i>
<i>Low quality:</i> Further research is very likely to have an important impact on our confidence in the estimate of effect and is likely to change the estimate.	<i>Translation of the evidence into practice in a particular setting:</i> Taking into consideration important factors that could be expected to modify the size of expected effects.
<i>Very low quality:</i> Any estimate of effect is very uncertain.	<i>Uncertainty about the baseline risk for the population of interest.</i>

**Table 5: Criteria for stool collection:**

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Clear instructions should be given to donors regarding hand hygiene.
Collect stool donations in a sealable clean container. A number of specifically designed devices are available commercially.
Stool should ideally be passed directly into the clean container for collection; alternatively, it may be collected in clean tissue and transferred to the clean container.
Stool should be transported to the FMT production site as soon as possible post defaecation (and within six hours); however, if a short period of storage is necessary, this should be at 4°C.

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Supplementary Material 1 for *Gut*

**The use of faecal microbiota transplant as treatment for recurrent or refractory *Clostridium difficile* infection and other potential indications: joint British Society of Gastroenterology (BSG) and Healthcare Infection Society (HIS) guidelines.**

**Supplementary Material 1: General additional information:**

**1. Additional information:**

**1.1. Lay summary:**

Faecal microbiota transplant (FMT) involves the transfer of a sample of faeces from a healthy donor to a recipient. There are several different ways to administer the transplant, including via endoscopy, rectally as an enema, via nasogastric/ nasoenteral tube (tube passed through the nose into the stomach/ upper part of the small intestine), or via oral ingestion of capsules that contain faecal material. The transplant may either be administered fresh (i.e. immediately after preparation), or may be prepared in advance, stored in a freezer and thawed when required. FMT is an accepted and effective treatment for recurrent infection by *Clostridium difficile*, a bacterium which can cause severe illness with diarrhoea, most commonly in frail elderly populations as a complication of antibiotic use. Despite adequate treatment, *Clostridium difficile* infection recurs in about 25% of patients, and some may suffer multiple recurrences.

This guideline reviews the evidence for FMT as a treatment for *Clostridium difficile* infection (CDI) and other conditions. Recommendations are made for: which patients are most likely to benefit, how donors should be selected and screened, how FMT should be prepared and administered, how patients should be followed up, and how FMT services should be configured.

**1.2. Working Party Report**

**1.2.1. What is the Working Party Report?**

The report is a set of recommendations covering key aspects of safe and efficacious delivery of a FMT service for recurrent/ refractory *Clostridium difficile* infection (CDI). The guidelines also review the evidence for the use of FMT for non-CDI indications.

Supplementary Material 1 for *Gut*

The working group recommendations have been developed systematically through multi-disciplinary discussions based on published evidence. They should be used in the development of local protocols for all relevant healthcare settings.

**1.2.2. Why do we need a Working Party Report for this topic?**

There is widespread and growing interest in the use of FMT as a treatment for recurrent CDI. The previous absence of randomised trials and lack of evidence-based guidelines describing best practice related to its use has led to uncertainty as to how to establish an FMT service. Existing services may be providing suboptimal clinical care. There is now a developing portfolio of randomised study evidence (including randomised controlled trial data) regarding the use of FMT in CDI and non-CDI indications, providing the opportunity to develop an evidence-based guideline for its use. There have also been recent changes to the UK regulatory framework for FMT (see **Supplementary Material 3**), which are not well-understood by clinicians.

**1.2.3. What is the purpose of the Working Party Report's recommendations?**

The main purpose is to inform clinicians about the use of FMT (and about the establishment of this service) for the treatment of recurrent and refractory CDI, and other possible future indications. The recommendations provide an evidence-based approach to a high quality clinical service, with appropriate governance structures. This document also serves to illustrate areas in which there are current gaps in knowledge, which will help to direct future areas of research.

**1.2.4. Who are these guidelines for?**

Any healthcare practitioner may use these guidelines and adapt them for their use. It is anticipated that users will include clinical staff, as well as healthcare infection prevention and control teams. It is expected that these guidelines will raise awareness of FMT amongst clinicians who care for patients with recurrent or refractory CDI, but who may be unaware that it is a feasible and accessible treatment option. The guidelines are also designed to be read by patients with CDI, helping them to understand whether FMT may be an appropriate treatment option for them.

**1.2.5. How are the guidelines structured?**

Supplementary Material 1 for *Gut*

Each section comprises an introduction, a summary of the evidence base with levels, and a recommendation graded according to the available evidence.

**1.2.6. Aim**

The primary aim of this report was to assess the current evidence for all aspects relating to provision of an FMT service as treatment for recurrent or refractory CDI. A secondary aim was to review the current evidence for the efficacy of FMT in treating non-CDI conditions.

**1.3. Implementation of these guidelines:**

**1.3.1. How can these guidelines be used to improve clinical effectiveness?**

Primarily, these guidelines will inform the development of local FMT services and appropriate local operational protocols, and will guide clinical decision-making. They also provide a framework for clinical audit, a tool for improving clinical effectiveness. In addition, the future research priorities identified by the working group will allow researchers to refine applications to funding bodies.

**1.3.2. How much will it cost to implement these guidelines?**

Where FMT is being provided under a MHRA license according to Good Manufacturing Practice (GMP) standards, there are significant costs associated with initial setup and maintenance of the service. These include the cost of obtaining the relevant license, laboratory design and equipment to enable quality assurance, storage facilities for samples, etc. However, there is counterbalance to this, as the expectation of the working group is that the publication of this guideline may encourage provision of FMT as treatment for recurrent or refractory CDI. This has consistently been shown to be cost effective in comparison with anti-*C. difficile* antimicrobial therapy<sup>1-4</sup>, so overall costs associated with treating the condition may actually decrease. Furthermore, there may be changes to the practice of clinicians already offering the service. For example, encouraging the use of healthy unrelated donors (who can provide multiple stool donations after one screening) reduces the cost of screening when compared to the use of an FMT recipient's relative as donor, who is likely to provide one donation only.

**1.3.3. E-learning tools:**

Supplementary Material 1 for *Gut*

Continuing Professional Development questions and their answers are provided for self-assessment in **Appendix 4** of this document.

## 2. Appendices

### Appendix 1: Glossary

*Clostridium difficile* infection (CDI) - Symptomatic infection caused by the spore-forming, toxin-secreting bacterium, *Clostridium difficile*. It is the most common cause of antibiotic-associated diarrhoea, and symptoms include watery stools, fever, nausea, and abdominal pain.

Refractory CDI – Failure of an episode of CDI to respond to metronidazole and oral vancomycin, although no uniform definition.

Recurrent CDI – Defined in ESMID guidelines as ‘when CDI re-occurs within 8 weeks after the onset of a previous episode, provided the symptoms from the previous episode resolved after completion of initial treatment’<sup>4</sup>; however, defined more variably within the reviewed literature within this guideline.

Faecal microbiota transplant – A procedure in which faecal matter (stool) is collected from a healthy screened donor, homogenised, strained, and introduced into the gastrointestinal tract of a patient.

Donor – In the context of FMT, this is a healthy screened individual that provides stool for the use in preparation of FMT.

Nasogastric – A means of reaching/ supplying the stomach via the nose for the purpose of treatment or investigation. This is usually achieved by the insertion of a tube.

Enema – A procedure in which liquid (or gas) is infused into the rectum as means for treatment or investigation.

Gut microbiota - Population of microorganisms that live in the gastrointestinal tract including bacteria, viruses and fungi.

Inflammatory bowel disease – Describes a group of chronic disorders (ulcerative colitis and Crohn’s diseases) in which the gastrointestinal tract becomes inflamed. The exact cause is unknown but it is thought to result from a combination of factors that trigger the body’s immune system to produce an inflammatory reaction in the gastrointestinal tract.

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Medicines and Healthcare Products Regulatory Agency - An executive agency of the Department of Health in the United Kingdom which is responsible for ensuring that medicines and medical devices are efficacious and are acceptably safe.

**Appendix 2: Guideline Development**

***Introduction***

The need for a guideline within this area was agreed at a HIS guideline scoping day, and a BSG Gut Microbiota for Health (GMfH) panel teaching/ meeting day, both in September 2015, and further meetings between both bodies confirmed the establishment of a working group. Members were chosen to reflect the range of stakeholders, but were not limited to members of BSG or HIS. Feedback from the HIS guideline scoping day (including patient representatives) was used to establish a basis for PICO questions, with the final structure of PICO questions agreed collectively by teleconference in July 2017. No payment was made to anyone involved in this guideline.

***Conflict of interest***

Conflict of interest was registered from all working group members and underwent ongoing review up until the point of completion. In the event of a potential conflict being identified, the working group agreed that the member should not contribute to the section affected.

***Search Strategy & Results***

***i. Literature search strategy: PICO Review Questions:***

**Review Question 1: Which patients with *Clostridium difficile* infection should be considered for faecal microbiota transplant, and how should they be followed up after treatment?**

Populations: Adults (18 years and over) with *Clostridium difficile* infection

Intervention: Faecal microbiota transplant

Comparison: Placebo

Vancomycin

Metronidazole

Supplementary Material 1 for *Gut*

Fidaxomicin

Intravenous immunoglobulin

Bezlotoxumab

Probiotics

Cessation of antibiotics for alternative indication

Outcomes: **Critical:** Cessation of diarrhoea and other symptoms/ relapse

Quality of life

Serious adverse events

**Important:** Negative tests for *Clostridium difficile* infection

Adverse events

Study design: Randomised trials

If no randomised trials identified – prospective cohort studies and retrospective case series

**Review Question 2: What recipient factors influence the outcome of faecal microbiota transplant when treating people with *Clostridium difficile* infection?**

Populations: Adults (18 years and over) with *Clostridium difficile* infection

Intervention: Faecal microbiota transplant

Comparison: **Preparation of patient:**

Use of bowel purgatives vs no bowel purgatives

For upper GI administration - use of PPI/ acid suppression prior to procedure vs no acid suppression

Use of agents affecting GI motility (e.g. metoclopramide for upper GI/ loperamide for lower GI) vs no use

Time before procedure that anti-CDI antibiotics are used and stopped (comparing time courses)

Supplementary Material 1 for Gut

	<b>Comorbidities:</b>	
		Severe CDI/ toxic megacolon vs non-severe disease
		Co-existing inflammatory bowel disease (IBD) vs no IBD
		Immunosuppression vs no immunosuppression
		Chronic liver disease/ cirrhosis vs no chronic liver disease
Outcomes:	<b>Critical:</b>	Cessation of diarrhoea and other symptoms/ relapse
		Quality of life
		Serious adverse events
	<b>Important:</b>	Negative tests for <i>Clostridium difficile</i> infection
		Adverse events
Study design:	Randomised trials	
		If no randomised trials identified – prospective cohort studies, retrospective case series

**Review Question 3: What donor factors influence the outcome of faecal microbiota transplant when treating people with *Clostridium difficile* infection?**

Populations:	Adults (18 years and over) with <i>Clostridium difficile</i> infection
Intervention:	Faecal microbiota transplant
Comparison:	Related vs unrelated donor
	Donor working in healthcare setting vs donor not from healthcare setting
	BMI (comparing cut-offs used)
	Age (comparing ages)
	Length of time since donor had antibiotics (comparing cut-offs used)
Outcomes:	<b>Critical :</b> Cessation of diarrhoea and other symptoms/ relapse



Supplementary Material 1 for *Gut*

Quality of life

Serious adverse events

**Important:** Negative tests for *Clostridium difficile* infection

Adverse events

Study design: Randomised trials

If no randomised trials identified – prospective cohort studies and retrospective case series

**Review Question 4: What factors related to the preparation of the transplant influence the outcome of faecal microbiota transplant when treating people with *Clostridium difficile* infection?**Populations: Adults (18 years and over) with *Clostridium difficile* infection

Intervention: Faecal microbiota transplant

Comparison: Time after delivery when transplant is prepared (comparing time points)

Anaerobic preparation vs preparation in ambient air

Manual preparation vs use of blender/ homogeniser

Diluent used (comparing normal saline, phosphate-buffered saline, water, milk/ yoghurt and others)

Amount of stool/ transplant administered (comparing amounts)

Fresh preparation vs frozen preparation:

-comparing glycerol vs other cryopreservative

-comparing concentration of cryopreservative used

-comparing length of time that frozen for before use

Outcomes: **Critical:** Cessation of diarrhoea and other symptoms/ relapse

Quality of life

Serious adverse events

**Important:** Negative tests for *Clostridium difficile* infection

Supplementary Material 1 for *Gut*

Adverse events

Study design: Randomised trials

If no randomised trials identified – prospective cohort studies and retrospective case series

**Review Question 5: What factors related to administration of the transplant influence the outcome of faecal microbiota transplant when treating people with *Clostridium difficile* infection?**

Populations: Adults (18 years and over) with *Clostridium difficile* infection

Intervention: Faecal microbiota transplant

Comparison: Upper GI administration (nasogastric, nasoduodenal or nasojejunal tube; upper GI endoscopy) vs lower GI administration (enema, rectal catheter, colonoscopy)

Encapsulated vs full transplant

Outcomes: **Critical:** Cessation of diarrhoea and other symptoms/ relapse

Quality of life

Serious adverse events

**Important:** Negative tests for *Clostridium difficile* infection

Adverse events

Study design: Randomised trials

If no randomised trials identified – prospective cohort studies, and retrospective case series

**Review Question 6: What is the clinical effectiveness of faecal microbiota transplant in treating conditions other than *Clostridium difficile* infection?**

Populations: Adults (18 years and over) with conditions of interest (e.g. inflammatory bowel disease)

Intervention: Faecal microbiota transplant

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Comparison: Standard care for the condition of interest

Autologous faecal microbiota transplant

Outcomes: **Critical:** Clinical improvement

Improvement in laboratory/ radiological/ endoscopic tests

Quality of life

Serious adverse events

**Important:** Adverse events

Study design: Randomised trials

**ii. Literature search terms:**

**Review Questions 1 – 5:**

*EMBASE*

1. exp Clostridium difficile infection/ or exp Clostridium difficile toxin B/ or exp Clostridium difficile toxin A/

2. clostridium difficile.ti,ab.

3. c diff\*.ti,ab.

4. (CDAD or RCDI or CDI).ti,ab.

5. pseudomembranous.ti,ab.

6. exp pseudomembranous colitis/

7. (antibiotic\* adj2 (diarrhea or diarrhoea or colitis)).ti,ab.

8. (FMT or HPI).ti,ab.

9. ((fecal or faecal or feces or faeces or stool or microbiota) adj2 (transplant\* or infus\* or transfus\* or implant\* or instil\* or donat\* or donor\* or reconstitut\* or therap\* or bacteriotherapy or encapsulated\* or capsul\*)).ti,ab.

10. (fecal or faecal or feces or faeces or stool or microbiota).ti,ab.

11. transplant\*.ti,ab.

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12. exp transplantation/

13. 8 or 9

14. 10 and (11 or 12)

15. 13 or 14

16. or/1-7

17. 15 and 16

*MEDLINE*

1. Clostridium difficile/

2. clostridium difficile.ti,ab.

3. c diff\$.ti,ab.

4. Enterocolitis, Pseudomembranous/

5. (antibiotic\$ adj2 (diarrhoea or colitis)).ti,ab.

6. (antibiotic\$ adj2 (diarrhea or colitis)).ti,ab.

7. pseudomembranous.ti,ab.

8. (CDAD or CDI).mp. [mp=title, abstract, original title, name of substance word, subject heading word, keyword heading word, protocol supplementary concept word, rare disease supplementary concept word, unique identifier, synonyms]

9. RCDI.ti,ab.

10. Clostridium Infections/

11. FMT.mp. or HPI.ti,ab. [mp=title, abstract, original title, name of substance word, subject heading word, keyword heading word, protocol supplementary concept word, rare disease supplementary concept word, unique identifier, synonyms]

12. ((fecal or faecal or feces or faeces or stool or microbiota) adj2 (transplant\$ or infus\$ or transfus\$ or implant\$ or instil\$ or donat\$ or donor or reconstitut\$ or therap\$ or bacteriotherapy or encapsulated\$ or capsul\$)).ti,ab.

13. (fecal or faecal or feces or faeces or stool or microbiota).ti,ab.

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14. (transplant\$ or infus\$ or transfus\$ or implant\$ or instil\$ or donat\$ or donor or reconstitut\$ or therap\$ or bacteriotherapy or encapsulated\$ or capsul\$).ti,ab.

15. Transplantation/

16. Transplants/

17. 11 or 12

18. 14 or 15 or 16

19. 13 and 18

20. 17 or 19

21. or/1-10

22. 20 and 21

*Limits:*

1. After 1980.
2. Studies in English only.
3. Human studies only.
4. Exclude case reports.
5. Exclude case series with less than 10 patients.

**Review Question 6:***EMBASE*

1. (FMT or HPI).ti,ab.
2. ((fecal or faecal or feces or faeces or stool or microbiota) adj2 (transplant\* or infus\* or transfus\* or implant\* or instil\* or donat\* or donor\* or reconstitut\* or therap\* or bacteriotherapy)).ti,ab.
3. (fecal or faecal or feces or faeces or stool or microbiota).ti,ab.
4. transplant\*.ti,ab.
5. exp transplantation/
6. 1 or 2

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7. 3 and (4 or 5)

8. 6 or 7

9. (clostridium difficile or CDAD or RCDI or CDI).ti.

10. 8 not 9

11. limit 10 to (clinical trial or randomized controlled trial or controlled clinical trial)

*MEDLINE*

1. FMT.mp. or HPI.ti,ab. [mp=title, abstract, heading word, drug trade name, original title, device manufacturer, drug manufacturer, device trade name, keyword, floating subheading word]

2. ((fecal or faecal or feces or faeces or stool or microbiota) adj2 (transplant\$ or infus\$ or transfus\$ or implant\$ or instil\$ or donat\$ or donor or reconstitut\$ or therap\$ or bacteriotherapy)).ti,ab.

3. (fecal or faecal or feces or faeces or stool or microbiota).ti,ab.

4. Transplantation/

5. Transplants/

6. transplant\$.ti,ab.

7. Fecal Microbiota Transplantation/

8. 4 or 5 or 6

9. 3 and 8

10. 1 or 2 or 7 or 9

11. (clostridium difficile or cdiff or CDAD or RCDI or CDI or pseudomembranous).ti.

12. 10 not 11

13. limit 12 to (clinical trial or randomized controlled trial or controlled clinical trial)

*Limits:*

1. After 1980.

2. Studies in English only.

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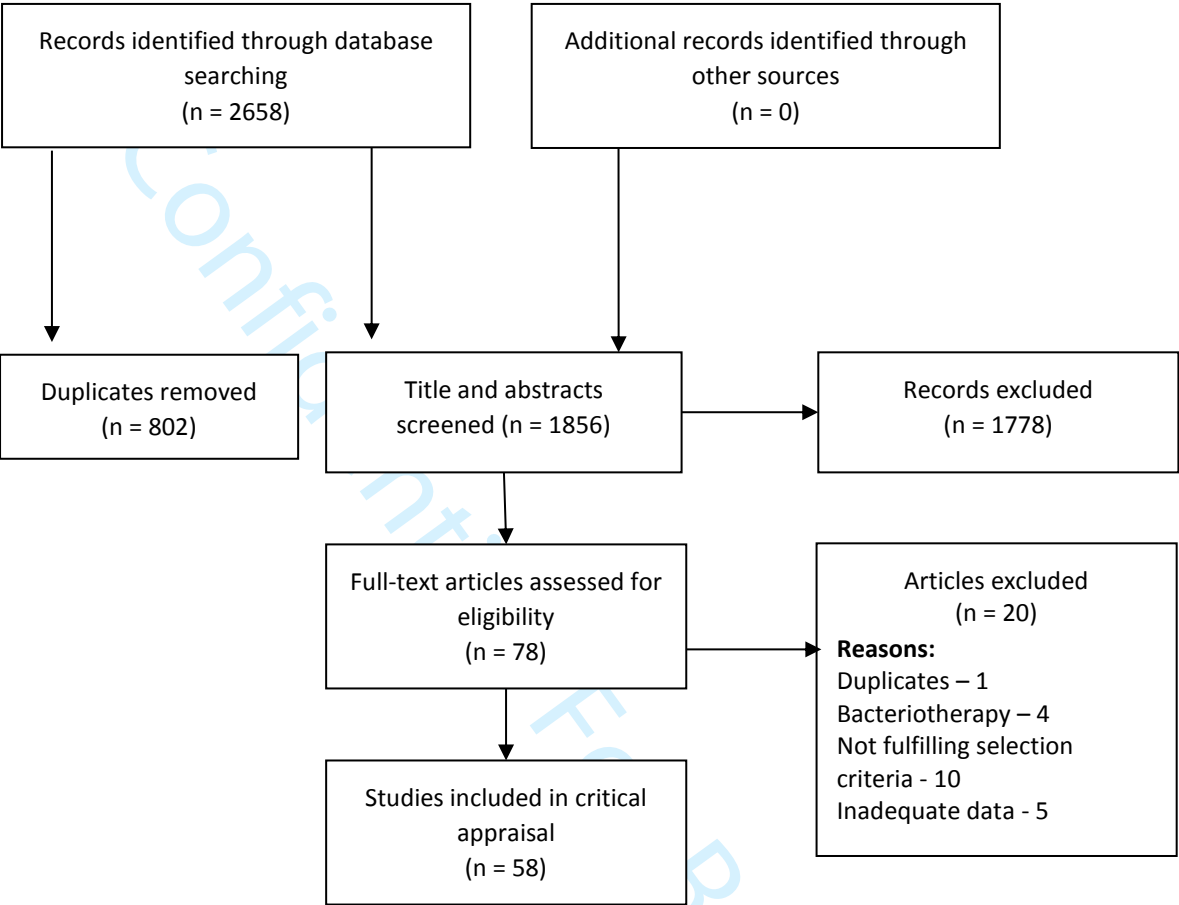
3. Human studies only.
4. Randomised trials only.

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Supplementary Material 1 for Gut

iii. Summary of the data extraction and literature review process (includes Q1-6):



Appendix 3: Consultation Stakeholders:

Individuals or organisation who were invited to and/ or attended the scoping day for these guidelines (as well as to provide feedback in stakeholder consultation) included:

- HSPA (Ireland) (Dr Eadaoin Griffin attended)
- Human Tissue Authority (Dr Robert Watson attended)
- NHS Wales
- NHS Scotland
- ECDC
- Royal College of Pathologists
- Royal College of General Practitioners
- Infection Prevention Society

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- Public Health England
- Royal College of Physicians
- Royal College of Nursing
- Royal College of Surgeons
- ESCMID
- MRSA Action
- HSCNI
- Institute of Microbiology and Infection, University of Birmingham (Prof Peter Hawkey and Dr Victoria McCune attended)
- Microbiology, Royal Devon and Exeter NHS Foundation Trust (Dr Ray Sheridan, Dr Alaric Colville, Dr Robert Porter and Dr Melissa Baxter attended)
- C diff support (Ms Graziella Kontkowski attended)
- OpenBiome (Dr Majdi Osman and Dr Carolyn Edelstein attended)
- Dr Sally Cudmore (University College Cork) attended
- Dr Ngozi Elumogo attended (Microbiology, Norfolk & Norwich University NHS Trust)
- Dr Vanya Gant (University College London Hospitals)
- Dr Simon Goldenberg attended (Guy's and St Thomas' NHS Foundation Trust)
- Dr Bram Goorguis attended (Academic Medical Centre, Amsterdam)
- Dr Geraldine Moloney attended (Microbiology, Trinity College Dublin)
- Dr Benjamin Mullish attended (Imperial College Healthcare NHS Trust)
- Dr Laura Prtak attended (Sheffield Teaching Hospitals NHS Trust)
- Mr Glenn Taylor attended (Taymount Clinic)
- Dr Mark Wilks attended (Microbiology, Barts and The London NHS Trust)

**Appendix 4. Continuing Professional Development material**

- 1) In which of the following settings would you **most strongly** avoid giving a patient FMT?
  - a) Immunocompromised patients
  - b) Decompensated liver disease
  - c) Heart failure
  - d) History of anaphylactic food allergy
  - e) A previous failed FMT

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Answer: d

2) Where is FMT best sourced, if available?

- a) Related healthy donor
- b) Health care professional
- c) Centralised stool bank
- d) Pooled from multiple donors
- e) Any of above

Answer: c

3) What is the maximum recommended length of time between stool donation and stool processing?

- a) 6 hours
- b) 7 hours
- c) 8 hours
- d) 9 hours
- e) 10 hours

Answer: a

4) For which non-CDI condition is FMT currently recommended?

- a) Irritable bowel syndrome
- b) Obesity and metabolic syndrome
- c) Parkinson's disease
- d) Ulcerative colitis
- e) None of the above

Answer: e

5) When considering setting up an FMT service in the UK, which organisation should be contacted to seek guidance in establishing the service?

- a) Medicines and Healthcare Products and Regulatory Agency
- b) Medicines and Healthcare Products Regulatory Authority
- c) Medical Drugs and Healthcare Products and Regulatory Agency

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- d) Medical Drugs and Healthcare Products Regulatory Authority  
e) None of the above

Answer: b

### 3. References:

1. Varier RU, Biltaji E, Smith KJ, et al. Cost-Effectiveness Analysis of Fecal Microbiota Transplantation for Recurrent *Clostridium difficile* Infection. *Infect Control Hosp Epidemiol*. 2015;36(4):438-444. doi:10.1017/ice.2014.80.
2. Konijeti GG, Sauk J, Shrimel MG, Gupta M, Ananthakrishnan AN. Cost-effectiveness of competing strategies for management of recurrent *Clostridium difficile* infection: a decision analysis. *Clin Infect Dis*. 2014;58(11):1507-1514. doi:10.1093/cid/ciu128.
3. Baro E, Galperine T, Denies F, et al. Cost-Effectiveness Analysis of Five Competing Strategies for the Management of Multiple Recurrent Community-Onset *Clostridium difficile* Infection in France. Green J, ed. *PLoS One*. 2017;12(1):e0170258. doi:10.1371/journal.pone.0170258.
4. Lapointe-Shaw L, Tran KL, Coyte PC, et al. Cost-Effectiveness Analysis of Six Strategies to Treat Recurrent *Clostridium difficile* Infection. Deshpande A, ed. *PLoS One*. 2016;11(2):e0149521. doi:10.1371/journal.pone.0149521.

Supplementary Material 2 for *Gut*

**The use of faecal microbiota transplant as treatment for recurrent or refractory *Clostridium difficile* infection and other potential indications: joint British Society of Gastroenterology (BSG) and Healthcare Infection Society (HIS) guidelines.**

**Supplementary Material 2: Additional Appendices**

**Appendix A. Scope**

**1. Guideline title**

The use of faecal microbiota transplant as treatment for recurrent or refractory *Clostridium difficile* infection and other potential indications: joint British Society of Gastroenterology (BSG) and Healthcare Infection Society (HIS) guidelines.

**1.1. Short title**

The use of faecal microbiota transplant

**2. The remit**

- i. To review the evidence (include randomised trial evidence) for the efficacy of faecal microbiota transplant (FMT) in the treatment of adults ( $\geq 18$  years), both in *Clostridium difficile* infection (CDI) and in other clinical conditions, and use this to make recommendations about optimal recipient selection and management, donor assessment, material preparation and administration, and other key elements of FMT delivery.
- ii. To provide specific guidance about best practice for an FMT service within the context of the regulatory framework for the intervention as it currently exists in the UK and beyond.

Whilst this is not a guideline specifically addressing the management of *Clostridium difficile* infection (CDI), the working group will include consideration of where FMT should be considered within the conventional treatment algorithm of patients with CDI (specifically, in which patients it should be considered, and at which point in their care).

The working group agreed that for the purposes of this guideline, faecal microbiota transplant would be defined as treatment that involves the administration of manipulated whole stool.

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There is a growing literature of the use of 'bacteriotherapy' originally deriving from healthy donor stool as a potential alternative to FMT (including commensal bacteria, spores, bacteriophages and/ or bacterial proteins or metabolites). However, the working group considered this to still be at the research stage, and would not be considered further.

**2.1. Population****2.1.1. Groups that will be covered**

Adults ( $\geq 18$  years) in whom:

- i. FMT has been used as treatment for CDI.
- ii. FMT has been used as treatment for a non-CDI indication.

Given the variability in the means used to diagnose CDI within different studies, the working group agreed to consider the suitability of the definition used on a study-by-study basis.

**2.1.2. Groups that will not be covered**

Children and young people ( $< 18$  years).

**2.2. Healthcare setting**

All settings in which National Health Service care is received, and/ or clinical trials are undertaken.

**2.3. Clinical management****2.3.1. Key clinical issues that will be covered**

- a) Appropriate selection of patients with CDI for FMT, and best practice in their management post-FMT.
- b) Optimal selection of donors of faecal material, and maintenance of a donor pool.
- c) Identification of the preferred means of preparation and administration of FMT to recipients.
- d) Evaluation of the safety and efficacy of FMT in treating non-CDI indications.
- e) Best practice in the development and delivery of an FMT service.

**2.3.2. Clinical issues that will not be covered**

- a) General management of CDI.

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- b) General management of non-CDI conditions in which FMT may have a role in therapy.

**2.4. Main outcomes**

Recommendations for practice

- a) Patient/ recipient selection, and peri-FMT management
- b) Donor selection
- c) Preparation and administration of FMT
- d) Efficacy and safety of FMT for non-CDI indications
- e) Provision of an FMT service

**2.5. Economic aspects**

Where FMT is being provided under a MHRA license according to Good Manufacturing Practice (GMP) standards, there are significant costs associated with initial setup and maintenance of the service. These include the cost of obtaining the relevant license, laboratory design and equipment to enable quality assurance, storage facilities for samples, etc. However, there is counterbalance to this, as the expectation of the working group is that the publication of this guideline may encourage provision of FMT as treatment for recurrent or refractory CDI. This has consistently been shown to be cost effective in comparison with anti-*C. difficile* antimicrobial therapy<sup>31–34</sup>, so overall costs associated with treating the condition may actually decrease. Furthermore, there may be changes to the practice of clinicians already offering the service. For example, encouraging the use of healthy unrelated donors (who can provide multiple stool donations after one screening) reduces the cost of screening when compared to the use of an FMT recipient’s relative as donor, who is likely to provide one donation only.

**2.6. Status**

**2.6.1. Scope**

This is the final scope.

**2.6.2. Timing**

The development of the guideline recommendation will begin in July 2017.



Supplementary Material 2 for *Gut***3. Related NICE guidance**

National Institute for Health and Care Excellence. *Faecal microbiota transplant for recurrent Clostridium difficile infection*. NICE Interventional Procedures Guidance IPG485. London: NICE; 2014. Available at: <https://www.nice.org.uk/guidance/ipg485> [last accessed 19th December 2017].

**4. Further information***Guideline development process*

Scottish Intercollegiate Guidelines Network. *SIGN 50: a guideline developer's handbook*. Revised edition. Edinburgh: Healthcare Improvement Scotland; 2014. Available at: <http://www.sign.ac.uk> [last accessed December 2017].

**Appendix B. Declarations of interest**

**B.1. *Introduction***

All members of the Working Group were required to make formal declarations of interest at the outset, and these were updated throughout the development process. No interests were declared that required any actions.

**B.2. *Tariq Iqbal***

First meeting 19/07/17: no declarations of interest; second meeting 04/10/17: no change.

Third meeting 19/10/17: consultant, advisor or speaker for: Pharmacosmos and Shield Therapeutics.

**B.3. *Simon Goldenberg (co-chair)***

First meeting 19/07/17

Advisory board and/ or consultancy and/ or speaker fees: Astellas, MSD, Pfizer.

Second meeting 04/10/17; third meeting 19/10/17: no change.

No action required.

**B.4. *Ailsa Hart***

First meeting 19/07/17

Advisory board and/ or consultancy and/ or speaker fees: AbbVie, Atlantic, Bristol-Myers Squibb, Celltrion, Falk, Ferring, Janssen, MSD, Napp Pharmaceuticals, Pfizer, Pharmacosmos, Shire and Takeda. Global steering committee for Genentech.

Second meeting 04/10/17; third meeting 19/10/17: no change.

No action required.

No declared conflict of interests for the other participants.

Supplementary Material 2 for *Gut***Appendix C. Clinical evidence tables****C.1. Reviewed case series of FMT for recurrent or refractory CDI**

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Supplementary Material 2 for Gut

Paper	Study and patient characteristics	Donor characteristics	FMT characteristics	Outcomes	Adverse events	CRD
Aas et al, Clinical Infectious Diseases, 2003	<p>Case series.</p> <p>Number of patients: 18.</p> <p>Female: male 13:5.</p> <p>Age (mean): 73+/-9 (range 53-88) years.</p> <p>Comorbidities: x1 patient with Crohn's colitis, x1 with leukaemia.</p> <p>CDI features: Recurrent (at least 2 x laboratory-confirmed CDI after initial antibiotic treatment).</p> <p>CDI diagnosis confirmation: Cytotoxin A and B positivity.</p> <p>Pre-FMT antibiotics: Metronidazole +/- vancomycin (not defined).</p>	<p>Donors were 15 family members, and 3 clinical volunteers.</p> <p>Working in healthcare: Yes - for 3 donors.</p> <p>Donor demographics: Not defined.</p> <p>Donor screening: Questionnaire not explicitly stated.</p> <p>Travel and antibiotic exclusion period: No antibiotics for 6 months prior; nil stated regarding travel.</p> <p>Screening blood tests: Hepatitis A, B and C, HIV-1/-2, syphilis.</p> <p>Screening stool tests: C.difficile, enteric pathogens, ova, cysts and parasites.</p>	<p>Amount of stool per transplant / administered to patients: 30g stool in 50-70ml normal saline; only 25ml of total administered to patient.</p> <p>Diluent used to prepare: Normal saline.</p> <p>Diluent used to store if frozen: N/A – fresh.</p> <p>Preparation methods: Homogenised in domestic blender, then coffee filter.</p> <p>Time from preparation to transplant (fresh): 6 hours.</p> <p>Time period for storage (frozen): N/A.</p> <p>Route administered: Upper GI: all nasogastric (18); lower GI: nil; capsules: nil.</p> <p>Number of infusions: Single infusion for all patients.</p> <p>Bowel purgative: Not described.</p> <p>PPI: 20mg omeprazole on day prior to FMT and day of FMT.</p> <p>Antimotility: Not described.</p> <p>Prokinetics: Not described.</p>	<p>Overall cure within stated follow up period: 83.3% (n=15/18).</p> <p>Cure with one infusion alone: 83.3% (n=15/18).</p> <p>Total follow-up period: 90 days.</p>	<p>Minor GI adverse events: Nil stated.</p> <p>Minor non-GI adverse events: Nil stated.</p> <p>Serious adverse events: Nil stated.</p> <p>Deaths: x2 - one related to ESRF, one related to COPD.</p>	<p>Selection/ eligibility reported: Yes.</p> <p>Consecutively recruited: Yes.</p> <p>Prospectively recruited: No.</p> <p>Loss to follow up explained: Yes.</p> <p>At least 90% followed up: No - 89%.</p>

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Time before CDI treatment was stopped  
before FMT: Continued until day of FMT.

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Agrawal <i>et al</i> , <i>Journal of Clinical Gastroenterology</i> , 2016	Case series.	Donors were identified by the patient or - if not available - provided by the physician.	Amount of stool per transplant / administered to patients: 60-100g of fresh stool.		Minor GI adverse events: Nil stated.	
	Number of patients: 146.	Working in healthcare: Not stated.	Diluent used to prepare: Normal saline, upper GI: 75-200ml; lower GI: 250-400ml; enema: 150-200ml.		Minor non-GI adverse events: Nil stated.	
	Female: male: 100: 46.	Donor demographics: No antibiotics for last three months. Excluded if significant GI disease, metabolic syndrome, chronic illness, immunocompromise, recent travel, and/ or high risk lifestyle in last three months.	Diluent used to store if frozen: N/A – fresh.		Serious adverse events: x2 microscopic colitis, x1 Sjögren’s, x1 scalp follicular lymphoma, x1 contact dermatitis and idiopathic Bence-Jones gammaglobulinaemia. In addition, x1 SCC, x1 ileus (died two weeks after ileus), x1 colonic perforation secondary to CMV colitis and subsequent death after 1 year. Patients developing cancers had underlying risk factors.	Selection/ eligibility reported: Yes.
	Age(mean): 78.6 (range 65-97) years.	Donor screening: Questionnaire - excluded if significant GI disease, metabolic syndrome, chronic illness, immunocompromise, recent travel, high risk lifestyle in last three months.	Preparation methods: Handstirred and blender, sifted through gauze.	Overall cure within stated follow-up period: 83% (n=121/146) .		Consecutively recruited: Yes.
	Comorbidities: Immunosuppression in 15 patients (x3 Crohn’s, x2 UC, x1 renal transplant)	Travel and antibiotic exclusion period: Exclusion if travel to an area of high incidence of infectious diarrhoea, and/ or antibiotics within past three months.	Time from preparation to transplant (fresh): Not stated.	Cure with one infusion alone: 83% (n=121/146) .		Prospectively recruited: No.
	CDI features: 89 with recurrent CDI.	Screening blood tests: Hepatitis A, B and C, HIV-1/-2, syphilis.	Time period for storage (frozen): N/A.	Total follow up period: mean follow up was 12.3 months (range 1-48 months).		Loss to follow up explained: No.
	CDI diagnosis confirmation: As per ACG guidelines.	Screening stool tests: <i>C difficile</i> , enteric pathogens, ova, cysts	Route administered: upper GI (16); lower GI (130); capsules: nil.		Deaths: x10 (x4 decompensated CCF, x3 malignancies, x1 dementia, x1	At least 90% followed up: No.
	Pre-FMT antibiotics: All had prior metronidazole, vancomycin and/ or fidaxomicin.		Number of infusions: 1 routinely; 2nd infusion given with vancomycin so data unable to be extracted.			
			Bowel purgative: PEG on day prior to FMT.			
			PPI: Not stated.			

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		and parasites, <i>Giardia</i> , <i>Cryptosporidium</i> , <i>Isospora</i> , <i>H.</i> <i>pylori</i> , Rotavirus.	Time before CDI treatment was stopped before FMT: Between 3 days prior to FMT and one day prior to FMT.		stroke, x1 pneumonia); deaths between 19 days to 7 months post-FMT.	
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Alrabaa et al, Transplant Infectious Diseases, 2017	Case series.		Amount of stool per transplant / administered to patients: 12.5g of stool in 28.5g of product.			
	Number of patients: 13.					
	Female: male: 8:5.		Diluent used to prepare: normal saline - diluted to approx 100-150ml to administer.		Minor GI adverse events: Several patients transient cramps and/ or diarrhoea.	
	Age (median): 69 (range 59-74) years.	Donors were unrelated.	Diluent used to store if frozen: Not clear.			
	Comorbidities: Yes - x4 OLT, x1 kidney/ liver transplant, x1 lung transplant, x1 HIV+ with CD4 count of 453. x1 immunocompromised patients with IBS, x1 immunocompetent patient with IBS; no IBD patients.	Working in healthcare: Nox Donor demographics: As per OpenBiome protocolx Donor screening: Questionnaire - as per OpenBiome protocolx Travel and antibiotic exclusion period: As per OpenBiome protocolx	Preparation methods: As per OpenBiome protocol. Time from preparation to transplant (fresh): N/A. Time period for storage (frozen): As per OpenBiome protocol - not described in paper.	Overall cure within stated follow up period: 84.6% (n=11/13) at eight weeks post-FMT. Cure with one infusion alone: 100% (n=13/13) at 5 days.	Minor non-GI adverse events: Nil noted. Serious adverse events: x1 patient had episode of CMV reactivation at the time of FMT - thought unrelated. X1 patient had episode of mild transplant rejection two months after FMT - thought unrelated.	Selection/ eligibility reported: Yes. Consecutively recruited: Not clearly described. Prospectively recruited: No.
	CDI features: Not clear if recurrent or refractory. Mean of 4 previous episodes of CDI prior to FMT.	Screening bloods: FBC, hepatitis A, B and C, LFTs, HIV, HTLV-1/-2, syphilis.	Route administered: Upper GI (nasoduodenal): 13; lower GI: 0; capsules: nil.	Total follow up period: Follow up up to 8 weeks described.		Loss to follow up explained: Yes.
	CDI diagnosis confirmation: PCR.	Screening stools: C.difficile toxin, MC&S, ova, cysts and parasites, H.pylori stool antigen.	Number of infusions: One routinely, but retreated if relapsed after primary outcome. However - one renal transplant patient received 2 doses of FMT on consecutive days (with successful outcome).			At least 90% followed up: Yes.
	Pre-FMT antibiotics: All patients had previously had oral vancomycin, x7 prev metronidazole (either with or without vancomycin). x5 received fidaxomicin		Bowel purgative: Bowel preparation used - GoLytely (PEG). PPI: 40mg pantoprazole night before and morning of procedure.		Deaths: None.	

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	with or after oral vancomycin.		Antimotility: Loperamide 4mg 1 hour post FMT.  Prokinetics: Not described.  Time before CDI treatment was stopped before FMT: See last box.			
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Brandt <i>et al</i> , <i>American Journal of Gastroenterology</i> , 2012	Case series.	Donors were 45 spouses/ partners; 21 relatives; 1 unknown person.	Amount of stool per transplant / administered to patients: 6 tablespoons of stool up to entire donation; 300-700ml of transplant administered.			
	Number of patients: 77.	Working in healthcare: No.	Diluent used to prepare: Normal saline.		Minor GI adverse events: Not stated.	
	Female: male: 56: 21.	Donor demographics: No antibiotics within past 3 months.	Diluent used to store if frozen: N/A – fresh.		Minor non-GI adverse events: Not stated.	Selection/ eligibility reported: Yes.
	Age (mean): 65+/-17 (range 22-87) years.	Donor screening: Questionnaire - not stated.	Preparation methods: Hand blender used to prep.	Overall cure within stated follow up period: N/A.	Serious adverse events: Nil.	Consecutively recruited: Not clear.
	Comorbidities: Not stated.	Travel and antibiotic exclusion period: Excluded if travel to area of high incidence of infectious diarrhoea, or if antibiotics within past three months.	Time from preparation to transplant (fresh): Within 8 hours.	Cure with one infusion alone: 90.9% (n=70/77).	Deaths: x7 deaths (cause unknown in one case, x1 metastatic colorectal cancer (present from pre-FMT), x1 metastatic ovarian cancer, x1 pneumonia (non-enteric organism), x1 MI, x1 stroke, x1 sepsis five months after FMT.	Prospectively recruited: No.
	CDI features: All recurrent/ refractory.	Screening blood tests: HIV-1, HIV-2, hepatitis A, B and C, Syphilis.	Time period for storage (frozen): N/A.	Total follow up period: not clear, but some patients followed-up to 3 years.		Loss to follow up explained: Reported but not explained.
	CDI diagnosis confirmation: Not clear.	Screening stool tests: <i>Clostridium difficile</i> toxin (if unavailable then EIA), MC&S, <i>Giardia</i> , <i>Cryptosporidium</i> , ova, cysts and parasites, <i>H.pylori</i> , Acid Fast stain for <i>Cyclospora</i> , <i>Isospora</i> .	Route administered: Upper GI: 0; lower GI: all 77 colonoscopic.			At least 90% followed up: No - only 77%.
	Pre-FMT antibiotics: 62 patients had had prior metronidazole, 76 vancomycin (25 tapered vancomycin), 17 rifaximin.		Number of infusions: 77 patients had one (patients that had second not included because given with concurrent vancomycin).			
			Bowel purgative: All patients given prep but no details.			
			PPI: Not described.			
			Antimotility: Not described.			
			Prokinetics: Not described.			

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Time before CDI treatment was stopped  
before FMT: 3 days.

Supplementary Material 2 for Gut

Brumbaugh <i>et al</i> , <i>Journal of Pediatrics</i> , 2017	Case series.		Amount of stool per transplant / administered to patients: 30ml OpenBiome aliquot/ capsule, although not defined re stool quantity.			
	Number of patients: 42.		Diluent used to prepare: As per OpenBiome protocol	Overall cure within stated follow up period: 71% (n=30/42).		
	Female: male: 23: 19.	Donor: OpenBiome-supplied FMT.	Diluent used to store if frozen: As per OpenBiome protocol	Cure with one infusion alone: 71% (n=30/42) - remission in 94% (n =16/17) otherwise healthy children, 54% (n =7/13) (54%) with IBD, 75% (n=9/12) medically complex.	Minor GI adverse events: 6/47 FMT administrations accompanied by vomiting within 24hrs; self-resolved.	Selection/ eligibility reported: Yes.
	Age (median): 9 (range 1 -18) years.	Working in healthcare: No.	Preparation methods: As per OpenBiome protocol	Success in 71% of children when via NGT, and 67% via gastrostomy (non-significant).		Consecutively recruited: Yes.
	Comorbidities: 31% had IBD (x4 Crohn's, x9 UC); 29% 'medically complex', including oncological, metabolic, cardiopulmonary or neurological diagnoses.	Donor demographics: Not stated.	Time from preparation to transplant (fresh): None given fresh		Minor non-GI adverse events: Nil reported.	Prospectively recruited: No.
	CDI features: All children had had at least one course of vancomycin. Previously recurrent - at least 2 episodes.	Donor screening: Questionnaire: As per OpenBiome protocol.	Time period for storage (frozen): N/A		Serious adverse events: Nil reported.	Loss to follow up explained: Yes.
		Travel and antibiotic exclusion period: As per OpenBiome protocol.	Route administered: Upper GI: 41, nasogastric administration (some children used pre-existing gastrostomy); lower GI: 0; capsules: 1 (1 x 30 capsules).	Total follow up period: 5 patients with initial failure opted for 2nd and 2 cured, so total success of 76% (n=32/42).	Deaths: Nil reported.	At least 90% followed up: Yes.
	CDI diagnosis: Diarrhoea, haematochezia and/ or crampy abdominal pain in combination with positive <i>C. difficile</i> PCR.	Screening bloods: As per OpenBiome protocol.	Number of infusions: 1 routinely			
	Pre-FMT antibiotics: Not stated.	Screening stools: As per OpenBiome protocol.	Bowel purgative: Not stated			
			PPI: Rantidine for 24hrs prior to FMT			
			Antimotility: N/A			
			Prokinetics: N/A			
			Time before CDI treatment was stopped			

Supplementary Material 2 for *Gut*

before FMT: 48 hours, after minimum of  
5 days of vancomycin.

Supplementary Material 2 for Gut

Chin et al, <i>Clinical Gastroenterology &amp; Hepatology</i> , 2016	<p>Case series.</p> <p>Number of patients: 35.</p> <p>Female: male: 16: 19.</p> <p>Age (mean): 43 (range 8 -93) years.</p> <p>Comorbidities: IBD in all, 8 on corticosteroids, 3 on Immunomodulators, 11 on biologics.</p> <p>CDI features: Recurrent - at least 2 episodes.</p> <p>CDI diagnosis confirmation: Not stated.</p> <p>Pre-FMT antibiotics: Not stated.</p>	<p>Donors were age 18 - 50, no medications, BMI 18.5 – 25.</p> <p>Working in healthcare: Not stated.</p> <p>Donor demographics: Not stated.</p> <p>Donor screening: Questionnaire - adapted from US blood bank.</p> <p>Travel and antibiotic exclusion period: Excluded if antibiotic within past six months.</p> <p>Screening blood tests: FBC, U&amp;E, LFTs, CRP, ANA, hepatitis A, B and C, HBV, HIV-1/-2, syphilis.</p> <p>Screening stool tests: Faecal occult blood, rotavirus, bacterial pathogens, ova, cysts and parasites, Acid fast stain for <i>Giardia</i> and <i>Cryptosporidium</i>, <i>C difficile</i>, <i>H. pylori</i>.</p>	<p>Amount of stool per transplant / administered to patients: 41g of stool on average.</p> <p>Diluent used to prepare: Normal saline.</p> <p>Diluent used to store if frozen: Frozen in 10% glycerol.</p> <p>Preparation methods: Ambient air.</p> <p>Time from preparation to transplant (fresh): N/A; given fresh.</p> <p>Time period for storage (frozen): Up to 156 days.</p> <p>Route administered: Upper GI: 5 via nasogastric tube; lower GI: 3 via colonoscopy; capsule: 27 patients.</p> <p>Number of infusions: Not stated.</p> <p>Bowel purgative: Not routinely - just for colonoscopy (4 litres of PEG).</p> <p>PPI: 7 on PPI not as premedications.</p> <p>Antimotility: Not described.</p> <p>Prokinetics: Not described.</p> <p>Time before CDI treatment was stopped before FMT: 2 days prior to FMT.</p>	<p>Overall cure within stated follow up period: N/A.</p> <p>Cure with one infusion alone: Not stated.</p> <p>Total follow up period: At least 2 months (range 2 to 6 months).</p>	<p>Minor GI adverse events: Not specified.</p> <p>Minor non-GI adverse events: Not specified.</p> <p>Serious adverse events: two required surgery (diverting colostomy and total proctectomy), two developed perianal disease with no prior history of it.</p> <p>Deaths: Ni.</p>	<p>Selection/ eligibility reported: No.</p> <p>Consecutively recruited: No.</p> <p>Prospectively recruited: No.</p> <p>Loss to follow up explained: No.</p> <p>At least 90% followed up: No.</p>
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Supplementary Material 2 for *Gut*

Cohen et al, <i>Israel Medical Association Journal</i> , 2016	<p>Case series.</p> <p>Number of patients: 22.</p> <p>Female: male: 9: 13.</p> <p>Age (median): Median 71.5 (range 16-92) years.</p> <p>Comorbidities: x1 IBD (colonoscopic group), x2 patients on chemotherapy, unclear why.</p> <p>CDI features: Recurrent or refractory.</p> <p>CDI diagnosis confirmation: Diarrhoea and toxin testing.</p> <p>Pre-FMT antibiotics: 19 patients given previous metronidazole, 9 vancomycin (with 13 both together).</p>	<p>Donors were 13 unrelated, 9 related.</p> <p>Working in healthcare: Yes - for unrelated.</p> <p>Donor demographics: No details - just says screening similar to blood donors.</p> <p>Donor screening: Questionnaire - no details.</p> <p>Travel and antibiotic exclusion period: Excluded if antibiotics within past six months.</p> <p>Screening bloods: No details.</p> <p>Screening stools: No details.</p>	<p>Amount of stool per transplant / administered to patients: 60g stool average (35-75g), 250ml total once mixed with saline (100 - 300ml range).</p> <p>Diluent used to prepare: Normal saline.</p> <p>Diluent used to store if frozen: Not stated.</p> <p>Preparation methods: Some fresh, some frozen.</p> <p>Time from preparation to transplant (fresh): Not stated.</p> <p>Time period for storage (frozen): No details.</p> <p>Route administered: Upper GI: nasoduodenal in 10; lower GI: colonoscopic in 12.</p> <p>Number of infusions: 1 FMT.</p> <p>Bowel purgative: 3l of PEG if colonoscopic administration.</p> <p>PPI: PPI if upper GI administration.</p> <p>Antimotility: Not described.</p> <p>Prokinetics: Metoclopramide just prior to upper GI administration.</p>	<p>Overall cure within stated follow up period: 72.7% (<math>n=16/22</math>) at 2 months.</p> <p>Cure with one infusion alone: 72.7% (<math>n=16/22</math>) (5/10 upper GI (out of 7 analysed), 91.7% (<math>n=11/12</math>) for lower GI (out of 11 analysed)).</p> <p>Total follow up period: Results reported at two months, but followed up to six months (7 months in the upper GI arm and 5 in the lower GI arm followed up to 6 months).</p>	<p>Minor GI adverse events: x5 transient constipation/ abdominal discomfort.</p> <p>Minor non-GI adverse events: Not stated.</p> <p>Serious adverse events: See deaths.</p> <p>Deaths: x7 (x1 due to CDI, x1 chronic resp disease, x1 related to dialysis, x2 pneumonia, x1 sepsis at ten days post-FMT (aspiration of stool; had been gastroscopic administration), x1 died at home ?cause).</p>	<p>Selection/ eligibility reported: Yes.</p> <p>Consecutively recruited: Yes.</p> <p>Prospectively recruited: No.</p> <p>Loss to follow up explained: Yes.</p> <p>At least 90% followed up: Yes.</p>
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Supplementary Material 2 for Gut

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			Time before CDI treatment was stopped before FMT: 12-24hrs.			
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Supplementary Material 2 for *Gut*

<p>Costello <i>et al</i>, <i>Alimentary Pharmacology and Therapeutics</i>, 2015</p>	<p>Case series.</p> <p>Number of patients: 20.</p> <p>Female: male: not stated.</p> <p>Age(median): 69 years.</p> <p>Comorbidities: Not stated.</p> <p>CDI features: All recurrent.</p> <p>CDI diagnosis confirmation: Not stated.</p> <p>Pre-FMT antibiotics: Conventional therapy with metronidazole, vancomycin and/or fidaxomicin had failed in all.</p>	<p>Donors were 4 healthy volunteers.</p> <p>Working in healthcare: No.</p> <p>Donor demographics: No details.</p> <p>Donor screening: Questionnaire - adapted from US blood bank.</p> <p>Travel and antibiotic exclusion period: Excluded if travel to diarrhoea-endemic areas within 6 months and/ or used antibiotics for 3 months.</p> <p>Screening blood tests: HIV -1 and -2, hepatitis A, B and C, and syphilis.</p> <p>Screening stool tests: <i>C difficile</i> toxin B PCR, routine MC&amp;S, faecal <i>Giardia</i> antigen, faecal <i>Cryptosporidium</i>, Acid-fast stain for <i>Cyclospora</i>, <i>Isospora</i>, ova, cysts and parasites, <i>H.pylori</i> fecal antigen.</p>	<p>Amount of stool per transplant / administered to patients: Not stated.</p> <p>Diluent used to prepare: Normal saline.</p> <p>Diluent used to store if frozen: 10% glycerol.</p> <p>Preparation methods: Anaerobically prepared.</p> <p>Time from preparation to transplant (fresh): all frozen.</p> <p>Time period for storage (frozen): 16 patients had stool stored for &lt; 2 months. 4 patients had stool stored &gt; 2 months.</p> <p>Route administered: Upper GI: 1; lower GI: 19; capsule: nil.</p> <p>Number of infusions: 17 patients had 1, 3 patients had 2.</p> <p>Bowel purgative: Not reported.</p> <p>PPI: Not reported.</p> <p>Antimotility: Not reported.</p> <p>Prokinetics: Not reported.</p> <p>Time before CDI treatment was stopped before FMT: Not reported.</p>	<p>Overall cure within stated follow up period: 85% (n=17/20).</p> <p>Cure with one infusion alone: 85% (n=17/20).</p> <p>Total follow up period: Minimum 3 months (but up to 14 months).</p>	<p>Minor GI adverse events: None.</p> <p>Minor non-GI adverse events: None.</p> <p>Serious adverse events: None.</p> <p>Deaths: None.</p>	<p>Selection/ eligibility reported: Yes.</p> <p>Consecutively recruited: Yes.</p> <p>Prospectively recruited: No.</p> <p>Loss to follow up explained: Yes.</p> <p>At least 90% followed up: Yes.</p>
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Supplementary Material 2 for Gut

Emanuelsson <i>et al</i> , <i>Scandinavian Journal of Infectious Diseases</i> , 2014	Case series.	Donors were spouses or close relative.	Amount of stool per transplant / administered to patients: 50g in 500mls.			
	Number of patients: 23.	Donor working in healthcare: No.	Diluent used to prepare: Normal saline.			
	Female: male: 14: 9.	Donor demographics: Not stated.	Diluent used to store if frozen: N/A - fresh.			
	Age (median): 66 years (range 25-99) years (including 8 additional patients treated with 'bacteriotherapy').	Donor screening: Questionnaire – asked regarding current and previous GI diagnoses/ symptoms.	Preparation methods: Anaerobically prepared.			
	Comorbidities: 3 with diabetes mellitus, 1 with microscopic colitis.	Travel and antibiotic exclusion period: Definitely an antibiotic use restriction but not clearly stated.	Time from preparation to transplant (fresh): Not stated.	Overall cure within stated follow up period: 65% (n=15/23).	Minor GI adverse events: None.	Selection/eligibility reported: Yes.
	CDI features: All recurrent.	Screening blood tests: HIV-1 and -2, hepatitis C virus, and hepatitis B surface antigen.	Time period for storage (frozen): N/A.	Cure with one infusion alone: 65% (n=15/23).	Minor non-GI adverse events: None.	Consecutively recruited: Yes.
	CDI diagnosis confirmation: Culture and/or toxin EIA.	Screening stool tests: <i>Salmonella</i> , <i>Shigella</i> , <i>Campylobacter</i> , enterohemolytic <i>Escherichia coli</i> , and <i>Clostridium difficile</i> .	Route administered: Upper GI: nil; ower GI: 23 (enema/ rectal catheter); capsules: nil.	Total follow up period: Median follow up of 18 months (range 0-201 months).	Serious adverse events: None.	Prospectively recruited: No.
	Pre-FMT antibiotics: Metronidazole and/or vancomycin used in all patients beforehand.		Number of infusions: 22 patients eceived 1 FMT, 1 patient received 2 FMTs.	Deaths: None.		Loss to follow up explained: Yes.
			Bowel purgative: Not stated.			At least 90% followed up: Yes.
			PPI: Not stated.			
			Antimotility: Not stated.			
			Prokinetics: Not stated.			
			Time before CDI treatment was stopped before FMT: Not stated.			

Supplementary Material 2 for *Gut*

Fischer <i>et al</i> , <i>Inflammatory Bowel Diseases</i> , 2016	<p>Case series</p> <p>Number of patients: 67 Female: male: 39:28</p> <p>Age (mean/ standard deviation): Mean 45.42 (+/-17.33) years.</p> <p>Comorbidities: x5 PSC, x4 liver transplant, x3 end stage liver disease, concurrent IBD in all (x35 Crohn's, x31 UC, x1 indeterminate colitis).</p> <p>CDI features: recurrent or refractory.</p> <p>CDI diagnosis confirmation: Return of diarrhoea and positive CDI testing within 12 weeks of FMT.</p> <p>Pre-FMT antibiotics: metronidazole in 47 patients, vancomycin in 63, vancomycin taper in 38 patients, fidaxomicin in 7, rifaximin in 7.</p>	<p>Donors were patient-directed donor or unrelated healthy volunteers.</p> <p>Donors working in healthcare: not stated.</p> <p>Donor demographics: As per Bakken <i>et al</i>, <i>Clin Gastroenterol Hepatol</i>, 2011.</p> <p>Donor screening: Questionnaire - as per Bakken <i>et al</i>, <i>Clin Gastroenterol Hepatol</i>, 2011.</p> <p>Travel and antibiotic exclusion period: Excluded as donor if travel within last 6 months where diarrheal illnesses are endemic or risk of travelers diarrhea is high, and/ or use of antibiotics within 3 months.</p> <p>Screening blood tests: HIV -1&amp;-2, hepatitis A, B and C, syphilis.</p> <p>Screening stool tests: As per Bakken <i>et al</i>, <i>Clin Gastroenterol Hepatol</i>, 2011.</p>	<p>Amount of stool per transplant / administered to patients: lower GI: -25-50ml; upper GI: 250-500ml.</p> <p>Diluent used to prepare: Preservative-free normal saline or 4% milk.</p> <p>Diluent used to store if frozen: N/A – fresh.</p> <p>Preparation methods: Household blender, homogenized and removal of particle matter with gauze/ urine strainers in a Biohazard Level 2 facility.</p> <p>Time from preparation to transplant (fresh): Certainly within 24 hours, and preferably within 6 hours.</p> <p>Time period for storage (frozen): N/A.</p> <p>Route administered: Upper GI: nil; lower GI: 67 (colonoscopy or sigmoidoscopy); capsule: nil.</p> <p>Number of infusions: 53 patients received one infusion, 14 received 2 infusions.</p> <p>Bowel purgative: Standard bowel preparation, but not specified.</p> <p>PPI: If upper GI administration, PPI on the evening before and morning of the procedure.</p>	<p>Overall cure within stated follow up period: 90% (n=60/67) within 3 months.</p> <p>Cure with one infusion alone: 79% (n=53/67).</p> <p>Total follow up period: average length 10.4 (range 3-36) months.</p>	<p>Minor GI adverse events: x1 IBD flare, managed as outpatient.</p> <p>Minor non-GI adverse events: x4 pneumonia.</p> <p>Serious adverse events: x1 colectomy for refractory IBD, x7 hospitalised, x2 CDI recurrence, x1 IBD exacerbation, x1 small bowel obstruction, x1 CMV colitis.</p> <p>Deaths: none.</p>	<p>Selection/ eligibility reported: Yes.</p> <p>Consecutively recruited: No.</p> <p>Prospectively recruited: No.</p> <p>Loss to follow up explained: N/A.</p> <p>At least 90% followed up: N/A.</p>
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Supplementary Material 2 for Gut

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			<p>Antimotility: Loperamide optional for lower GI administration.</p> <p>Prokinetics: Not stated.</p> <p>Time before CDI treatment was stopped before FMT: 24-48 hrs.</p>			
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Supplementary Material 2 for *Gut*

Fischer <i>et al</i> , <i>American Journal of Gastroenterology</i> , 2016	<p>Case series.</p> <p>Number of patients: 328.</p> <p>Female: male: 241: 87.</p> <p>Age (mean/ standard deviation): 61.4 (+/-19.3) years.</p> <p>Comorbidities: 77 immunocompromised (x3 CVID, x3 selective IgA deficiency, x71 immunosuppressants (20 for solid organ transplant, 29 for IBD, 6 for rheumatoid arthritis, 2 for SLE, 1 for pemphigoid, 1 for chronic obstructive airway disease, 1 for psoriasis)), x11 chemotherapy for malignancy, x63 IBD (25 UC, 33 Crohn's), x118 diverticulosis.</p> <p>CDI features: Recurrent disease in 87.2% and severe or severe-complicated in 12.8%.</p> <p>CDI diagnosis confirmation: Postive stool <i>C difficile</i> toxin or</p>	<p>Donors were 130 (40%) patient-directed donors, and 198 universal (60%).</p> <p>Donor working in healthcare: Not stated.</p> <p>Donor demographics: Not stated.</p> <p>Donor screening: Questionnaire – depended upon individual centre.</p> <p>Travel and antibiotic exclusion period: Depended upon individual centre.</p> <p>Screening blood tests: Depended upon individual centre.</p> <p>Screening stool test: Depended upon individual centre.</p>	<p>Amount of stool per transplant / administered to patients: Not specified.</p> <p>Diluent used to prepare: Not specified.</p> <p>Diluent used to store if frozen: Both fresh and frozen, but specific details not given.</p> <p>Preparation methods: Dependent upon individual centre.</p> <p>Time from preparation to transplant (fresh): Dependent upon individual centre.</p> <p>Time period for storage (frozen): Dependent upon individual centre.</p> <p>Route administered: Not specified ('predominantly colonoscopy').</p> <p>Number of infusions: Dependent upon individual centre.</p> <p>Bowel purgative: Not specified.</p> <p>PPI: Not specified.</p> <p>Antimotility: Not specified.</p> <p>Prokinetics: Not specified.</p> <p>Time before CDI treatment was stopped before FMT: Dependent upon each centre.</p>	<p>Overall cure within stated follow up period: 1 month 81.4% (<i>n</i>=267/328).</p> <p>Cure with one infusion alone: Not specified.</p> <p>Total follow up period: Not specified.</p>	<p>Minor GI adverse events: Not specified.</p> <p>Minor non-GI adverse events: Not specified.</p> <p>Serious adverse events: Not specified.</p> <p>Deaths: Not specified.</p>	<p>Selection/ eligibility reported: Yes.</p> <p>Consecutively recruited: No.</p> <p>Prospectively recruited: No.</p> <p>Loss to follow up explained: N/A.</p> <p>At least 90% followed up: N/A.</p>
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Supplementary Material 2 for *Gut*

	PCR.					
	Pre-FMT antibiotics: vancomycin.					

Supplementary Material 2 for *Gut*

Fischer <i>et al</i> , <i>Gut Microbes</i> , 2017	<p>Case series.</p> <p>Number of patients: 57.</p> <p>Female: male: 34: 23.</p> <p>Age (median): Median 72 (range 25-99) years.</p> <p>Comorbidities: x7 toxic megacolon, x12 acute kidney injury (x3 needing dialysis), x10 with hypovolaemic/ septic shock, x7 mental status changes, x4 on mechanical ventilation. x10 patients had inflammatory bowel disease (x5 with Crohn's and x5 with ulcerative colitis) and x10 patients were on immunosuppressive medications.</p> <p>CDI features: Severe, recurrent and severe-complicated.</p> <p>CDI diagnosis confirmation: Positive stool <i>C.difficile</i> PCR.</p> <p>Pre-FMT antibiotics: Included vancomycin,</p>	<p>Donors were screened patient-selected donors for first 29 patients, whilst next 28 from OpenBiome stool bank.</p> <p>Donors working in healthcare: Not specified.</p> <p>Donor demographics: Not specified.</p> <p>Donor screening: Questionnaire – for patient-selected donors, this was as for Bakken <i>et al</i>, <i>Clin Gastroenterol Hepatol</i>, 2011; for OpenBiome, as per OpenBiome protocol.</p> <p>Travel and antibiotic exclusion period: For patient-selected donors, this was as for Bakken <i>et al</i>, <i>Clin Gastroenterol Hepatol</i>, 2011; for OpenBiome, as per OpenBiome protocol.</p> <p>Screening blood tests: For patient-selected donors, this was as for Bakken <i>et al</i>, <i>Clin Gastroenterol Hepatol</i>, 2011; for OpenBiome, as per OpenBiome protocol.</p> <p>Screening stool tests: For patient-selected donors, this was as for Bakken <i>et al</i>, <i>Clin Gastroenterol Hepatol</i>, 2011;</p>	<p>Amount of stool per transplant / administered to patients: As per Fischer <i>et al</i>, <i>Alim Pharm Ther</i>, 2015 or OpenBiome.</p> <p>Diluent used to prepare: As per Fischer <i>et al</i>, <i>Alim Pharm Ther</i>, 2015 or OpenBiome.</p> <p>Diluent used to store if frozen: As per Fischer <i>et al</i>, <i>Alim Pharm Ther</i>, 2015 or OpenBiome .</p> <p>Preparation methods: As per Fischer <i>et al</i>, <i>Alim Pharm Ther</i>, 2015 or OpenBiome.</p> <p>Time from preparation to transplant (fresh): 6 hours.</p> <p>Time period for storage (frozen): As per OpenBiome protocols.</p> <p>Route administered Upper GI: nil; lower GI: 57 via colonoscopy or sigmoidoscopy.</p> <p>Number of infusions: 32 patients: x1, 20 patients x2, 5 patients x3, 1 patient x4, 1 patient x5. Pre-planned protocol for serial FMTs +/- vancomycin, as described in Fischer <i>et al</i>, <i>Alim Pharm Ther</i>, 2015.</p> <p>Bowel purgative: Not stated.</p>	<p>Overall cure within stated follow up period: 91% (<math>n=52/57</math>), i.e. 100% severe CDI (<math>n=19/19</math>), and 87% (<math>n=33/38</math>).</p> <p>Cure with one infusion alone: 52.6% (<math>n= 30/57</math>).</p> <p>Total follow up period: Up to 6 months.</p>	<p>Minor GI adverse events: Not stated.</p> <p>Minor non-GI adverse events: Not stated.</p> <p>Serious adverse events: Not stated.</p> <p>Deaths: x7 unrelated deaths, x4 CDI-related deaths.</p>	<p>Selection/ eligibility reported: Yes.</p> <p>Consecutively recruited: Yes.</p> <p>Prospectively recruited: Yes.</p> <p>Loss to follow up explained: Yes.</p> <p>At least 90% followed up: Yes.</p>
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Supplementary Material 2 for *Gut*

	fidaxomicin, rectal vancomycin, intravenous metronidazole.	for OpenBiome, as per OpenBiome protocol.	PPI: Not stated.  Antimotility: Not stated.  Prokinetics: Not stated.  Time before CDI treatment was stopped before FMT: Not stated.			
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Supplementary Material 2 for *Gut*

Fischer <i>et al</i> , <i>Alimentary Pharmacology and Therapeutics</i> , 2015	<p>Case series.</p> <p>Number of patients: 29.</p> <p>Female: male: 17: 12.</p> <p>Age (mean/ standard deviation): Overall, mean 65.2 (+/-17.9) years (range 25-92 years); mean 60.8 (range 26-87) years in severe; 67.6 (range 60-78) years in severe-complicated.</p> <p>Comorbidities: x3 Crohn's, x2 UC, x1 hypogammaglobulinaemia, x1 ESKD, x1 ESLD, x1 renal transplant, x1 liver transplant, x4 on immunosuppressive meds. 12/19 of pts treated in ITU at the time with following complications: x5 patients with toxic megacolon (caecal diam &gt;12cm or rectosigmoid &gt; 6.5cm diameter); x7 AKI and hypovolaemic/ septic shock, x4 of which required vasopressors, x3 with change in mental status, x2 patients ventilated. x22 with</p>	<p>Donors were either patient selected-donor, or universal donors. If patient-directed, same donor used for subsequent FMTs if required. 44 FMTs in all - patient-selected for 16 FMTs, universal donor for 28 FMTs.</p> <p>Donors working in healthcare: Not described.</p> <p>Donor demographics: Not clear.</p> <p>Donor screening: Questionnaire: As per Bakken <i>et al</i>, <i>Clin Gastroenterol Hepatol</i>, 2011.</p> <p>Travel and antibiotic exclusion period: As per Bakken <i>et al</i>, <i>Clin Gastroenterol Hepatol</i>, 2011.</p> <p>Screening blood tests: As per Bakken <i>et al</i>, <i>Clin Gastroenterol Hepatol</i>, 2011.</p> <p>Screening stool tests: As per Bakken <i>et al</i>, <i>Clin Gastroenterol Hepatol</i>, 2011.</p>	<p>Amount of stool per transplant / administered to patients: 50-200g of stool.</p> <p>Diluent used to prepare: 300ml of saline.</p> <p>Diluent used to store if frozen: N/A – all fresh.</p> <p>Preparation methods: No additional details.</p> <p>Time from preparation to transplant (fresh): Six hours.</p> <p>Time period for storage (frozen): N/A.</p> <p>Route administered: Upper GI: nil; lower GI: flexible sigmoidoscopy or colonoscopy either proximal or distal to the splenic flexure at the discretion of the endoscopist. In practice – proximal to the splenic flexure in 18 FMTs, distal in 26.</p> <p>Number of infusions: As many as per protocol until end point. 16 x 1 FMT (7 severe, 9 complicated), 11 x 2nd FMT (3 severe, 8 compl), 2 x 3rd FMT (0 severe, 2 complicated).</p> <p>N.B. Oral vancomycin (125 mg every 6 hours) was resumed 24–48 hours after FMT for a minimum of 5 days if there were pseudomembranes present at colonoscopy. For patients who did not</p>	<p>Overall cure within stated follow up period: By 3 months, 62% (n=18/29) in remission.</p> <p>Cure with one infusion alone: 70% (n=7/10) in severe arm; 47% (n=9/19) in severe-complicated arm.</p> <p>Total follow up period: Up to 3 months.</p>	<p>Minor GI adverse events: Not stated.</p> <p>Minor non-GI adverse events: Not stated.</p> <p>Serious adverse events: Nil.</p> <p>Deaths: x2 deaths by 1 month; x1 death from sepsis within 24 hours of FMT); death following colectomy after 3x failed FMT in patient who was six weeks post-OLT. By 3 months – x2 further deaths from CDI recurrence, x1 death from cirrhosis, x1 death from heart failure, x1 death from respiratory failure, x1 death from aspiration.</p>	<p>Selection/ eligibility reported: Yes.</p> <p>Consecutively recruited: Yes.</p> <p>Prospectively recruited: Yes.</p> <p>Loss to follow up explained: Yes.</p> <p>At least 90% followed up: Yes.</p>
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Supplementary Material 2 for Gut

	<p>pseudomembranes at first FMT.</p> <p>CDI features: 9 patients with first episode of CDI; all others with previous episodes.</p> <p>CDI diagnosis confirmation: Diarrhoea (at least 3 loose stools/ day) and positive toxin.</p> <p>Pre-FMT antibiotics: Not stated.</p>		<p>improve by days 6–7, the vancomycin was stopped, and bowel prep was administered if no ileus was present. The next day (day 7–8), a repeat FMT, from the same donor as the first FMT if patient-directed, was performed by sigmoidoscopy or colonoscopy. If pseudomembranes were present, oral vancomycin was resumed for an additional 5 days. If no pseudomembranes were detected, antibiotics were not resumed following the repeat FMT.</p> <p>Bowel purgative: Split dose 4l Golytely if no ileus/ obstruction.</p> <p>PPI: Not described.</p> <p>Antimotility: Not described.</p> <p>Prokinetics: Not described.</p> <p>Time before CDI treatment was stopped before FMT: 12-24hr prior to FMT.</p>			
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Supplementary Material 2 for *Gut*

<p>Garborg <i>et al</i>, <i>Scandinavian Journal of Infectious Diseases</i>, 2010</p>	<p>Case series.</p> <p>Number of patients: 40.</p> <p>Female: male: 21: 19.</p> <p>Age (mean): Mean age 75 (range 53-94) years.</p> <p>Comorbidities: x1 Wegener's, x1 AML. Repeated courses of antibiotics, not formally described.</p> <p>CDI features: Not described.</p> <p>CDI diagnosis confirmation: Diarrhoea and + <i>C difficile</i> toxin (testing for A and B).</p> <p>Pre-FMT antibiotics: All patients had had at least two courses of oral metronidazole (500mg three times daily) or vancomycin (125mg po four times daily).</p>	<p>Donors were close relatives/ household members.</p> <p>Donors working in healthcare: No.</p> <p>Donor demographics: Not stated.</p> <p>Donor screening: Questionnaire - "Symptoms of GI disease or history of chronic infectious disease".</p> <p>Travel and antibiotic exclusion period: Not stated.</p> <p>Screening bloods: Hepatitis A, B and C, HIV.</p> <p>Screening stools: MC&amp;S, <i>Yersinia</i>. No routine parasite screening ("low prevalence in Norway").</p>	<p>Amount of stool per transplant / administered to patients: 50-100g.</p> <p>Diluent used to prepare: 250ml sterile normal saline.</p> <p>Diluent used to store if frozen: All fresh.</p> <p>Preparation methods: Stool placed on gauze pad and strained; flushed with saline; drawn up into syringes ready for administration.</p> <p>Time from preparation to transplant (fresh): Same day.</p> <p>Time period for storage (frozen): N/A.</p> <p>Route administered: Upper GI: OGD with delivery in distal duodenum; 38; lower GI: Colonoscopy; 2.</p> <p>Number of infusions: One at baseline; follow up if 'did not respond' although not specifically defined.</p> <p>Bowel purgative: Not mentioned, even for colonoscopy.</p> <p>PPI: Not stated.</p> <p>Antimotility: Not stated.</p> <p>Prokinetics: Not stated.</p>	<p>Overall cure within stated follow up period: 835 (n=33/40).</p> <p>Cure with one infusion alone: 73% (n=29/ 40) (28 in duodenum, 1 in colon).</p> <p>Total follow up period: Up to 80 days.</p>	<p>Minor GI adverse events: Not stated.</p> <p>Minor non-GI adverse events: Not stated.</p> <p>Serious adverse events: Not stated.</p> <p>Deaths: x5 deaths within 3 weeks - 2 months post-FMT but none attributable to FMT. x2 deaths attributed to 'frailty', x1 advanced Wegener's, x1 AML/ antibiotics, one patients with advanced cardiovascular disease who had fulminant colitis, underwent colectomy, but died.</p>	<p>Selection/ eligibility reported: Yes.</p> <p>Consecutively recruited: Yes.</p> <p>Prospectively recruited: No.</p> <p>Loss to follow up explained: Yes.</p> <p>At least 90% followed up: Yes.</p>
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Supplementary Material 2 for *Gut*

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			Time before CDI treatment was stopped before FMT: Evening prior to FMT.			
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Supplementary Material 2 for *Gut*

<p>Girotra <i>et al</i>, <i>Digestive Diseases and Sciences</i>, 2016</p>	<p>Case series.</p> <p>Number of patients: 29.</p> <p>Female: male: 6: 23.</p> <p>Age (mean/ standard deviation): 80.1 (+/-6.49) years (13 patients 70-79, 14 patients 80-89, 2 patients &gt; 90 years).</p> <p>Comorbidities: x8 patients with diabetes mellitus.</p> <p>CDI features: No specific details - purely symptoms &gt; 6 months, failed at least 3 antibiotic regimens.</p> <p>CDI diagnosis confirmation: At least three unformed stools in 24 hour and positive stool <i>C difficile</i> test by toxin (by ELISA) or toxin gene B (by PCR). All patients here defined RCDI by symptoms &gt;6 months and at least x3 failed antibiotics.</p> <p>Pre-FMT antibiotics: Not indicated.</p>	<p>Donors were patient-selected family or friends.</p> <p>Donors working in healthcare: No.</p> <p>Donor demographics: Not stated.</p> <p>Donor screening: Questionnaire – peptic ulcer disease/GORD, IBS, IBD, polyps, malignancy, antibiotic use/ hospitalisation within past 3 months.</p> <p>Travel and antibiotic exclusion period: Excluded as donor if antibiotic use within the past three months.</p> <p>Screening bloods: HIV, HTLV-I/-II, syphilis enzyme immunoassay, hepatitis A immunoglobulin M, hepatitis B surface antigen, hepatitis C antibody, and <i>Helicobacter pylori</i> antibody.</p> <p>Screening stools: MC&amp;S/ ova, cysts and parasites x3, <i>Cryptosporidium</i>, <i>Microspora</i>, <i>C difficile</i> toxin.</p>	<p>Amount of stool per transplant / administered to patients: 450cc - 270cc via colonoscopy AND 180cc into jejunum via enteroscopy.</p> <p>Diluent used to prepare: Saline - whole stool sample (&gt;30g) mixed with 50-70ml of sterile saline, made up to 5 x 90cc aliquots.</p> <p>Diluent used to store if frozen: Fresh.</p> <p>Preparation methods: Stool mixed with saline, homogenised in blender for &lt;4 minutes, filtered x2 with coffee filter paper.</p> <p>Time from preparation to transplant (fresh): Within 6 hours.</p> <p>Time period for storage (frozen): N/A.</p> <p>Route administered: Enteroscopy into jejunum AND colonoscopy in all 29 patients.</p> <p>Number of infusions: 1 FMT per patient (combined upper and lower GI administration).</p> <p>Bowel purgative: Not described.</p> <p>PPI: 20mg omeprazole evening before/ morning of procedure.</p> <p>Antimotility: Not described.</p>	<p>Overall cure within stated follow up period: 100% (n=29/29).</p> <p>Cure with one infusion alone: 100% (n=29/29).</p> <p>Total follow-up period: Reported 25.37 +/- 12.8 months follow-up (range 8-50 months).</p> <p>In addition - researchers report 60% weight gain, 40% stable weight, 75% improved 'failure to thrive' (defined as decrease of weight &gt;10% from baseline, with no improvement despite medical treatment of CDI and nutritional treatment).</p>	<p>Minor GI adverse events: Bloating 10% (n=3/29).</p> <p>Minor non-GI adverse events: Fever 7% (n=2/29) (transient for one day).</p> <p>Serious adverse events: None.</p> <p>Deaths: None.</p>	<p>Selection/ eligibility reported: Yes.</p> <p>Consecutively recruited: Yes.</p> <p>Prospectively recruited: No.</p> <p>Loss to follow up explained: N/A.</p> <p>At least 90% followed up: Yes.</p>
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Supplementary Material 2 for *Gut*

			<p>Prokinetics: Not described.</p> <p>Time before CDI treatment was stopped before FMT: &gt;12 hours.</p>			
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Supplementary Material 2 for *Gut*

Hagel <i>et al</i> , <i>Deutsches Arzteblatt International</i> , 2016	<p>Case series.</p> <p>Number of patients: 133.</p> <p>Female: male: 86: 47.</p> <p>Age (median): Median 75 (IQR 59.5 - 81.5) years.</p> <p>Comorbidities: x3 chemotherapy, x19 immunosuppressants, x5 solid organ transplant, x1 allogeneic stem cell transplant, x43 GI comorbidities (no details).</p> <p>CDI features: Median of 3 recurrences (IQR 1-4); no specific details re recurrent vs refractory confirmation.</p> <p>Pre-FMT antibiotics: x4 metronidazole only, x13 vancomycin only, x2 fidaxomicin only, x61 metronidazole/ vancomycin, x8 vancomycin/ fidaxomicin, x34 metronidazole/ vancomycin/</p>	<p>Donors working in healthcare: not stated</p> <p>Donor demographics: Not stated.</p> <p>Donor screening: Questionnaire - not stated.</p> <p>Travel and antibiotic exclusion period: Not stated.</p> <p>Screening blood tests.: Rapid plasma reagin and fluorescent <i>Treponemal</i> antibody-absorbed.</p> <p>Screening stool tests: Not stated.</p>	<p>Amount of stool per transplant / administered to patients: Not stated.</p> <p>Diluent used to prepare: Not stated.</p> <p>Diluent used to store if frozen: Yes, in some cases - no details given.</p> <p>Preparation methods: Not stated.</p> <p>Time from preparation to transplant (fresh): Not stated.</p> <p>Time period for storage (frozen): Not stated.</p> <p>Route administered: Upper GI: 4 OGD, 40 enteroscopy, 19 nasoenteric tube; lower GI: 55 'endoscopic' (no further details); capsule: 13. x2 combination of jejunal and colonoscopic FMT.</p> <p>Number of infusions: 1 FMT.</p> <p>Bowel purgative: Yes - 117 (no details given).</p> <p>PPI: Yes - 31 (no details given).</p> <p>Antimotility: Yes - 31 (no details given).</p> <p>Prokinetics: Not stated.</p> <p>Time before CDI treatment was stopped before FMT: Not stated.</p>	<p>Overall cure within stated follow up period: Primary cure on day 30 and 90 was achieved in 84.2% (<math>n=101/120</math>) and 78.3% (<math>n=72/92</math>).</p> <p>Cure with one infusion alone: No diarrhoea at 30 days in 84.2% (<math>n=101/120</math>); no diarrhoea at 90 days in 78.3% (<math>n=72/92</math>).</p> <p>Total follow up period: Median follow up 141 days (IQR 50-353 days).</p>	<p>Minor GI adverse events: x5 nausea, x3 abdominal pain, 2 belching, x2 vomiting, x2 'food intolerance', x1 IBS.</p> <p>Minor non-GI adverse events: x3 fever, x2 throat discomfort.</p> <p>Serious adverse events: x1 aspiration pneumonia, x1 haemorrhage (during endoscopy - no details), x1 loss of tooth, x1 polyneuropathy, x1 weight gain &gt; 10kg in 12 months post-FMT.</p> <p>Deaths: x7 died during follow up, x2 within 90 days of FMT. In x6 cases, definitely not related to CDI (in one patient, recurrence of CDI one week after FMT contributed to her death (but</p>	<p>Selection/eligibility reported: Yes.</p> <p>Consecutively recruited: Not clear.</p> <p>Prospectively recruited: No.</p> <p>Loss to follow up explained: No.</p> <p>At least 90% followed up: Yes.</p>
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Supplementary Material 2 for Gut

	fidaxomicin, x11 unknown.				stroke described as primary cause of death).	
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Supplementary Material 2 for *Gut*

<p>Hamilton <i>et al</i>, <i>American Journal of Gastroenterology</i>, 2012</p>	<p>Case series.</p> <p>Number of patients: 43.</p> <p>Female: male: 31: 12.</p> <p>Age (mean/ standard deviation): Mean 59 (+/- 21) years.</p> <p>Comorbidities: x14 IBD patients.</p> <p>CDI features: Recurrent.</p> <p>CDI diagnosis confirmation: Toxin positive with at least two subsequent recurrences.</p> <p>Pre-FMT antibiotics: All had vancomycin, 17 patients had addition of vancomycin and 2 weeks of rifaximin (one of these 17 had 4 weeks of rifaximin); 3 patients took 2-4 weeks of nitazoxanide.</p>	<p>Donors were standard donors for 33 FMTs, and individual donors for 10 FMTs.</p> <p>Donors working in healthcare: Not stated.</p> <p>Donor demographics: Not stated.</p> <p>Donor screening: Questionnaire - before recruitment, the donors were required to submit available medical records and have a separate medical history interview away from the recipient patient. The history included assessment of infectious risk, including identification of known risk factors for HIV and hepatitis, current communicable diseases, and recent travel to areas of the world with a higher prevalence of diarrheal illnesses.</p> <p>Travel and antibiotic exclusion period: Excluded as donors if recent travel to areas where high prevalence of diarrheal illness (not specified), and/ or antibiotic use within the past six months.</p>	<p>Amount of stool per transplant / administered to patients: 50g.</p> <p>Diluent used to prepare: 250ml sterile, non-bacteriostatic normal saline.</p> <p>Diluent used to store if frozen: 10% glycerol.</p> <p>Preparation methods: Stool from individual donors was passed through stainless steel tea strainers; stool from universal donors was transported on ice to the lab, and processed within 2 hours. Material was weighed and homogenised in commercial blender under nitrogen gas. Slurry then passed through 2.0, 1.0, 0.5 and 0.25mm stainless steel lab sieves. The resulting material was then centrifuged at 6000 x g for 15 minutes and resuspended to one-half the original volume in normal saline.</p> <p>Time from preparation to transplant (fresh): 1-2 hours.</p> <p>Time period for storage (frozen): 1-8 weeks.</p> <p>Route administered: Upper GI: nil; lower GI: colonoscopy (with majority into terminal ileum or caecum, with a small proportion into other colonic areas) in all 43; capsules: nil.</p> <p>Number of infusions: 1x FMT in 37</p>	<p>Overall cure within stated follow up period: 95% (n=41/43) within 2 months follow-up.</p> <p>Cure with one infusion alone: 86% (n=37/43).</p> <p>Total follow up period: 2 months following FMT.</p>	<p>Minor GI adverse events: ~1/3 of patients reported flatulence and excessive bowel movements within fortnight following procedure.</p> <p>Minor non-GI adverse events: None.</p> <p>Serious adverse events: None.</p> <p>Deaths: None.</p>	<p>Selection/ eligibility reported: Yes.</p> <p>Consecutively recruited: Yes.</p> <p>Prospectively recruited: No.</p> <p>Loss to follow up explained: No.</p> <p>At least 90% followed up: Yes.</p>
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Supplementary Material 2 for Gut

		<p>Screening blood tests: HIV, hepatitis B/C, RPR, LFTs.</p> <p>Screening stool tests: <i>Clostridium difficile</i> toxin B PCR, MC&amp;S, ova, cysts and parasites, <i>Giardia</i>, <i>Cryptosporidium</i>, <i>H pylori</i> antigen.</p>	<p>patients, 2x FMT in 6 patients.</p> <p>Bowel purgative: Yes - GoLYTELY or Moviprep.</p> <p>PPI: Not described.</p> <p>Antimotility: Not described.</p> <p>Prokinetics: Not described.</p> <p>Time before CDI treatment was stopped before FMT: 2 days.</p>			
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Supplementary Material 2 for *Gut*

<p>Hefazi <i>et al</i>, <i>Mayo Clinic Proceedings</i>, 2017</p>	<p>Case series.</p> <p>Number of patients: 23.</p> <p>Female: male: 13: 10.</p> <p>Age (median): 66 (range 23-88) years.</p> <p>Comorbidities: x13 patients had haematological malignancy (x4 diffuse large B cell lymphoma, x2 Hodgkin's lymphoma, x1 chronic myeloid leukaemia, x1 follicular lymphoma, x1 stage IV cutaneous T cell lymphoma, x1 B cell acute lymphocytic leukaemia, x1 hairy cell leukaemia, x1 chronic lymphocytic leukaemia, x1 severe aplastic anaemia); x1 with active disease at time of FMT, x2 with recent chemotherapy use, x2 with neutropenia within 12 weeks prior to FMT. x10 patients with solid organ malignancy (x4 breast, x2 anal, x1 colon, x1 pancreatic, x1 tonsillar, x1 non-small</p>	<p>Donors: Fresh stool from family/ friends in 10 patients, frozen stool from standard donors in 13 patients.</p> <p>Donor working in healthcare: Not stated.</p> <p>Donor demographics: Not stated.</p> <p>Donor screening: As per Patel <i>et al</i>, <i>Mayo Clin Proc</i>, 2013.</p> <p>Travel and antibiotic exclusion period: As per Patel <i>et al</i>, <i>Mayo Clin Proc</i>, 2013.</p> <p>Screening blood tests: As per Patel <i>et al</i>, <i>Mayo Clin Proc</i>, 2013.</p> <p>Screening stools: As per Patel <i>et al</i>, <i>Mayo Clin Proc</i>, 2013.</p>	<p>Amount of stool per transplant / administered to patients: ~50g.</p> <p>Diluent used to prepare: 250ml normal saline.</p> <p>Diluent used to store if frozen: Not stated.</p> <p>Preparation methods: As per Patel <i>et al</i>, <i>Mayo Clin Proc</i>, 2013.</p> <p>Time from preparation to transplant (fresh): Not stated.</p> <p>Time period for storage (frozen): Not stated.</p> <p>Route administered: Upper GI: nil; lower GI: All 23 patients received FMT via colonoscopy into caecum.</p> <p>Number of infusions: 1 FMT.</p> <p>Bowel purgative: Not stated.</p> <p>PPI: Not stated.</p> <p>Antimotility: Not stated.</p> <p>Prokinetics: Not stated.</p> <p>Time before CDI treatment was stopped before FMT: 24 hours.</p>	<p>Overall cure within stated follow up period: 92% (<math>n=11/12</math>) of haematological malignancy patients (other patient died), and 805 (<math>n=8/10</math>) solid malignancy patients.</p> <p>Cure with one infusion alone: 86% (<math>n=19/22</math>) by primary outcome criteria.</p> <p>Total follow up period: x1 CLL patient recurred at 22 months post-FMT in context of ibrutinib and coamoxiclav; successfully treated with 10 days of metronidazole. x1 tonsillar cancer patient had CDI recurrence at 14 months after exposure to cefalexin; successfully treated with 10 days of</p>	<p>Minor GI adverse events: x3 chronic diarrhoea for at least six months (despite negative <i>C difficile</i> laboratory tests), x8 transient diarrhoea, x3 abdominal cramps, x2 faecal urgency, x2 constipation, x1 nausea.</p> <p>Minor non-GI adverse events: None.</p> <p>Serious adverse events: None.</p> <p>Deaths: x1 death after cardiac arrest of Hodgkin's lymphoma patient at day 5 (multiple medical comorbidities thought likely cause, not FMT); x2 deaths at &gt; 60 days related to the underlying malignancy progressing.</p>	<p>Selection/ eligibility reported: Yes.</p> <p>Consecutively recruited: Yes.</p> <p>Prospectively recruited: No.</p> <p>Loss to follow up explained: Yes.</p> <p>At least 90% followed up: Yes.</p>
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Supplementary Material 2 for *Gut*

	<p>cell lung. x5 with metastasis at time of FMT, x3 recent chemotherapy use, x1 with recent neutropenia. Other comorbidities include x1 COPD, x1 ESKD on haemodialysis, x1 graft versus host disease (on immunosuppression), x1 granulomatosis with polyangiitis (Wegener's) on immunosuppression, x1 hypogammaglobulinaemia on intravenous immunoglobulin, x1 inflammatory arthritis on corticosteroids.</p> <p>CDI features: All recurrent.</p> <p>CDI diagnosis confirmation: Not explicitly defined, but definitions of recurrent, severe and complicated CDI as per American College of Gastroenterology.</p> <p>Pre-FMT antibiotics: All given additional vancomycin until 24hrs</p>			<p>vancomycin then 10 days of fidaxomicin. N.B. In all - x10 more chemotherapy courses and x8 more antibiotic courses after FMT.</p>		
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Supplementary Material 2 for *Gut*

	prior to FMT. Median of 2.5 standard treatment courses per patient (defined as at least 10 days of metronidazole, vancomycin or fidaxomicin), x1 previous vancomycin taper, and x4 total treatment courses for CDI).					
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Supplementary Material 2 for Gut

Hirsch et al, BMC Infectious Diseases, 2015	Case series.	Donors were 3 unrelated participants.	Amount of stool per transplant / administered to patients: 2.3g.			
	Number of patients: 19.	Donors working in healthcare: Not stated.	Diluent used to prepare: 350ml in 0.9% normal saline.			
	Female: male: 13: 6.		Diluent used to store if frozen: 15% glycerol.			
	Age (mean): 61 (range 26-92) years.	Donor demographics: Excluded if BMI>25, diabetes mellitus, psychiatric history, IBD, or IBS.	Preparation methods: Strict environmental control <6 hours after defaecation. All sterile, wet weight of stool was homogenised in 350ml 0.9% normal saline and aliquoted; samples were then centrifuged at 200 x g for 10 mins. Supernatant was decanted and centrifuged at 4600 x g for 15 minutes. supernatant removed and pellet re-suspended in 0.9% normal saline with glycerol. The typical concentration was 0.5g/ml. The resulting FMT slurry was put in 5-10ml syringes and frozen at -80°C.	Overall cure within stated follow up period: 68% (n=13/19).	Minor GI adverse events: x5 abdominal pain 5 (x4 self-resolved; x1 required opiates and was hospitalised).	Selection/ eligibility reported: Yes.
	Comorbidities: x3 IBS, x2 diabetes mellitus, x1 diverticulitis, x1 lymphoma, x1 acute myeloid leukaemia, x1 renal cancer, x1 chronic renal failure.	Donor screening: Questionnaire - standard questionnaire, with details as above.		Cure with one infusion alone: 68% (n=13/19) at 90 days.	Minor non-GI adverse events: None.	Consecutively recruited: Not clear.
	CDI features: Refractory and recurrent (2 or more episodes).	Travel and antibiotic exclusion period: Excluded if travel outside the USA within 30 days prior to donation, and/ or use of antibiotics within the past 6 months.		Total follow up period: Primary outcome assessed at 90 days, whilst secondary outcome assessed at 6 weeks after this.	Serious adverse events: None.	Prospectively recruited: No.
	CDI diagnosis confirmation: Not stated.	Screening blood tests: HIV, hepatitis A, B,C, <i>Treponema</i> /syphilis, and HTLV-1.	Time from preparation to transplant (fresh): N/A.		Deaths: x1 died from respiratory failure after failing FMT treatment.	Loss to follow up explained: No.
	Pre-FMT antibiotics: metronidazole, vancomycin +/- fidaxomicin.	Screening stool tests: <i>Clostridium difficile</i> toxin B, <i>Salmonella</i> , <i>Shigella</i> , <i>Campylobacter</i> , <i>E. coli</i> , <i>Yersinia</i> , <i>Vibrio</i> , <i>Aeromonas</i> , <i>Plesiomonas</i> .	Time period for storage (frozen): 1-3 weeks at -80°C; prior to use, syringes were transferred to -20°C and used within six weeks.			At least 90% followed up: Yes.
			Route administered: Nil upper or lower GI; all capsules. Aliquots of 0.4 mL of FMT slurry were dispensed into Size 1 acid-resistant hypromellose capsules,			

Supplementary Material 2 for *Gut*

			<p>subsequently placed within Size 0 acid-resistant hypromellose capsules and then nested within Size 00 gelatin Caps. Capsules were administered immediately upon filling and capping.</p> <p>Number of infusions: One course was 8-12 capsules (one only took 6).</p> <p>Bowel purgative: Not described.</p> <p>PPI: Yes - evening and morning of procedure.</p> <p>Antimotility: Not described.</p> <p>Prokinetics: Yes - encouraged to drink 4 ounces of Kefir fermented milk product twice a day, and also given a list of prebiotics to consume for 3 days.</p> <p>Time before CDI treatment was stopped before FMT: On day prior to FMT.</p>			
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Supplementary Material 2 for Gut

Ianiro <i>et al</i> , <i>Clinical Microbiology and Infection</i> , 2017	Case series.	Donors were unrelated for 36 FMTs, and related for 28 FMTs..	Amount of stool per transplant / administered to patients: not reported.			
	Number of patients: 64.		Diluent used to prepare: 500ml of 0.9% saline.			
	Female:male: 39: 25.	Donor working in healthcare: No.	Diluent used to store if frozen: N/A – fresh.			
	Age (mean): Mean 74 years.	Donor demographics: Not specified.	Preparation methods: After dilution, the solution was blended and supernatant strained and poured into sterile container.			
	Comorbidities: Not reported.	Donor screening: As per Cammarota <i>et al</i> , <i>Alim Pharm Ther</i> , 2015.	Time from preparation to transplant (fresh): 6 hours.	Overall cure within stated follow up period: 975 (n=62/64) at 8 weeks.	Minor GI adverse events: Not specified.	Selection/ eligibility reported: Yes.
	CDI features: Recurrent CDI - all patients had 3 recurrences on average range (range 2-6).	Travel and antibiotic exclusion period: As per Cammarota <i>et al</i> , <i>Alim Pharm Ther</i> , 2015.	Time period for storage (frozen): Not specified.	Cure with one infusion alone: 69% (n=44/64).	Minor non-GI adverse events: Not specified.	Consecutively recruited: Yes.
	CDI diagnosis confirmation: Defined using ESCMID guidelines.	Screening blood tests: As per Cammarota <i>et al</i> , <i>Alim Pharm Ther</i> , 2015.	Route administered: Upper GI: nil: lower GI: all 64 given FMT via colonoscopy; capsules: nil.	Total follow up period: 8 weeks.	Serious adverse events: Not specified.	Prospectively recruited: No.
	Pre-FMT antibiotics: All patients had had prior metronidazole, vancomycin and/ or fidaxomicin.	Screening stool tests: As per Cammarota <i>et al</i> , <i>Alim Pharm Ther</i> , 2015.	Number of infusions: 44 patients had x1 FMT, 20 patients had >1 FMT (undefined).		Deaths: Not specified.	Loss to follow up explained: Yes.
			Bowel purgative: 4l macrogol on last 1-2 days of antibiotics treatment.			At least 90% followed up: Yes.
			PPI: Not specified.			
			Antimotility: Not specified.			
			Prokinetics: Not specified.			

Supplementary Material 2 for *Gut*

Time before CDI treatment was stopped  
before FMT: FMT given on last 1 or two  
days of CDI treatment.

Supplementary Material 2 for Gut

Kassam <i>et al</i> , <i>Archives of Internal Medicine</i> , 2012	Case series.					
	Number of patients: 27.			Amount of stool per transplant / administered to patients: 150g of stool.		
	Female: male 13: 14.	Donors were two healthy volunteers.		Diluent used to prepare: 300mls sterile water.		
	Age (mean): 69.4 (range 26-87) years.	Donors working in healthcare: Not specified.		Diluent used to store if frozen: N/A.		
	Comorbidities: Not specified.	Donor demographics: Not specified.		Preparation methods: Not specified.	Overall cure within stated follow up period: 81% (n=22/27).	Minor GI adverse events: Not specified.
	CDI features: Recurrent and refractory.	Donor screening: Questionnaire - not specified.		Time from preparation to transplant (fresh): Not specified.		Selection/ eligibility reported: Yes.
	CDI diagnosis confirmation: (1) Laboratory-confirmed <i>C difficile</i> toxin using EIA with no other cause for diarrhea; (2) refractory CDI (defined as ongoing diarrhea despite antimicrobial treatment) or recurrent CDI (defined as symptom resolution for at least 2 days after discontinuation of treatment with recurrence of diarrhea).	Travel and antibiotic exclusion period: Excluded if used antibiotics within last 6 months.		Time period for storage (frozen): N/A – fresh.	Cure with one infusion alone: 81% (n=22/27).	Consecutively recruited: Yes.
		Screening blood tests: Hepatitis B surface antigen, hepatitis C antibody, <i>Helicobacter pylori</i> and syphilis serologic markers, HIV types -1 and -2, and HTLV types -I and -II.		Route administered: Upper GI: nil; lower GI: 27 via retention enema.		Prospectively recruited: No.
		Screening stool tests: Stool was processed for enteric bacterial pathogens, <i>C difficile</i> toxin, and ova and parasites.		Number of infusions: 1 enema in 22 patients, 2 enemas in 5 patients.	Total follow up period: Mean follow-up of 427.3 days after transplant.	Loss to follow up explained: Yes.
	Pre-FMT antibiotics: All had at least prior metronidazole; 19 had subsequent vancomycin monotherapy. 8 had			Bowel purgative: Not specified.	Serious adverse events: Not specified.	At least 90% followed up: Yes.
				PPI: Not specified.	Deaths: Not specified.	
				Antimotility: Not specified.		
				Prokinetics: Not specified.		
				Time before CDI treatment was stopped before FMT: At least 24 hours before.		

Supplementary Material 2 for *Gut*

	combination metronidazole and vancomycin therapy.					
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Supplementary Material 2 for Gut

Kelly et al, Journal of Clinical Gastroenterology, 2012	Case series.	Donors were family members in 25 cases, and friend in 1 case.	Amount of stool per transplant / administered to patients: "6:8 tablespoons of donor stool".			
	Number of patients: 26.					
	Female: male: 24:2.	Donor working in healthcare: No.	Diluent used to prepare: 1 litre of sterile water passed through gauze. Aliquoted in 60ml syringes.			
	Age (mean): 59 years.	Donor demographics: Not specified.	Diluent used to store if frozen: N/A – fresh.			
	Comorbidities: Not stated.	Donor screening: Questionnaire – asked regarding known exposure to HIV within 12 months, high-risk sexual behaviours, use of illicit drugs, tattoo within 6 months, incarceration within 12 months, risk factors for Creutzfeldt-Jakob disease, GI co-morbidities, recent ingestion of allergen, systemic autoimmunity, chronic pain syndromes.	Preparation methods: As above.	Overall cure within stated follow up period: 92.3% (n=24/26).	Minor GI adverse events: Mild diarrhoea post-FMT in x3 patients.	Selection/ eligibility reported: Yes.
	CDI features: Recurrent. Mean duration of diagnosis of CDI prior to FMT of 12.6 (range 4 to 84) months.		Time from preparation to transplant (fresh): 6 hours prior to transplant.			Consecutively recruited: Yes.
	CDI diagnosis confirmation: Not stated.	Travel and antibiotic exclusion period: No antibiotics for preceeding 90 days.	Time period for storage (frozen): N/A.	Cure with one infusion alone: 92.3% (n=24/26).	Minor non-GI adverse events: No.	Prospectively recruited: No.
	Pre-FMT antibiotics: All had previous treatment with metronidazole, and repeated tapering courses of vancomycin. 19 had failed at least one course of rifaximin. Some patients had prior <i>Saccharomyces boulardii</i> or <i>Lactobacillus</i> GG. Pre-FMT, all had 2 weeks of metronidazole or vancomycin, discontinued 2-3 days before FMT.	Screening blood tests: blood for hepatitis A, B and C, HIV-1&-2, <i>Treponema pallidum</i> .	Route administered: Upper GI: nil; lower GI: all 26 via colonoscopy; capsules: nil.	Total follow up period: follow up of mean 10.7 months (ranged from 2-30 months).	Serious adverse events: No.	Loss to follow up explained: Yes.
		Screening stool tests: Stool for culture for bacteria, stain for ova and parasites, <i>C difficile</i> toxin A and B.	Number of infusions: not explicitly stated but implies single infusion for all patients.	Deaths: No.		At least 90% followed up: Yes
			Bowel purgative: PEG bowel prep night before transplant.			
			PPI: Not stated.			
			Antimotility: Not stated.			
			Prokinetics: Not stated.			
			Time before CDI treatment was stopped			

Supplementary Material 2 for *Gut*

before FMT: 2-3 days.

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Supplementary Material 2 for Gut

Kelly et al, <i>American Journal of Gastroenterology</i> , 2014	Case series.				Minor GI adverse events: x3 self limiting diarrhoea, x3 bloating and abdominal discomfort, x1 Crohn's flare, x1 nausea, x1 minor mucosal tear at colonoscopy.	
	Number of patients: 80.		Amount of stool per transplant / administered to patients: Varied by centre.			
	Female: male: 42: 38.		Diluent used to prepare: Varied by centre.			
	Age (mean): N.B. 75 adults, and 5 children.		Diluent used to store if frozen: Varied by centre.			
	Mean age of adults: 53 (range 20-88) years; mean age of paediatric patients: 10.9 (range 6.5-16) years.	Donors working in healthcare: Not specified.	Preparation methods: Varied by centre.	Overall cure within stated follow up period: 89% (n=71/80) within a minimum of 12 weeks.	Minor non-GI adverse events: x1 fever, x1 hip pain, x1 pertussis.	Selection/ eligibility reported: Yes.
	Comorbidities: x36 IBD, x19 solid organ transplant, x3 HIV/AIDS, x7 cancer, x4 rheumatoid arthritis, x1 adrenal insufficiency, x6 cirrhosis, x1 ESKD, x1 panhypopituitarism, x1 end-stage COPD, x1 ESKD with allograft failure, x1 Sjögrens.	Donor demographics: Not specified.	Time from preparation to transplant (fresh): Varied by centre.	Cure with one infusion alone: 78% (n=62/80).	Serious adverse events: x10 hospitalization (x1 for fever, encephalopathy and pancytopenia; x1 abdo pain post FMT, x3 IBD flares (x2 Crohn's, x1 UC), x1 stroke, x1 colectomy, x1 fall and sustained hip fracture, x1 influenza B and diarrhoea, x1 catheter infection.	Consecutively recruited: No.
	CDI features: Both refractory and recurrent patients included as well as severe/ complicated disease.	Donor screening: Questionnaire: Varied by centre.	Time period for storage (frozen): Varied by centre.	Total follow up period: 12 weeks post-FMT.		Prospectively recruited: No.
	CDI diagnosis: Not clearly specified.	Travel and antibiotic exclusion period: Varied by centre.	Route administered: Not specified.			Loss to follow up explained: No.
	Pre-FMT antibiotics:	Screening blood tests: Varied by centre.	Number of infusions: 85% (n=68/80) had single FMT, 15% (n=12/80) had > 1 FMT.			At least 90% followed up: Yes.
		Screening stool tests: Varied by centre.	Bowel purgative: Varied by centre.			
			PPI: Varied by centre.			
			Antimotility: Varied by centre.			
			Prokinetics: Varied by centre.			
			Time before CDI treatment was stopped before FMT: Varied by centre.		Deaths: x2 deaths (x1 pneumonia and x1 aspiration after	

Supplementary Material 2 for *Gut*

	Vancomycin 67 (84%), fidaxomicin 23 (29%), rifaximin 13 (16%), metronidazole 55 (69%).				sedation for colonoscopic FMT).	
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Supplementary Material 2 for Gut

Khoruts <i>et al</i> , <i>Clinical Gastroenterology &amp; Hepatology</i> , 2016	Case series.		Amount of stool per transplant / administered to patients: As per Hamilton <i>et al</i> , <i>Am J Gastroenterol</i> , 2012.		Minor GI adverse events: Not specified.	
	Number of patients: 272.  Female: male: 189: 83.  Age (mean/ median/ standard deviation): Mean 57.2 (+/- 19.2) years; median 59.0 (range 16-100) years.  Comorbidities: x10 dialysis, x22 established Crohn's, x21 established UC, x15 lymphocytic colitis, x5 diagnosed with Crohn's during colonoscopy for FMT, x1 diagnosed UC during colonoscopy for FMT, x14 newly-diagnosed lymphocytic colitis. x13 reclassified in terms of IBD. x8 solid organ recipients, x30 patients without IBD were taking biologics (anti-TNF, rituximab), immunomodulators (methotrexate, purine analogues), and/ or corticosteroids.  CDI features: All patients had at least two	Donors working in healthcare: As per Hamilton <i>et al</i> , <i>Am J Gastroenterol</i> , 2012.  Donor demographics: As per Hamilton <i>et al</i> , <i>Am J Gastroenterol</i> , 2012.  Donor screening: Questionnaire - as per Hamilton <i>et al</i> , <i>Am J Gastroenterol</i> , 2012.  Travel and antibiotic exclusion period: As per Hamilton <i>et al</i> , <i>Am J Gastroenterol</i> , 2012.  Screening blood tests: As per Hamilton <i>et al</i> , <i>Am J Gastroenterol</i> , 2012.  Screening stools: As per Hamilton <i>et al</i> , <i>Am J Gastroenterol</i> , 2012.	Diluent used to prepare: As per Hamilton <i>et al</i> , <i>Am J Gastroenterol</i> , 2012.  Diluent used to store if frozen: As per Hamilton <i>et al</i> , <i>Am J Gastroenterol</i> , 2012.  Preparation methods: As per Hamilton <i>et al</i> , <i>Am J Gastroenterol</i> , 2012.  Time from preparation to transplant (fresh): As per Hamilton <i>et al</i> , <i>Am J Gastroenterol</i> , 2012.  Time period for storage (frozen): As per Hamilton <i>et al</i> , <i>Am J Gastroenterol</i> , 2012.  Route administered: Upper GI: nil; lower GI: colonoscopy (272); capsule: nil.  Number of infusions: One routinely, more than one if required - specific criteria not defined.  Bowel purgative: Yes - all had purgative on day prior to procedure (as per Hamilton <i>et al</i> , <i>Am J Gastroenterol</i> , 2012).	Overall cure within stated follow up period: 74% (n= 32/43) in IBD patients and 92.2% (n=211/229) in non-IBD patients.  Cure with one infusion alone: 74% (n= 32/43) in IBD patients and 92.2% (n=211/229) in non IBD patients.  Total follow up period: Up to 6 years.	Minor non-GI adverse events: Not specified.  Serious adverse events: 25.6% (n=11/43) of IBD patients diagnosed with FMT-related flare. x2 patients hospitalised with IBD flare within two months of FMT. Clearance of CDI by FMT generally associated with improved control of IBD over the long term. x6 patients struggled with IBD despite optimisation of immunosuppressive treatment, x3 of whom underwent colectomies.  Deaths: Nil.	Selection/ eligibility reported: Yes.  Consecutively recruited: Yes.  Prospectively recruited: No.  Loss to follow up explained: Yes.  At least 90% followed up: Yes.

Supplementary Material 2 for *Gut*

	<p>spontaneous relapses of CDI following initial episode, defined as recurrence within three months of discontinuation of anti-CDI antibiotics treatment in conjunction with diarrheal symptoms.</p> <p>CDI diagnosis confirmation: Positive stool testing within two months of FMT - not clearly defined.</p> <p>Pre-FMT antibiotics: x206 patients had had prior metronidazole, x270 vancomycin, x69 fidaxomicin, x71 rifaximin, x104 probiotics.</p>		<p>PPI: Not described.</p> <p>Antimotility: Not described.</p> <p>Prokinetics: Not described.</p> <p>Time before CDI treatment was stopped before FMT: 2 days.</p>			
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Supplementary Material 2 for Gut

Lagier <i>et al</i> , <i>European Journal of Clinical Microbiology and Infectious Diseases</i> , 2015	Case series.		Amount of stool per transplant / administered to patients: >30g.			
	Number of patients: 61.					
	Female: male: 40:21.	Donors were preferentially healthy family members, but also used healthy volunteer students and residents.	Diluent used to prepare: Whole stool mixed with 400ml normal saline, homogenised for 10 minutes.	Overall cure within stated follow up period: Global death rate of 19% ( <i>n</i> =3/16) in early transplant arm (day 20, day 37, day 166),	Minor GI adverse events: x24 diarrhoea (resolved day 1 after FMT), x1 nausea.	
	Age (mean): 84 (range 66-101) years.	Donor working in healthcare: Yes - some residents.	Diluent used to store if frozen: N/A – fresh.	67% ( <i>n</i> =2/3) died in arm of those treated by tardive transplant (day 28, day 54).	Minor non-GI adverse events: Not specified.	Selection/ eligibility reported: Yes.
	Comorbidities: Not Specified.	Donor demographics: BMI<30, exclude active cancer, diarrhoea, current immunosuppressive drugs, antibiotics within past three months.	Preparation methods: 10 minutes of homogenisation in blender, filtered, put into a syringe at room temperature.	None of these patients died with evidence of CDI.	Serious adverse events: x1 acute heart failure - no details.	Consecutively recruited: No - not stated.
	CDI features: Some patients refractory/ recurrent; some during first CDI.	Donor screening: Questionnaire: As above.	Time from preparation to transplant (fresh): <6 hours.	Cure with one infusion alone: 33% ( <i>n</i> =1/3) treated by tardive FMT dead at day 31; 4.2% ( <i>n</i> =1/16) treated by early FMT dead at day 31.	Deaths: 3/16 in early transplant arm (vs 29/45 treated by abx only or tardive transplant). No sign of CDI at time of death (days 20, 37, 166).	Prospectively recruited: No.
	CDI diagnosis confirmation:PCR that detects toxin and B genes, and toxin C gene deletion that characterises 027.	Travel and antibiotic exclusion period: Excluded as donor if antibiotic use within past three months.	Route administered: Upper GI: Via nasogastric tube in 61 patients; nil lower GI or capsules.	Total follow up period: No details on absolute length of follow-up.		Loss to follow up explained: Yes.
	Pre-FMT antibiotics: Patients divided into 'tardive transplant' (i.e. only after x3 antibiotic failures) or 'early transplant' (during first week of infection during first treatment, accompanied by antibiotics). Antibiotics were for non-severe disease: metronidazole	Screening blood tests: HIV, hepatitis A, B,C, E, active CMV, active EBV, <i>Treponema pallidum</i> , HTLV.	Number of infusions: In early FMT arm - one FMT routine; but offered 2nd FMT if relapse.			At least 90% followed up: Yes.
		Screening stool tests: MC&S, parasites, toxigenic <i>C difficile</i> .	Bowel purgative: 4l Klean Prep/ two glasses of Fast Prep day before FMT.			
			PPI: No - but used 200ml 1.4% bicarbonate 15 minutes before FMT.			
			Antimotility: Not specified.			
			Prokinetics: Not specified.			

Supplementary Material 2 for *Gut*

	orally three times a day for 14 days, then vancomycin 125mg four times a day for 14 days, then fidaxomicin 200mg twice a day for 10 days; for severe disease (defined as AKI, paralytic ileus, or peritoneal fluid), used vancomycin and metronidazole for primary infection, then fidaxomicin if relapse/failure.		Time before CDI treatment was stopped before FMT: Not specified.			
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Supplementary Material 2 for Gut

Lee et al, European Journal of Clinical Microbiology and Infectious Diseases, 2014	Case series.		Amount of stool per transplant / administered to patients: Not specified.			
	Number of patients: 94		Diluent used to prepare: 300ml water.		Minor GI adverse events: "10% experienced transient constipation and excess flatulence post-FMT".	
	Female: male: 53: 41.	Donors were volunteers.	Diluent used to store if frozen: N/A – fresh.		Minor non-GI adverse events: None described.	Selection/ eligibility reported: Yes.
	Age (mean): Mean 71.8 (range 24-95) years.	Donor working in healthcare: Not specified	Preparation methods: Homogenisation of stool in water using a disposable spatula.	Overall cure within stated follow up period: At 6 months – 87% (n=81/94) in remission after FMT.	Serious adverse events: None described.	Consecutively recruited: Yes.
	Comorbidities: x3 IBD, x3 post-renal transplant.	Donor demographics: Not specified.	Time from preparation to transplant (fresh): Not specified.	Cure with one infusion alone: 47.9% (n=45/94) with single FMT in remission at 6 months.	Deaths: 75% (n=6/8) patients not responding to FMT died (not clear when). All "over 70 years of age", with multiple underlying significant comorbidities and passed away due to critical illnesses; none had deaths attributable to FMT or directly due to CDI.	Prospectively recruited: No.
	CDI features: Some patients refractory (defined as ongoing diarrhea despite treatment with at least 5 days of oral vancomycin, 125mg four times daily), or recurrent (symptom resolution for at least two days after the discontinuation of treatment with recurrence of diarrhoea.	Donor screening: Questionnaire - describes use of questionnaire but no details given - "similar to the Full Length Donor History Questionnaire documents (US Food and Drug administration, DHQ version 1.3, May 2008"	Time period for storage (frozen): N/A.	Total follow up period: 24 months.		Loss to follow up explained: Yes.
	CDI diagnosis confirmation: Toxin positive by enzyme immunoassay or polymerase chain reaction.	Travel and antibiotic exclusion period: Not specified.	Route administered: Upper GI: nil; lower GI: retention enema in all 94 patients; nil capsules.			At least 90% followed up: Yes.
		Screening blood testss: HIV-1/-2, HTLV-1 and -2. Hepatitis A IgG/M, hepatitis B surface antigen, hepatitis C antibody, Treponema pallidum.	Number of infusions: No fixed number - as many as required to achieve remission. No clear definition of non-response.			
		Screening stools: Ova, cysts and parasites, MC&S, C difficile toxin, norovirus, adenovirus, rotavirus.	Bowel purgative: Not specified.			
	Pre-FMT antibiotics: Average of 2.1 previous anti-CDI antibiotic		PPI: Not specified.			
			Antimotility: Not specified.			
			Prokinetics: Not specified.			
			Time before CDI treatment was stopped before FMT: Not specified.			

Supplementary Material 2 for *Gut*

	courses (range 1-4), specifically: x74 metronidazole courses (79.3%), x71 vancomycin (75%), x14 vancomycin taper (15.2%), x3 probiotic monotreatment (0.03%), x16 concomitant metronidazole/vancomycin (17.4%).					
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Supplementary Material 2 for Gut

MacConnachie <i>et al, QJM</i> , 2009	<p>Case series.</p> <p>Number of patients: 15.</p> <p>Female: male: 14: 1.</p> <p>Age (median): 81.5 (range 68-95) years.</p> <p>Comorbidities: no haematological or IBD.</p> <p>CDI features: Relapsing defined as recurrence of loose stool following successful antibiotic treatment in a patient with previous toxin positive CDI.</p> <p>CDI diagnosis confirmation: Not specified.</p> <p>Pre-FMT antibiotics: All had had previous metronidazole and vancomycin; x3 patients tapering vancomycin and intravenous Immunoglobulin.</p>	<p>Donors were healthy related volunteers.</p> <p>Working in healthcare: Yes – in three cases where relatives could not be identified.</p> <p>Donor demographics: Not specified.</p> <p>Donor screening: HIV-1/-2, HTLV- 1 and -2, hepatitis A IgG/M, hepatitis B surface antigen, hepatitis C antibody, <i>Treponema pallidum</i>.</p> <p>Questionnaire: Yes, but not specified.</p> <p>Travel and antibiotic exclusion period: Not specified.</p> <p>Screening stools: Ova, cysts and parasites, MC&amp;S, <i>C difficile</i> toxin.</p>	<p>Amount of stool per transplant administered to patients: 30g.</p> <p>Diluent used to prepare: 0.9% normal saline.</p> <p>Diluent used to store if frozen: N/A – fresh.</p> <p>Preparation methods: Stool sample prepared in less than 6 hours; add 50-70ml of normal saline, homogenise with handheld stool blender, gradually advance speed, continue for 2-4 mins until smooth, filter suspension in coffee filter paper.</p> <p>Time from preparation to transplant (fresh): 6 hours.</p> <p>Time period for storage (frozen): Not applicable.</p> <p>Route administered: Upper GI: All 15 patients received FMT via nasogastric tube; lower GI and capsules: nil.</p> <p>Number of infusions: 1 FMT per patient routinely, repeat if required.</p> <p>Bowel purgative: Not given.</p> <p>PPI: Omeprazole 20mg eve before and on morning.</p> <p>Antimotility: Not given.</p>	<p>Overall cure within stated follow up period: 84% (n=15/18) “resolution”.</p> <p>Cure with one infusion alone: 884% (n=15/18) “resolution”.</p> <p>Total follow-up period: 90 days.</p>	<p>Minor GI adverse events: x1 diarrhoea.</p> <p>Minor non-GI adverse events: Nil.</p> <p>Serious adverse events: Nil.</p> <p>Deaths: x2 (not felt related to FMT).</p>	<p>Selection/ eligibility reported: Yes.</p> <p>Consecutively recruited: Yes.</p> <p>Prospectively recruited: No.</p> <p>Loss to follow up explained: Yes.</p> <p>At least 90% followed up: Yes.</p>
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Supplementary Material 2 for *Gut*

Prokinetics: Not given.

Time before CDI treatment was stopped  
before FMT: Stopped on the evening  
before FMT.



Supplementary Material 2 for Gut

Mattila <i>et al</i> , <i>Gastroenterology</i> , 2012	Case series.	Donors: 61 donors were close relatives/ other household members; in 9 cases, healthy volunteers.	Amount of stool per transplant / administered to patients: 20-30ml stool.			
	Number of patients: 70.	Donors working in healthcare: Not specified.	Diluent used to prepare: 100-200ml water; 100ml of suspension administered to caecum.		Minor GI adverse events: Not specified.	
	Female: male: 42: 28.	Donor demographics: Not specified.	Diluent used to store if frozen: N/A – all fresh.	Overall cure within stated follow up period: 94% (n=66/70) (100% (n=34/34) of those with non-027, 89% (n=32/36) with 027) within 12 weeks.	Minor non-GI adverse events: Not specified.	Selection/ eligibility reported: Yes.
	Age (mean): Mean 73 (range 22-90) years.	Donor screening: Questionnaire - "No antibiotics and no intestinal symptoms within 6 months".	Preparation methods: Not specified.		Serious adverse events: Not specified.	Consecutively recruited: Not clear.
	Comorbidities: No IBD, one adenocarcinoma of colon diagnosed during colonoscopy for FMT.	Travel and antibiotic exclusion period: Excluded as donor if any antibiotic use within past six months; no details of travel restrictions.	Time from preparation to transplant (fresh): 6 hours.			
	CDI features: Recurrent, mean of 3.5 previous episodes of CDI pre-FMT (range 1-12).	Screening blood tests: Hepatitis B surface antigen, Hepatitis C antibody, HIV-1/-2 , <i>Treponema pallidum</i> plasma reagin test; total blood count, C-reactive protein, creatinine, liver enzymes.	Time period for storage (frozen): N/A.	Cure with one infusion alone: 94% (n=66/70) (100% (n=34/34) of those with non-027, 89% (n=32/36) with 027) within 12 weeks.	Deaths: x4 patients infected with 027 did not respond to FMT and died within 3 months. 10 other patients died of 'unrelated illnesses' during one year of follow-up.	Prospectively recruited: No.
	CDI diagnosis confirmation: Positive culture and toxin.	Screening stool tests: <i>C difficile</i> culture/ tox A/ B; MC&S, ova cysts and parasites.	Route administered: Upper GI: nil; lower GI: colonoscopy (70); capsules: nil.	Total follow up period: One year.		Loss to follow up explained: Yes.
	Pre-FMT antibiotics: Mixture of metronidazole, vancomycin, rifaximin - no patient-level data.		Number of infusions: 1 FMT.			At least 90% followed up: Yes.
			Bowel purgative: 4l PEG (Colonsteril).			
			PPI: Not specified.			

Supplementary Material 2 for *Gut*

<p>Meighani <i>et al</i>, <i>European Journal of Gastroenterology and Hepatology</i>, 2016</p>	<p>Case series.</p> <p>Number of patients: 201.</p> <p>Female: male: 125: 76.</p> <p>Age (mean/ standard deviation): Mean age 66.6 (+/-18.3) years.</p> <p>Comorbidities: x37 cancer, x30 immunosuppressed, x26 CKD. Immunosuppressed defined as chemotherapy within 1 year of FMT, HIV with CD4 &lt; 200, or prednisolone use greater than or equal to 20mg for more than 1 month.)</p> <p>CDI features: 61 with refractory, 140 with recurrent.</p> <p>CDI diagnosis confirmation: Positive toxin or polymerase chain reaction.</p> <p>Pre-FMT antibiotics: Not specified.</p>	<p>Donors working in healthcare: not specified.</p> <p>Donor demographics: not specified.</p> <p>Donor screening: Questionnaire - not specified.</p> <p>Travel and antibiotic exclusion period: Not specified.</p> <p>Screening blood tests: Not specified.</p> <p>Screening stool tests: Not specified.</p>	<p>Amount of stool per transplant / administered to patients: Not specified.</p> <p>Diluent used to prepare: Not specified.</p> <p>Diluent used to store if frozen: Not specified.</p> <p>Preparation methods: Not specified.</p> <p>Time from preparation to transplant (fresh): Not specified.</p> <p>Time period for storage (frozen): Not specified.</p> <p>Route administered: Upper GI: nasogastric tube x 76, PEG x5; lower GI: x45 enema, x75 colon; capsules: nil.</p> <p>Number of infusions: Some people received multiple FMT procedures - repeat FMTs within 90 days of previous FMT were still maintained as a 'single infection unit'.</p> <p>Bowel purgative: Not specified.</p> <p>PPI: Not specified.</p> <p>Antimotility: Not specified.</p> <p>Prokinetics: Not specified.</p>	<p>Overall cure within stated follow up period: 88% (n=176/201) over 90 days.</p> <p>Cure with one infusion alone: 73.1% (n=147/201).</p> <p>Total follow-up period: Each patient for 90 days.</p>	<p>Minor GI adverse events: Not specified.</p> <p>Minor non-GI adverse events: Not specified.</p> <p>Serious adverse events: Not described.</p> <p>Deaths: 18 deaths in cohort but no clear timeframe, and not clear if any related to FMT. Described as mortality rate of 6.25% in response group, 28% in failure rate.</p>	<p>Selection/ eligibility reported: Yes.</p> <p>Consecutively recruited: Yes.</p> <p>Prospectively recruited: No.</p> <p>Loss to follow up explained: Yes.</p> <p>At least 90% followed up: Yes.</p>
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Supplementary Material 2 for *Gut*

			Time before CDI treatment was stopped before FMT: 24 hour - not specifically stated as anti-CDI treatment.			
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Supplementary Material 2 for *Gut*

<p>Meighani <i>et al</i>, <i>Digestive Diseases and Sciences</i>, 2017</p>	<p>Case series.</p> <p>Number of patients: 201.</p> <p>Female: male: 124: 77.</p> <p>Age (mean/ standard deviation): Mean 68.79 (+/-16.78) years for x181 non-IBD patients, mean 46.9 (+/-19.97) for the x20 IBD patients.</p> <p>Comorbidities: 13/20 IBD patients were immunosuppressed (no further details); no further specific details about immunosuppression).</p> <p>CDI features: Recurrent CDI in 13/20 of IBD patients, primary refractory in 7/20. 1.90 (+/- 1.02) CDI infections in past three months for IBD patients, 1.79 (+/1.17) CDI infections in past three months for non-IBD patients.</p> <p>CDI diagnosis confirmation: GDH first, then toxin A and B; PCR</p>	<p>Donors were typically family members, but small number of unrelated universal donors. Amongst IBD cohort - 6 patients had family members as donor, universal donor in other 14.</p> <p>Donor working in healthcare: Not defined.</p> <p>Donor demographics: Not defined.</p> <p>Donor screening: Questionnaire - not defined.</p> <p>Travel and antibiotic exclusion period: Not defined.</p> <p>Screening blood tests: Not defined.</p> <p>Screening stool tests: Not defined.</p>	<p>Amount of stool per transplant / administered to patients: Not defined.</p> <p>Diluent used to prepare: Not defined.</p> <p>Diluent used to store if frozen: Not defined.</p> <p>Preparation methods: Not defined.</p> <p>Time from preparation to transplant (fresh): Not defined.</p> <p>Time period for storage (frozen): Not defined.</p> <p>Route administered: Upper GI: 5 nasogastric (IBD patients only; not described re non-IBD patients) lower GI: 13 colonoscopy (IBD patients only; not described in non-IBD patients); 2 retention enema (IBD patients only; not described re non-IBD patients) (15).</p> <p>Number of infusions: Any relapse beyond 90 days was defined as 'new infection'. However, not made clear if patients given more than one FMT.</p> <p>Bowel purgative: Not described.</p> <p>PPI: Not described.</p> <p>Antimotility: Not described.</p> <p>Prokinetics: Not described.</p>	<p>Overall cure within stated follow up period: As per primary outcome - difficult to give more specific information than already given.</p> <p>Cure with one infusion alone: 87.3% (<math>n=158/181</math>) in non-IBD, 75% (15/20) in IBD; but 17.15 (<math>n=31/181</math>) non-IBD relapse within 90 days/ 13.9% (<math>n=25/180</math>) beyond 90 days, and 25% (<math>n=5/20</math>) IBD relapse within 90 days/ 20% (<math>n=4/20</math>) beyond 90 days. 3/5 failures in IBD arm had newly-diagnosed IBD, other had severe active disease.</p> <p>Total follow up period: At least 90 days.</p>	<p>Minor GI adverse events: None.</p> <p>Minor non-GI adverse events: None.</p> <p>Serious adverse events: None.</p> <p>Deaths: None.</p>	<p>Selection/ eligibility reported: Yes.</p> <p>Consecutively recruited: Yes.</p> <p>Prospectively recruited: No.</p> <p>Loss to follow up explained: Yes.</p> <p>At least 90% followed up: Yes.</p>
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Supplementary Material 2 for *Gut*

	used if discordance.  Pre-FMT antibiotics: Not defined for non-IBD; for IBD, 15 vancomycin alone, 5 vancomycin and oral metronidazole.		Time before CDI treatment was stopped before FMT: No specific details.			
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Supplementary Material 2 for *Gut*

<p>Patel <i>et al</i>, <i>Mayo Clinic Proceedings</i>, 2013</p>	<p>Case series.</p> <p>Number of patients: 31.</p> <p>Female: male: 17: 14.</p> <p>Age (mean/ standard deviation): Mean 61.26 (+/- 19.34) years.</p> <p>Comorbidities: x5 diverticulosis, x5 IBS, x3 UC, x1 Crohn's, x1 gastroparesis, x1 coloanal fistula, x3 prev sigmoid surgery for diverticulitis, x2 subtotal colectomy with ileosigmoid anastomosis, x1 left hemicolectomy with colostomy, x3 long term corticosteroids, x2 hypogammaglobulinaemia, x1 OLT, x1 renal transplant, x1 long term methotrexate.</p> <p>CDI features: Recurrent - mean +/- SD number of confirmed relapses before FMT of 4 +/- 1.4 (range 2-7) episodes.</p> <p>CDI diagnosis confirmation: At least 3x unformed stools/ day, at</p>	<p>Donors were healthy family/ contacts of recipients - 14 spouses, 9 children, 5 siblings, 3 parents, 1 niece, 1 friend.</p> <p>Working in healthcare: Not stated.</p> <p>Donor demographics: No stated age/ BMI limits.</p> <p>Donor screening: Questionnaire - exclude if: chronic GI disease, active peptic ulcer disease, GORD requiring daily PPI, IBS, IBD, history of colon polyps/ cancer, antibiotics or hospitalisation in past three months.</p> <p>Travel and antibiotic exclusion period: No stated travel restrictions; excluded as donor if antibiotic use within past 3 months.</p> <p>Screening blood tests: hepatitis A IgM, HBsAg, HBc IgG/M, hepatitis C antibody, HIV-1/-2 antibody, HTLV-1/-2 antibody, RPR/ syphilis EIA.</p> <p>Screening stool tests: MC&amp;S, ova, cysts and parasites, <i>Cryptosporidium</i> antigen,</p>	<p>Amount of stool per transplant / administered to patients: Whole stool - median transplanted weight of 115g (range 18-397g).</p> <p>Diluent used to prepare: Normal saline - "added in 100ml increments until mixture suitable for instillation through working channel of colonoscope". Median volume of FMT 360 (range 180-900) ml.</p> <p>Diluent used to store if frozen: N/A – fresh.</p> <p>Preparation methods: Blender/ pitcher.</p> <p>Time from preparation to transplant (fresh): Six hours; kept at room temperature until processing.</p> <p>Time period for storage (frozen): N/A.</p> <p>Route administered: Upper GI: nil; lower GI: colonoscopy (31); capsule: nil.</p> <p>Number of infusions: One initially.</p> <p>Bowel purgative: Yes - PEG day before FMT.</p> <p>PPI: Not described.</p> <p>Antimotility: 4mg loperamide either pre- or immediately after colonoscopy.</p>	<p>Overall cure within stated follow up period: At 3 months – 91.3% (<i>n</i>=21/23) said diarrhoea no longer present; at 1 year, 100% (<i>n</i>=6/6) reported maintained improvement or resolution.</p> <p>Cure with one infusion alone: Of 29 with diarrhoea – 24.1% (<i>n</i>=7/29) reported improvement and 75.9% (<i>n</i>=22/29) resolution of diarrhoea by median time of three days.</p> <p>Total follow up period: One year.</p>	<p>Minor GI adverse events: Not described.</p> <p>Minor non-GI adverse events: Not described.</p> <p>Serious adverse events: Microperforation - caused by biopsy of an area of presumed ischaemic small bowel injury during the FMT procedure; managed conservatively.</p> <p>Deaths: x1 death at three months - directly related to recently diagnosed metastatic pancreatic cancer, not related to FMT .</p>	<p>Selection/ eligibility reported: Yes.</p> <p>Consecutively recruited: Yes, implied that were.</p> <p>Prospectively recruited: No.</p> <p>Loss to follow up explained: Yes.</p> <p>At least 90% followed up: Yes - at least as far as primary outcome.</p>
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Supplementary Material 2 for Gut

	<p>least 2 x toxin positive episodes previously to participate.</p> <p>Pre-FMT antibiotics: All 31 previous methotrexate, all 31 previous vancomycin, 6 previous fidaxomicin, 10 previous rifaximin, 23 prior probiotic.</p>	<p><i>Microsporidia</i> smear, <i>C difficile</i> toxin (PCR or EIA).</p>	<p>Prokinetics: Not described.</p> <p>Time before CDI treatment was stopped before FMT: Antibiotics continued until 4 hours before prep (i.e. stopped day prior to FMT).</p>			
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Supplementary Material 2 for *Gut*

<p>Pathak <i>et al</i>, <i>Clinical &amp; Experimental Gastroenterology</i>, 2013</p>	<p>Case series.</p> <p>Number of patients: 12.</p> <p>Female: male: 8: 4.</p> <p>Age (mean): Mean 71.9 (range 37 – 90) years.</p> <p>Comorbidities: x1 UC, 1 renal transplant, x1 left colon adenocarcinoma and diverticulitis; x1 ruptured appendix; x2 ventilator-dependent.</p> <p>CDI features: Recurrent; full details not given. Two of the patients had had recurrent CDI treated with FMT 'many years ago'.</p> <p>CDI diagnosis confirmation: Not specifically defined.</p> <p>Pre-FMT antibiotics: All vancomycin, 8 patients fidaxomicin, 4 patients methotrexate.</p>	<p>Donors were preferably family/ first degree relatives; family used in all cases here.</p> <p>Working in healthcare: Not specifically addressed.</p> <p>Donor demographics: Not given.</p> <p>Donor screening: Questionnaire - exposure to HIV, hepatitis, STDs; high risk sexual behaviour; drug use, tattoos/ piercings, imprisonment, other high risk behaviour; known current communicable disease; GI morbidities including IBD or GI malignancy; antibiotic use within 90 days.</p> <p>Travel and antibiotic exclusion period: Excluded as donor if antibiotic use within last 90 days.</p> <p>Screening blood tests: HIV-1/-2, hepatitis A/B/C, STDs.</p> <p>Screening stool tests: MC&amp;S, ova, cysts and parasites, <i>C difficile</i> toxin A and B.</p>	<p>Amount of stool per transplant / administered to patients: About 6-8 tablespoons.</p> <p>Diluent used to prepare: 1l of tap water.</p> <p>Diluent used to store if frozen: N/A - all fresh.</p> <p>Preparation methods: No specific details.</p> <p>Time from preparation to transplant (fresh): 6 hours.</p> <p>Time period for storage (frozen): N/A.</p> <p>Route administered: Upper GI: nasoduodenal tube (1; as a second FMT); lower GI: colonoscopy (12).</p> <p>Number of infusions: 1 FMT initially.</p> <p>Bowel purgative: PEG the night before FMT.</p> <p>PPI: Not described.</p> <p>Antimotility: 2 tablets diphenoxylate/ atropine post-FMT.</p> <p>Prokinetics: Not described.</p> <p>Time before CDI treatment was stopped before FMT: 24 hours.</p>	<p>Overall cure within stated follow-up period: 91.7% (<math>n=11/12</math>).</p> <p>Cure with one infusion alone: 91.7% (<math>n=11/12</math>).</p> <p>Total follow up period: 2-26 months.</p>	<p>Minor GI adverse events: Not stated.</p> <p>Minor non-GI adverse events: Not stated.</p> <p>Serious adverse events: Not stated.</p> <p>Deaths: x1 death. Patient with perforated appendix developed rCDI; didn't respond to six months of anti-CDI treatment, went to ITU. Donor was husband - no screening, and no response to colonoscopic FMT. For 2<sup>nd</sup> FMT, used healthy volunteer donor FMT via nasoduodenal tube - responded. Urinary tract infection at nursing home few months later – antibiotic treatment precipitated further CDI. Further sepsis, returned to ITU -</p>	<p>Selection/eligibility reported: Yes.</p> <p>Consecutively recruited: Yes, implied that were.</p> <p>Prospectively recruited: No.</p> <p>Loss to follow up explained: Yes.</p> <p>At least 90% followed up: Yes.</p>
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Supplementary Material 2 for *Gut*

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					declined treatment, then died, four months after initial FMT.	
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Supplementary Material 2 for *Gut*

<p>Rohlke <i>et al</i>, <i>Journal of Clinical Gastroenterology</i>, 2010</p>	<p>Case series.</p> <p>Number of patients: 19. Female: male: 17: 2.</p> <p>Age (mean): Mean age 49 years.</p> <p>Comorbidities: Not described.</p> <p>CDI features: Recurrent CDI.</p> <p>CDI diagnosis confirmation: Positive <i>C difficile</i> toxin and consistently recurring symptoms over a span of six months.</p> <p>Pre-FMT antibiotics: Not given in detail - all at least three courses of conventional anti-CDI antibiotics, including pulsed and tapered vancomycin.</p>	<p>Donors were 4 family members, 14 partners, and 1 housemate.</p> <p>Donors working in healthcare: Excluded.</p> <p>Donor demographics: Donor screening: Questionnaire – included current or recent diarrhoeal illness, sexual behaviour.</p> <p>Travel and antibiotic exclusion period: Excluded if 'recent antibiotic use'; not further defined.</p> <p>Screening blood tests.: HIV, hepatitis A, B and C, and <i>Treponema</i> serology.</p> <p>Screening stool tests: <i>C difficile</i>, bacterial culture, ova, cysts and parasites, <i>Giardia</i>, <i>Cryptosporidium</i>.</p>	<p>Amount of stool per transplant / administered to patients: 350mls.</p> <p>Diluent used to prepare: Normal saline.</p> <p>Diluent used to store if frozen: N/A - fresh.</p> <p>Preparation methods: Fresh preparation, with manual shaking of stool and saline in large suction canister, followed by filtering.</p> <p>Time from preparation to transplant (fresh): Not stated.</p> <p>Time period for storage (frozen): N/A.</p> <p>Route administered: Upper GI: nil; lower GI: all given via colonoscopy.</p> <p>Number of infusions: One routinely, with one patient having a second FMT.</p> <p>Bowel purgative: PEG.</p> <p>PPI: Not described.</p> <p>Antimotility: Loperamide post-FMT.</p> <p>Prokinetics: Not described.</p> <p>Time before CDI treatment was stopped before FMT: 1-3 days.</p>	<p>Overall cure within stated follow up period: 100% (n=20/20).</p> <p>Cure with one infusion alone: 95% (n=19/20).</p> <p>Total follow-up period: 6 months to 5 years.</p>	<p>Minor GI adverse events: Nil reported.</p> <p>Minor non-GI adverse events: Nil reported.</p> <p>Serious adverse events: Nil reported.</p> <p>Deaths: Nil reported.</p>	<p>Selection/ eligibility reported: Yes.</p> <p>Consecutively recruited: Yes.</p> <p>Prospectively recruited: No.</p> <p>Loss to follow up explained: Yes – variable follow-up.</p> <p>At least 90% followed up: Yes.</p>
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Supplementary Material 2 for Gut

Rubin <i>et al</i> , <i>Anaerobe</i> , 2013	<p>Case series.</p> <p>Number of patients: 75.</p> <p>Female: male: 49: 26.</p> <p>Age (median): Median 63 (range 6-94) years.</p> <p>Comorbidities: x10 diabetes mellitus, x8 malignancy, x7 corticosteroids in prior three months.</p> <p>CDI features: Not stated.</p> <p>CDI diagnosis confirmation: Not described.</p> <p>Pre-FMT antibiotics: Oral metronidazole or vancomycin alone or in combination for initial FMT in all cases; not clear exact breakdown/ use for recurrences.</p>	<p>Donors were healthy people from the same household as the patient.</p> <p>Donors working in healthcare: Not stated.</p> <p>Donor demographics: Not described.</p> <p>Donor screening: Questionnaire – as per Aas <i>et al</i>, <i>Clin Infect Dis</i>, 2003.</p> <p>Travel and antibiotic exclusion period: As per Aas <i>et al</i>, <i>Clin Infect Dis</i>, 2003.</p> <p>Screening blood tests: As per Aas <i>et al</i>, <i>Clin Infect Dis</i>, 2003.</p> <p>Screening stool tests: As per Aas <i>et al</i>, <i>Clin Infect Dis</i>, 2003.</p>	<p>Amount of stool per transplant/ administered to patients: 30g of stool.</p> <p>Diluent used to prepare: Saline - As per Aas <i>et al</i>, <i>Clin Infect Dis</i>, 2003. 25ml of stool/ saline mixture per FMT.</p> <p>Diluent used to store if frozen: N/A - fresh.</p> <p>Preparation methods: As per Aas <i>et al</i>, <i>Clin Infect Dis</i>, 2003.</p> <p>Time from preparation to transplant (fresh): As per Aas <i>et al</i>, <i>Clin Infect Dis</i>, 2003.</p> <p>Time period for storage (frozen): N/A – fresh.</p> <p>Route administered: Upper GI: 64 nasogastric, 4 PEG, 7 OGD (75 administrations to 74 patients); lower GI: nil; capsule: nil.</p> <p>Number of infusions: One routinely.</p> <p>Bowel purgative: Not described.</p> <p>PPI: Evening prior to/ morning of procedure - no further details.</p> <p>Antimotility: Not described.</p> <p>Prokinetics: Not described.</p>	<p>Overall cure within stated follow up period: 78.7% (n=59/75).</p> <p>Cure with one infusion alone: 78.7% (n=59/75).</p> <p>Total follow up period: Up to 60 days.</p>	<p>Minor GI adverse events: Nil.</p> <p>Minor non-GI adverse events: Nil.</p> <p>Serious adverse events: Nil.</p> <p>Deaths: No - up to 60 days.</p>	<p>Selection/ eligibility reported: Yes.</p> <p>Consecutively recruited: Yes.</p> <p>Prospectively recruited: No.</p> <p>Loss to follow up explained: Yes.</p> <p>At least 90% followed up: Yes.</p>
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Supplementary Material 2 for *Gut*

Time before CDI treatment was stopped  
before FMT: Stopped on the day prior to  
procedure.

Supplementary Material 2 for Gut

Satokari <i>et al</i> , <i>Alimentary Pharmacology and Therapeutics</i> , 2015	Case series.	Donors were: 15 fresh FMTs with individual donors, 11 fresh FMTs with universal donors; and 23 frozen FMTs with universal donor.	Amount of stool per transplant / administered to patients: Fresh - approximately 30g of stool.	Overall cure within stated follow up period: Fresh: 96% (n=25/26); frozen: 96% (n=22/23).  Total follow up period: 12 weeks.	Minor GI adverse events: N/A.	Selection/ eligibility reported: Yes.  Consecutively recruited: Yes.  Prospectively recruited: No.  Loss to follow up explained: Yes.  At least 90% followed up: Yes.
	Number of patients: 49.		Diluent used to prepare: Fresh - approximately 150ml of tap water.			
	Female: male: 34: 15.		Diluent used to store if frozen: Frozen - 30g of stool added to 150ml N/saline and then glycerol		Minor non-GI adverse events: Mild transient fever in x2 patients with frozen FMT.	
	Age (mean): Fresh: 52 (range 22-81) years; frozen: 61 (range 20-88) years.	Donor working in healthcare: Not stated.	Preparation methods: As described.		Serious adverse events: N/A.	
	Comorbidities: Not described in significant details.	Donor demographics: No clear age or BMI limits.	Time from preparation to transplant (fresh): Fresh - less than 6 hours between delivery and administration; less than 15 minutes between making FMT and delivery.		Deaths: x1 fresh faeces patient died within one year of FMT - not related; x2 frozen patients had relapse within one year, both treated with further antibiotics – x1 died of recurrent CDI, x1 died of arterial thrombosis.	
	CDI features: Recurrent - mean 4.6 (range 2-12) relapses in fresh; mean 4.9 (range 1-6) relapses in frozen.	Donor screening: Questionnaire - "No antibiotics in past six months and no intestinal symptoms".	Time period for storage (frozen): Up to 16 weeks; thawed over 4-5 hours at room temp or in 37°C water bath.			
		Travel and antibiotic exclusion period: Excluded as donors if had used antibiotics in past six months.	Route administered: Upper GI: nil; lower GI: colonoscopy (49); capsules: nil.			
	CDI diagnosis confirmation: "Positive culture and toxin".	Screening bloods: Total blood count, CRP, creatinine, LFTs, hepatitis B and C, HIV-1/-1, <i>Treponema</i> .	Number of infusions: One FMT routinely.			
	Pre-FMT antibiotics: Describes using vancomycin with all, but no specific details.	Screening stools: <i>C difficile</i> culture and toxin A/B test, MC&S, ova, cysts and parasites.	Bowel purgative: 4l Colonsteril PEG/ 2l Moviprep.			
			PPI: Not described.			
			Antimotility: Not described.			

Supplementary Material 2 for *Gut*

Prokinetics: not described.

Time before CDI treatment was stopped  
before FMT: Stopped at an average of 36  
hours prior to administration.

Supplementary Material 2 for Gut

Yoon et al, Journal of Clinical Gastroenterology, 2010	Case series.		Amount of stool per transplant / administered to patients: Stool (unclear how much) mixed with 1l normal saline; approx 250-450cc of FMT administered in total.			
	Number of patients: 12.					
	Female: male: 9: 3.		Diluent used to prepare: Normal saline.			
	Age (mean)*: Mean 66 (range 30 - 86) years.	Donors were spouses/ partners in 8 patients; for other 4 patients, donors were one son, two daughters, and one granddaughter.	Diluent used to store if frozen: N/A.			
	Comorbidities: 9 with diverticulosis (with 2 of these having diverticulitis as index infection).	Donors working in healthcare: No.	Preparation methods: Manually shaken then filtered through gauze.		Minor GI adverse events: Nil described.	Selection/ eligibility reported: Yes.
	CDI features: 1 patient with first CDI, 2 with 2nd, 5 with 3rd, 1 with 4th, 1 with 5th, 1 with 6th, 1 with 8 <sup>th</sup> .	Donor demographics: No details.	Time from preparation to transplant (fresh): No details.	Overall cure within stated follow up period: 100% (n=12/12).	Minor non-GI adverse events: Nil described.	Consecutively recruited: Yes.
	CDI diagnosis confirmation: Toxin testing for either toxin A or B, or assessment of both via EIA.	Donor screening: Questionnaire - no details.	Time period for storage (frozen): N/A.	Total follow up period: 3 weeks to 8 years - no details on relation to individual patients.		Prospectively recruited: No.
		Travel and antibiotic exclusion period: No details given	Route administered: Upper GI: (N/A)		Serious adverse events: Nil described.	Loss to follow up explained: No.
		Screening bloods: Hepatitis B and C, HIV.	Lower GI: 10-20cc of FMT administered every 5-10cm of withdrawal distance in all 12 patients.		Deaths: Nil described.	At least 90% followed up: Yes.
	Pre-FMT antibiotics: 12 had oral metronidazole, 3 had intravenous metronidazole, 12 had oral vancomycin, 4 x rifaximin, no mention of fidaxomicin.	Screening stools: <i>C difficile</i> toxin, enteric pathogens, ova, cysts and parasites - at treating clinician's discretion.	Number of infusions: Single.			
			Bowel purgative: All colonoscopic, but no specific details given.			
			PPI: Not described.			
			Antimotility: Not described.			
			Prokinetics: Not described.			

Supplementary Material 2 for *Gut*

Time CDI treatment was stopped before  
FMT: 3 days.



Supplementary Material 2 for Gut

Youngster <i>et al</i> , JAMA, 2014	Prospective case series.	Donors were unrelated adult volunteers.	Amount of stool per transplant / administered to patients: 30 capsules (single treatment) - total 48g of stool.		Minor GI adverse events: Transient abdominal cramping and bloating in 6 patients (30%) that resolved in 72 hours.	
	Number of patients: 20.		Diluent used to prepare: saline in 1/10th volume of stool.			
	Female: male: 9: 11.	Donor working in healthcare: Not stated.	Diluent used to store if frozen: 10% glycerol.			
	Age (median): Median 64.5 (range 11-89) years.	Donor demographics: Age range 18-50 years, BMI 18.5 - 25.	Preparation methods: Faecal matter solution was pipetted into size 0 capsules (650 µL), which were closed and then secondarily sealed in size 00 capsules. Capsules were stored frozen at -80°C until use.	Overall cure within stated follow up period: 90% (n=18/20).	Minor non-GI adverse events: Not described.	Selection/ eligibility reported: Yes.
	Comorbidities: Specific comorbidities not described.	Donor screening: Questionnaire - American Association of Blood Banks donor questionnaire.	Time from preparation to transplant (fresh): N/A.	Cure with one infusion alone: 70% (n=14/20).	Serious adverse events: x1 hospitalised with a documented relapse of severe CDI after taking 15 capsules, but had successful treatment after receiving the remaining 15 capsules. No other severe adverse events (grade 2 or above).	Consecutively recruited: Yes.
	CDI features: Included patients with both recurrent or refractory CDI.	Travel and antibiotic exclusion period: Excluded as potential donors if used antibiotics within preceeding 6 months.	Time period for storage (frozen): Mean 113 days (30-252 days).	Total follow up period: 8 weeks.		Prospectively recruited: Yes.
	CDI diagnosis confirmation: Toxin and ELISA, PCR if toxin negative but ELISA is positive or indeterminate.	Screening blood tests: Antibodies to hepatitis A, B, and C; HIV; and <i>Treponema pallidum</i> within 2 weeks of donations.	Route administered: All courses were 30 oral capsules.			Loss to follow up explained: Yes.
	Pre-FMT antibiotics: Failed vancomycin taper and/ or fidaxomicin.	Screening stool tests: " Enteric pathogens".	Number of treatments: 1 course (given as 15 capsules on 2 consecutive days). If failed, retreated at a mean of 7 days.			At least 90% followed up: Yes.
			Bowel purgative: Not described.		Deaths: none.	
			PPI: Not described.			
			Antimotility: Not described.			

Supplementary Material 2 for *Gut*

Prokinetics: Not described.

Time before CDI treatment was stopped  
before FMT: 48 hours prior to FMT.

Supplementary Material 2 for Gut

Youngster <i>et al</i> , <i>BMC Medicine</i> , 2016	Case series.	Donors were healthy volunteers.	Amount of stool per transplant / administered to patients: 30 capsules derived from a mean of 48g of faeces.	Overall cure within stated follow up period: 91% (n=164/180)  Cure with one infusion alone: 82% (n=147/180)  Total follow up period: 8 weeks for primary response.	Minor GI adverse events: x5 vomiting, x112 diarrhoea, x45 nausea/ bloating, x40 abdominal pain.  Minor non-GI adverse events: x3 fever, x54 fatigue, malaise, and headache, x12 other complaints.  Serious adverse events: Related serious (x1 fever, x2 new UC, x6 hospitalisations for CDI/ diarrhoea).  Unrelated serious adverse events: x26 hospitalisations, x14 deaths.  Deaths: x14 (unrelated).	Selection/eligibility reported: Yes.  Consecutively recruited: Yes.  Prospectively recruited: No.  Loss to follow up explained: Yes.  At least 90% followed up: Yes.
	Number of patients: 180.	Donors working in healthcare: Not mentioned.	Diluent used to prepare: Normal saline.			
	Female: male: Not stated.	Donor demographics: 18-50 years of age, on no medications, with a 'normal body mass index'.	Diluent used to store if frozen: 10% glycerol.			
	Age (median): Median 64 (range 7–95) years.	Donor screening: Questionnaire - initial screening using the American Association of Blood Banks donor questionnaire for exposure to infectious agents.	Preparation methods: Homogenised using a commercial blender then passed through sieves in ambient air.			
	Comorbidities: Not described.	Travel and antibiotic exclusion period: Excluded as donor if antibiotic use within 6 months.	Time from preparation to transplant (fresh): N/A.			
	CDI features: Three or more mild-to-moderate episodes of CDI or two episodes requiring hospitalisation.	Screening bloods: Blood was screened for antibodies to hepatitis A, B, and C; HIV; and <i>Treponema pallidum</i> within 2 weeks of donations.	Time period for storage (frozen): Study of capsulised FMT. Faecal slurry was double-encapsulated in hypromellose capsules (Capsugel, Cambridge, MA) and stored at –80 °C for up to 6 months pending use.			
	CDI diagnosis confirmation: Not specifically described.	Screening stools: Donor faeces were screened for enteric bacterial pathogens including rotavirus, <i>Listeria monocytogenes</i> , <i>Vibrio cholerae</i> , <i>Escherichia coli</i> O157, ova and parasites (including general microscopy, acid-fast staining, and/or antigen testing	Route administered: All received 30 capsules as a 'dose'.			
	Pre-FMT antibiotics: Not described.		Number of infusions: 1 course of capsules in 147 patients, 2 courses in 26 patients and 3 course in 4 patients.			
			Bowel purgative: not mentioned.			
			PPI: not mentioned.			
			Antimotility: not mentioned.			

Supplementary Material 2 for *Gut*

		for <i>Giardia</i> , <i>Cryptosporidium</i> , <i>Isospora</i> , and <i>Microsporidia</i> ), <i>C. difficile</i> , and <i>Helicobacter pylori</i> antigen.	Prokinetics: not mentioned.  Time before CDI treatment was stopped before FMT: 24–48 hours prior.			
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Supplementary Material 2 for Gut

Zainah et al, Digestive Diseases and Sciences, 2014	Case series.		Amount of stool per transplant / administered to patients: 30-50g.			
	Number of patients: 14.					
	Female: male: 9:5.		Diluent used to prepare: Warm tap water.			
	Age (mean +/-range)*: 73.4 (+/-11.9) years.	Donors: 12 patients received FMT from related donor (7 spouse, 5 children); the other two used unrelated donors.	Diluent used to store if frozen: N/A.			
	Comorbidities: x4 patients with cancer, x1 OLT patient.	Donors working in healthcare: Not stated.	Preparation methods: Homogenised mixture, then filtered through gauze; 120-180ml of suspension if through nasogastric tube, 300-500ml if through colonoscopy.	Overall cure within stated follow up period: 79% (n=11/14) by seven days.	Minor GI adverse events: Not described.	Selection/ eligibility reported: Yes.
	CDI features: 8 patients had had prev CDI episodes (2-5 episodes prior).	Donor demographics: Not stated.	Time from preparation to transplant (fresh): "Same day".		Minor non-GI adverse events: Not described.	Consecutively recruited: Yes.
	CDI diagnosis: Diarrhoea (at least 3 unformed stool/d for 2 consecutive days) + positive <i>C difficile</i> EIA and/or PCR. All patients here severe by definition - defined here as age >60 years, albumin <2.5mg/dl, temp at least 38.3°C, WBC > 15 within 48 hour of CDI diagnosis; or at least one of the following: pseudomembranes, treatment in intensive care.	Donor screening: Questionnaire - not described.	Time period for storage (frozen): N/A.	Cure with one infusion alone: 71% (n=10/14).	Serious adverse events: Not described.	Prospectively recruited: No.
		Travel and antibiotic exclusion period: No details.	Route administered: Upper GI: Nasogastric administration in all but one patient (13 patients); lower GI: colonoscopic administration in one patient (1 patient).	Total follow up period: Up to 100 days .	Deaths: x1 within 7 days of FMT - but died of their malignancy.	Loss to follow up explained: Yes.
		Screening blood tests: HIV-1/-2, hepatitis A IgM, hepatitis B serology, hepatitis C antibody, syphilis (RPR and FTA-Abs).	Number of infusions: One routinely; repeated if no response at 48-72hr.			At least 90% followed up: Yes.
		Screening stools: <i>C difficile</i> toxin by PCR, stool ova, cysts and parasites.	Bowel purgative: No details.			
			PPI: Yes, pre nasogastric administration - no details given.			
			Antimotility: Not described.			

Supplementary Material 2 for *Gut*

	Pre-FMT antibiotics: 14 patients prior vancomycin, 12 prior metronidazole too.		Prokinetics: Not described.  Time before CDI treatment was stopped before FMT: 24 hours.			
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3                   **C.2.     Reviewed randomised studies of FMT for recurrent or refractory CDI**  
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Paper	Study and patient characteristics	Donor characteristics	FMT characteristics	Outcomes	Adverse events
Camacho-Ortiz <i>et al</i> , <i>PLoS ONE</i> , 2017	<p>Intervention: FMT (pooled from three donors).  Number of patients: 9.  Female: male: 3: 4 (data only presented for 7 patients).  Age: Mean of 39.7 (+/- 24.8) years.</p> <p>Comparator: Vancomycin (250mg every 6 hours for 10-14 days).  Number of patients: 10.  Female: Male: 3: 6 (data only presented for 9 patients).  Age (mean/median): Mean of 46.7 (+/- 15.8) years.</p> <p>Comorbidities: In FMT arm – x1 abdominal abscess, x1 Child B cirrhotic, x1 pulmonary TB; in vancomycin arm – x2 haemodialysis patients, x1 meningeal TB, x1 'abscessed squamous cell carcinoma'.</p> <p>CDI features: All first episode of CDI, occurring at least 48hrs after admission.</p> <p>CDI diagnosis confirmation: &gt;3 bowel movements during the previous 24 hours, Bristol scale &gt; 5, positive <i>C. difficile</i> EIA or PCR.</p> <p>Pre-FMT antibiotics: no antibiotics within FMT arm; patients in vancomycin arm received 250mg</p>	<p>Donors working in healthcare: Not stated.</p> <p>Donor demographics: &gt;18 years, non-pregnant, BMI 20-25kg/m<sup>2</sup></p> <p>Donor screening: On questionnaire, rejected potential donors who in the past three months had had use of PPI, use of immunosuppressives, hospitalisation and/ or diarrhoea. Also excluded if high risk sexual behaviour, first degree relative with diabetes mellitus, abdominal surgery, and any GI disease/ cancer.</p> <p>Travel and antibiotic exclusion period: Excluded if antibiotics within the past 3 months.</p> <p>Screening blood tests: Normal full blood count and liver enzymes essential for inclusion. Also screened for HAV, HBV, HCV, HIV, CMV, EBV, <i>Trypanosoma</i>, <i>Brucella</i>, <i>Treponema pallidum</i>.</p> <p>Screening stool tests: Included parasites, enteropathogenic bacteria, rotavirus.</p>	<p>Amount of stool per transplant: 45ml of pooled donor stool (from three donors), at ~0.19g/ml.</p> <p>Diluent used to prepare: 0.9% saline.</p> <p>Diluent used to store if frozen: 15% v/v glycerol.</p> <p>Preparation methods: Stool from donors pooled, mixed, resuspended in saline, filtered to remove particles &gt; 330µm .</p> <p>Time from preparation to transplant (fresh): N/A.</p> <p>Time period for storage (frozen): Not stated.</p> <p>Route administered: Upper GI: 14 by OGD; 1 by nasojejunal tube. Lower GI: colonic (1; patient with anatomical abnormality due to head and neck neoplasia). Capsule: nil.</p> <p>Number of infusions: routinely 1; patients not resolving after first FMT received 2<sup>nd</sup> FMT (as did patients not improving with vancomycin).</p> <p>Bowel purgative: Not stated.</p>	<p>Treatment arm: FMT  Overall cure rate: 71.4% (n=5/7) (after 2 x FMT)  Cure with one infusion alone: 57.1% (n=4/7).</p> <p>Treatment arm: Vancomycin  Overall cure rate: 88.9% (n=8/9) (not clear if failed patient received FMT subsequently, as is described in protocol).</p>	<p>Minor GI adverse events: Nil stated.</p> <p>Minor non-GI adverse events: Nil stated.</p> <p>Serious adverse events: Nil stated.</p> <p>Deaths: Nil.</p>



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	<p>every 6hrs for 10-14 days.</p> <p>Total follow up period: up to one year.</p> <p>Cochrane Collaboration risk of bias assessment: uncertain risk of bias.</p>		<p>PPI: Not stated.</p> <p>Antimotility: Not stated.</p> <p>Prokinetics: Not stated.</p> <p>Time before CDI treatment was stopped before FMT: Nil given.</p>		
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<p>Cammarota <i>et al</i>, <i>Alimentary Pharmacology and Therapeutics</i>, 2015</p>	<p>Intervention: FMT. Number of patients: 20. Female: Male: 12: 8. Age (mean/median): Mean 71 (range 29-89) years.</p> <p>Comparator: Vancomycin (125mg four times daily for 10 days, follow by a pulse regimen (125-500mg/day every 2-3 days, for at least three weeks). Number of patients: 19. Female: Male: 11: 8. Age (mean/median): Mean 75 (range 49-93) years.</p> <p>Comorbidities: No significant difference of Charlson comorbidity index between groups.</p> <p>CDI features: All recurrent. 7/20 in FMT arm with pseudomembranous colitis.</p> <p>CDI diagnosis confirmation: Diarrhoea and CDT positive within 10 weeks of previous antibiotic treatment.</p> <p>Pre-FMT antibiotics: All had had vancomycin or metronidazole. 19/20 of FMT arm and 16/20 of vancomycin arm had had previous vancomycin taper.</p> <p>Total follow up period: 10 weeks.</p>	<p>Donors working in healthcare: no.</p> <p>Donor demographics: Less than 50 years of age, no antibiotics within past 6 months.</p> <p>Donor screening: Questionnaire - no antibiotics for last 6/12. Excluded if significant GI disease, metabolic syndrome, chronic illness, immunocompromise, recent travel, high risk lifestyle in last three months.</p> <p>Travel and antibiotic exclusion period: 3 month travel exclusion period, 6 month antibiotic exclusion period.</p> <p>Screening blood tests: Hepatitis A, B, and C, HIV, EBV, syphilis, <i>Stongyloides</i>, <i>Entamoeba histolytica</i>, FBC, LFTs, creatinine, CRP.</p> <p>Screening stool tests: <i>C. difficile</i> cult and toxin, enteric bacteria, ova, cysts and parasites, VRE, MRSA, Gram negative multi-drug resistant bacteria.</p>	<p>Amount of stool per transplant / administered to patients: Not specified.</p> <p>Diluent used to prepare: Normal saline 500mls.</p> <p>Diluent used to store if frozen: N/A – fresh.</p> <p>Preparation methods: Blended and strained.</p> <p>Time from preparation to transplant (fresh): 6 hours.</p> <p>Time period for storage (frozen): N/A.</p> <p>Route administered: Upper GI: nil; lower GI: colonic (20); capsule: nil.</p> <p>Number of infusions: 14 had 1 infusion, 4 had 2 infusions, 1 had 3 infusions and 1 had 4 infusions. Initial protocol was that if non-response to first FMT, then second FMT after one week; however, after first two patients, changed to all patients with pseudomembranous colitis receiving repeat FMT every 3 days until resolution of CDI.</p> <p>Bowel purgative: Macrogol.</p> <p>PPI: No.</p>	<p>Treatment arm: FMT Overall cure rate: 90% (<math>n=18/20</math>). Cure with one infusion alone: 65% (<math>n=13/20</math>); none of these were patients with pseudomembranous colitis. The 7 patients not cured with first FMT all had pseudomembranous colitis; of these, 5/7 cured with protocol of recurrent FMTs.</p> <p>Treatment arm: Vancomycin: Overall cure rate: Cure with one infusion alone: 26% (<math>n=5/19</math>).</p>	<p>Minor GI adverse events: x19 diarrhoea, x12 bloating (all resolved at 12 hours).</p> <p>Minor non-GI adverse events: None.</p> <p>Serious adverse events: None.</p> <p>Deaths: x2 from <i>C difficile</i>-related complications.</p>
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	Cochrane Collaboration risk of bias assessment: uncertain risk of bias.		Antimotility: No.  Prokinetics: No.  Time before CDI treatment was stopped before FMT: Between five and two days prior to FMT.		
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<p>Allegretti <i>et al</i>, <i>Gastroenterology</i> (abstract), 2016</p>	<p>Intervention: Low dose FMT capsules (30 pills once). Number of patients: 10. Female: male: Not stated. Age (mean/median): Not stated.</p> <p>Comparator: High dose FMT. capsules (30 pills daily on two consecutive days). Number of patients: 9. Female: male: Not stated. Age (mean/median): Not stated.</p> <p>Comorbidities: Not stated.</p> <p>CDI features: Not stated.</p> <p>CDI diagnosis confirmation: Not stated.</p> <p>Pre-FMT antibiotics: Not stated.</p> <p>Total follow up period: 8 weeks.</p> <p>Cochrane Collaboration risk of bias assessment: uncertain risk of bias.</p>	<p>Donors were unrelated donors from universal stool bank (OpenBiome).</p> <p>Donors working in healthcare: No.</p> <p>Donor demographics: mean age 26, mean BMI 22.2.</p> <p>Donor screening: Questionnaire - as per OpenBiome protocol.</p> <p>Travel and antibiotic exclusion period: As per OpenBiome protocol.</p> <p>Screening bloods: As per OpenBiome protocol.</p> <p>Screening stools: As per OpenBiome protocol.</p>	<p>Amount of stool per transplant / administered to patients: 30 pills a day for one day.</p> <p>Diluent used to prepare: Not stated.</p> <p>Diluent used to store if frozen: Stored at -80°C prior to use.</p> <p>Preparation methods: Capsules physically stable for 30 days at 25°C using an emulsion-based production protocol.</p> <p>Time from preparation to transplant (fresh): Not stated.</p> <p>Time period for storage (frozen): Not stated.</p> <p>Route administered: All capsule – as described above.</p> <p>Number of infusions: 30 tablets (over one day).</p> <p>Bowel purgative: Not stated.</p> <p>PPI: Not stated.</p> <p>Antimotility: Not stated.</p> <p>Prokinetics: Not stated.</p> <p>Time before CDI treatment was stopped before FMT: Not stated.</p>	<p>Treatment arm: Low dose FMT capsules (30 pills once). Overall cure rate: 70% (n=7/10).</p> <p>Treatment arm: High dose FMT capsules (30 pills daily on two consecutive days). Overall cure rate: 77.8% (n=7/9).</p>	<p>Minor GI adverse events: None.</p> <p>Minor non-GI adverse events: None.</p> <p>Serious adverse events: None.</p> <p>Deaths: None.</p>
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Hota <i>et al</i> , <i>Clinical Infectious Diseases</i> , 2016	Intervention: FMT. Number of patients: 16. Female: male: 11: 5. Age (mean/ standard deviation): Mean 75.7 +/- 14.5 years.		Amount of stool per transplant / administered to patients: 50g.		
	Comparator: 6 week vancomycin taper. Number of patients: 12. Female: male: 8: 4. Age (mean/ standard deviation): Mean 69.6 +/- 14.2 years.	Donors working in healthcare: Not stated.	Diluent used to prepare: 500mls normal saline.		
	Comorbidities: Not stated, but similar Charlson comorbidity index score between groups.	Donor demographics: ≥18yrs.	Diluent used to store if frozen: N/A – fresh.		Minor GI adverse events: abdominal pain, tenderness and bloating, equal in both groups.
	CDI features: All recurrent.	Donor screening: Questionnaire - self-screening questionnaire of behaviours associated with risk for blood-borne pathogens.	Preparation methods: Stomacher laboratory blender.	Treatment arm: FMT: Overall cure rate: 43.8% (n=7/16). Cure with one infusion alone: 43.8% (n=7/16).	Minor non-GI adverse events: Nil.
	CDI diagnosis confirmation: Symptoms and toxin or PCR detection.	Travel and antibiotic exclusion period: Antibiotic use for at least two days in the preceding three months.	Time from preparation to transplant (fresh): 48 hours.		Serious adverse events: x1 developed anasarca from liver disease, x1 had perforated bowel from diverticulitis at 35 days post-FMT.
	Pre-FMT antibiotics: At least 1 course of vancomycin for a minimum of 10 days. The majority of patients in both arms had had prior vancomycin tapers.	Screening blood tests: Extensive screening comparable with previous studies.	Time period for storage (frozen): N/A.	Treatment arm: 6 week vancomycin taper. Overall cure rate: 58.3% (n=7/12).	Deaths: None.
	Total follow up period: 120 days.	Screening stool tests: Extensive screening comparable with previous studies.	Route administered: Upper GI: nil; lower GI: 16; capsule: nil.		
	Cochrane Collaboration risk of bias assessment: uncertain risk of bias.		Number of infusions: All had 1 infusion.		
			Bowel purgative: None.		
			PPI: None.		
			Antimotility: None.		
			Prokinetics: None.		
			Time before CDI treatment was stopped before FMT: Day prior to FMT.		

<p>Jiang <i>et al</i>, <i>Alimentary Pharmacology and Therapeutics</i>, 2017</p>	<p>Intervention: Fresh FMT. Number of patients: 25. Female: male: 21:4. Age (mean): Mean 75 (range 19-97) years.</p> <p>Comparator: Lyophilised FMT. Number of patients: 23. Female: Male: 13: 10. Age (mean): Mean 63 (range 20-87) years.</p> <p>Comparator: Frozen FMT. Number of patients: 24 Female: Male: 18: 6. Age (mean): Mean 62.5 (range 33-88) years.</p> <p>CDI features: All recurrent.</p> <p>CDI diagnosis confirmation: Not explicitly stated, but includes CDI toxin.</p> <p>Pre-FMT antibiotics: Not stated.</p> <p>Total follow up period: 2 months.</p> <p>Cochrane Collaboration risk of bias assessment: high risk of bias.</p>	<p>Donors working in healthcare: Not stated.</p> <p>Donor demographics: "Normal BMI".</p> <p>Donor screening: Questionnaire - as per van Nood <i>et al</i>, <i>NEJM</i>, 2013.</p> <p>Travel and antibiotic exclusion period: As per van Nood <i>et al</i>, <i>NEJM</i>, 2013.</p> <p>Screening blood tests: As per van Nood <i>et al</i>, <i>NEJM</i>, 2013.</p> <p>Screening stool tests: As per van Nood <i>et al</i>, <i>NEJM</i>, 2013.</p>	<p>Amount of stool per transplant / administered to patients: 50g.</p> <p>Diluent used to prepare: Normal saline.</p> <p>Diluent used to store if frozen: Implied use of glycerol for frozen product but not clearly stated.</p> <p>Preparation methods: mix stool with normal saline (1:10), aerobic conditions, use Stomacher to homogenise.</p> <p>Time from preparation to transplant (fresh): Within 2 hours of preparation.</p> <p>Time period for storage (frozen): Not specified.</p> <p>Route administered: All colonoscopic.</p> <p>Number of infusions: 1</p> <p>Bowel purgative: PEG on night before FMT.</p> <p>PPI: No.</p> <p>Antimotility: 4mg loperamide 3 hours before.</p> <p>Prokinetics: No.</p>	<p>Treatment arm: Fresh: Overall cure rate: 100% (<math>n=25/25</math>).</p> <p>Cure with one infusion alone: 100% (<math>n=25/25</math>).</p> <p>Treatment arm: Frozen: Overall cure rate: 83% (<math>n=20/24</math>).</p> <p>Cure with one infusion alone: 83% (<math>n=20/24</math>).</p> <p>Treatment arm: Lyophilised: Overall cure rate: 78% (<math>n=20/23</math>).</p> <p>Cure with one infusion alone: 78% (<math>n=20/23</math>).</p>	<p>Minor GI adverse events: no differences in the three groups. Mild transient abdominal pain and diarrhoea in 86% of patients. x6 experienced fatigue and x4 had a headache. x2 gained weight.</p> <p>Minor non-GI adverse events: None stated.</p> <p>Serious adverse events: None.</p> <p>Deaths: None.</p>
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			Time before CDI treatment was stopped before FMT: Not specified.		
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<p>Kao <i>et al</i>, JAMA, 2017</p>	<p>Comparator: Oral FMT capsules. Number of patients: 57. Female: male: 43: 14. Age (median/standard deviation): 58.7 (+/-18.5) years.</p> <p>Comparator: Colonoscopic FMT. Number of patients: 59. Female: male: 36: 13. Age (median/standard deviation): 57.4 (+/-19.1) years.</p> <p>CDI features: All recurrent.</p> <p>CDI diagnosis: Recurrence of diarrhea (&gt;3 unformed bowel movements every 24 hours) within 8 weeks of completing a prior course of treatment, with either a positive <i>C difficile</i> toxin by glutamate dehydrogenase and <i>C difficile</i> toxins A/B (<i>C diff</i> QuikChek Complete; Techlab) or by detection of glutamate dehydrogenase and <i>C difficile</i> cytotoxin B gene (Cepheid), plus resolution of diarrhea for the current episode.</p> <p>Pre-FMT antibiotics: Oral vancomycin (125mg twice daily) up to 24hrs before FMT.</p> <p>Total follow-up period: 12 weeks.</p>	<p>Donors were unrelated volunteers.</p> <p>Working in healthcare: Not stated.</p> <p>Donor demographics: Not stated. Donor screening: Questionnaire: As per Kelly <i>et al</i>, <i>Gastroenterology</i>, 2015.</p> <p>Travel and antibiotic exclusion period: As per Kelly <i>et al</i>, <i>Gastroenterology</i>, 2015.</p> <p>Screening blood tests: As per Kelly <i>et al</i>, <i>Gastroenterology</i>, 2015.</p> <p>Screening stool tests: As per Kelly <i>et al</i>, <i>Gastroenterology</i>, 2015.</p>	<p>Amount of stool per transplant / administered to patients: 80-100g.</p> <p>Diluent used to prepare: Normal saline.</p> <p>Diluent used to store if frozen: 100% glycerol.</p> <p>Preparation methods: Mix stool with 200ml of normal saline, and filtered using a Stomacher to homogenise 180ml of faecal slurry.</p> <p>Time from preparation to transplant (fresh): up to 2 months frozen, collected fresh within 12 hours.</p> <p>Time period for storage (frozen): up to 2 months.</p> <p>Route administered: lower GI: 59 (colonoscopy); capsule: 57.</p> <p>Number of infusions: x1 of colonoscopy, or x40 capsules as one-off.</p> <p>Bowel purgative: PEG on the night before.</p> <p>PPI: No.</p> <p>Antimotility: Not stated.</p> <p>Prokinetics: Not stated.</p>	<p>Treatment arm: Oral FMT capsules: 96.2% (n=51/53) absence of CDI at 12 weeks.</p> <p>Cure with one treatment alone: 96.2% (n=51/53).</p> <p>Treatment arm: FMT via colonoscopy: 96.2% (n=50/52).</p> <p>Cure with one infusion alone: 96.2% (n=50/52).</p>	<p>Minor GI adverse events: Capsule group: x3 nausea, x2 vomiting, x1 abdominal pain. Colonoscopy group: x1 nausea, x1 vomiting, x1 fever, x5 abdominal pain.</p> <p>Minor non-GI adverse events: .1 developed confusion in the colonoscopy group between time of screening and delivery of FMT. This was not communicated to team, and despite an uneventful FMT she died three days later from heart failure.</p> <p>Serious adverse events: None.</p> <p>Deaths: x1 in each group from cardiopulmonary disease (see above for colonoscopy). The other patient developed <i>Staphylococcus epidermis</i> bacteraemia 10 weeks after capsule</p>
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			Time before CDI treatment was stopped before FMT: 24 hours.		treatment and died from sepsis.
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<p>Kelly <i>et al</i>, <i>Annals of Internal Medicine</i>, 2016</p>	<p>Intervention: Donor FMT. Number of patients: 22. Female: male: 18: 4. Age (mean/ standard deviation): Mean age 48 (+/-16) years.</p> <p>Comparator: Autologous FMT. Number of patients: 24. Female: male: 19: 5. Age (mean/ standard deviation): Mean age 55 (+/-14) years.</p> <p>Comorbidities: Similar median Charlson comorbidity scores between groups.</p> <p>CDI features: Recurrent.</p> <p>CDI diagnosis confirmation: <math>\geq 3</math> unformed stools over 24 hours for 2 consecutive days, and either a positive stool test result for <i>C difficile</i> or pseudomembranes on colonoscopy.</p> <p>Pre-FMT antibiotics: All patients had had prolonged prior courses of vancomycin.</p> <p>Total follow up period: 8 week outcome follow up, 6 month safety follow-up.</p> <p>Cochrane Collaboration risk of bias assessment: low risk of bias.</p>	<p>Donors working in healthcare: Not stated.</p> <p>Donor demographics: Not stated.</p> <p>Donor screening: Questionnaire - potential donors also completed a modified AABB full-length donor history questionnaire, and those with risk factors for infectious agents were excluded.</p> <p>Travel and antibiotic exclusion period: Excluded as donor if antibiotics within preceeding 90 days.</p> <p>Screening bloods: Testing for HIV- 1 and HIV-2 was performed within 2 weeks before donation for FMT. Other serologic testing was performed within 1 month before FMT and included testing for hepatitis A, B, and C viruses; also, testing for <i>Treponema pallidum</i>.</p> <p>Screening stool tests: polymerase chain reaction (PCR) testing for detection of <i>C difficile</i> toxin; culture for enteric pathogens (<i>Escherichia coli</i>, <i>Salmonella</i>, <i>Shigella</i>, <i>Yersinia</i>, <i>Campylobac- ter</i>, <i>Listeria monocytogenes</i>, <i>Vibrio parahaemolyticus</i>, and <i>V cholerae</i>); testing for fecal <i>Giardia</i> and <i>Cryptosporidium</i> antigens; acid-fast stain for detection of <i>Cyclospora</i> and</p>	<p>Amount of stool per transplant / administered to patients: Mean stool dose of 64 g (standard deviation of 25 g; range, 20 to 100g).</p> <p>Diluent used to prepare: 100g of stool in 500mls of normal saline.</p> <p>Diluent used to store if frozen: N/A.</p> <p>Preparation methods: Not reported.</p> <p>Time from preparation to transplant (fresh): 6 hours.</p> <p>Time period for storage (frozen): N/A.</p> <p>Route administered: Upper GI: nil; lower GI: all patients in both groups (colonoscopy); capsule: nil.</p> <p>Number of infusions: 1 infusion only.</p> <p>Bowel purgative: polyethylene glycol (PEG).</p> <p>PPI: No.</p> <p>Antimotility: Not described.</p> <p>Prokinetics: No.</p> <p>Time before CDI treatment was stopped before FMT: continued</p>	<p>Treatment arm: Donor FMT: Overall cure rate: 90.9% (<math>n=20/22</math>). Cure with one infusion alone: 90.9% (<math>n=20/22</math>).</p> <p>Treatment arm: Autologous FMT Overall cure rate: 62.5% (<math>n=15/24</math>). Cure with one infusion alone: 62.5% (<math>n=15/24</math>).</p>	<p>Minor GI adverse events: Low rates of abdominal pain, bloating, nausea, vomiting, diarrhea, flatulence, anorexia, and constipation; these did not differ significantly between groups.</p> <p>Minor non-GI adverse events: None described.</p> <p>Serious adverse events: None described.</p> <p>Deaths: None.</p>
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		<i>Isospora</i> ; ova and parasite testing; and enzyme immunoassay for detection of Rotavirus.	therapy until 2 to 3 days before the procedure.		
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<p>Lee <i>et al</i>, JAMA, 2016</p>	<p>Intervention: Frozen FMT. Number of patients: 108. Female: male: 72: 36. Age (mean/ standard deviation): Mean age 73.0 (+/- 16.4) years.</p> <p>Comparator: Fresh FMT. Number of patients: 111. Female: Male: 74: 37. Age (mean/ standard deviation): Mean age 72.5 (+/- 16.2) years.</p> <p>Comorbidities: Not described.</p> <p>CDI features: All recurrent disease.</p> <p>CDI diagnosis confirmation: Toxin and PCR.</p> <p>Pre-FMT antibiotics: All had had prior metronidazole, vancomycin, or both in combination. Almost all patients had had prior vancomycin taper.</p> <p>Total follow up period: 13 weeks.</p> <p>Cochrane Collaboration risk of bias assessment: low risk of bias.</p>	<p>Donors were unrelated volunteers.</p> <p>Donors working in healthcare: Not specifically described.</p> <p>Donor demographics: Not defined.</p> <p>Donor screening: questionnaire – comparable to blood donor screening questionnaire.</p> <p>Travel and antibiotic exclusion period: Excluded as donor if travel (within the last 6 months) to areas of the world where diarrheal illnesses are endemic or risk of traveler's diarrhea is high; also excluded if antibiotics within the preceding 3 months.</p> <p>Screening blood tests: HIV-1 and -2, hepatitis A IgM, HBsAg, anti-HBc (both IgG and IgM), and anti-HBs, hepatitis C antibody, RPR and FTA-ABS.</p> <p>Screening stool tests: <i>Clostridium difficile</i> toxin B by PCR; if unavailable, then evaluation for toxins A and B by EIA; routine bacterial culture for enteric pathogens; faecal <i>Giardia</i> antigen; faecal <i>Cryptosporidium</i> antigen; Acid-fast stain for <i>Cyclospora</i>,</p>	<p>Amount of stool per transplant / administered to patients: 100g of stool.</p> <p>Diluent used to prepare: 300mls of water.</p> <p>Diluent used to store if frozen: no solvents used for storage.</p> <p>Preparation methods: 100g of stool homogenised and mixed in 300mls of water.</p> <p>Time from preparation to transplant (fresh): If fresh, administered within 24hrs.</p> <p>Time period for storage (frozen): If frozen, kept for 30 days at -20°C.</p> <p>Route administered: Upper GI: nil; lower GI: enema FMT for all patients in both groups; capsule: nil.</p> <p>Number of infusions in frozen arm: 57 patients had 1 infusion; 24 patients had 2 infusions; rest had &gt;2 infusions; in fresh arm: 56 patients had 1 infusion; 22 patients had 2 infusion; rest had &gt;2 infusions.</p> <p>Bowel purgative: Not described.</p> <p>PPI: Nil.</p>	<p>Treatment arm: Frozen: Overall cure rate: 90.7% (n=98/109). Cure with one infusion alone: 52.8% (n=57/108).</p> <p>Treatment arm: Fresh: Overall cure rate: 85.6% (n=95/111). Cure with one infusion alone: 50.5% (n=56/111).</p>	<p>Minor GI adverse events: Transient diarrhoea (70%), abdominal cramps (10%), nausea (5%) in 24 hours post-FMT; constipation (20%) and flatulence (25%) in follow-up period. No difference between the two groups.</p> <p>Minor non-GI adverse events: None described.</p> <p>Serious adverse events: x12 patients required hospitalization because of illnesses unrelated to FMT.</p> <p>Deaths: x6 deaths in frozen and x13 deaths in fresh arm (all unrelated to FMT).</p>
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		<i>Isospora</i> and, if antigen testing unavailable, <i>Cryptosporidium</i> ; ova, cysts and parasites.	Antimotility: Not described.  Prokinetics: Not described.  Time before CDI treatment was stopped before FMT: Discontinued 24 - 48 hours prior to FMT.		
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<p>van Nood <i>et al</i>, <i>New England Journal of Medicine</i>, 2013</p>	<p>Intervention: FMT + bowel lavage. Number of patients: 16. Female: male: 8: 8. Age (mean/ standard deviation): 73 (+/- 13) years.</p> <p>Comparator: Vancomycin (500mg orally four times daily for 14 days). Number of patients: 13. Female: male: 7: 6. Age (mean/ standard deviation): 66 (+/-14) years.</p> <p>Comparator: Vancomycin (500mg orally four times daily for 14 days) + bowel lavage. Number of patients: 13. Female: Male: 3: 10. Age (mean/ standard deviation): 69 (+/-16) years.</p> <p>Comorbidities: No significant difference in median Charlson comorbidity index between groups.</p> <p>CDI features: All recurrent.</p> <p>CDI diagnosis confirmation: Toxin and PCR.</p> <p>Pre-FMT antibiotics: At least one course of adequate antibiotic therapy (<math>\geq 10</math> days of vancomycin at a dose of <math>\geq 125</math>mg four times a day or <math>\geq 10</math> days of metronidazole</p>	<p>Donors were healthy volunteers.</p> <p>Donors working in healthcare: No.</p> <p>Donor demographics: &lt;60 years of age.</p> <p>Donor screening: questionnaire: questionnaire addressed risk factors for potentially transmissible diseases.</p> <p>Travel and antibiotic exclusion period: Excluded as donor if travel to tropical area within past 3 months, or antibiotic use within the past two months.</p> <p>Screening blood tests: Blood was screened for HIV; human T-cell lymphotropic virus types 1 and 2; hepatitis A,B, and C; cytomegalovirus; Epstein-Barr virus; <i>Treponema pallidum</i>; <i>Strongyloides stercoralis</i>; and <i>Entamoeba histolytica</i>.</p> <p>Screening stool tests: Donor feces were screened for parasites, including <i>Blastocystis hominis</i> and <i>Dientamoeba fragilis</i>; <i>C</i></p>	<p>Amount of stool per transplant / administered to patients: A mean (+/-standard deviation) of 141+/- 71g of faeces was infused.</p> <p>Diluent used to prepare: Faeces were diluted with 500mls of sterile saline, 0.9%.</p> <p>Diluent used to store if frozen: N/A.</p> <p>Preparation methods: The solution was stirred, and the supernatant strained and poured in a sterile bottle.</p> <p>Time from preparation to transplant (fresh): Mean time from defecation to infusion was 3.1+/- 1.9 hours.</p> <p>Time period for storage (frozen): N/A.</p> <p>Route administered: Upper GI: 16 (via nasoduodenal tube); lower GI: nil; capsule: nil.</p> <p>Number of infusions: 16 patients had 1 infusion; 3 who did not respond in this group had 2nd infusion.</p> <p>Bowel purgative: 4 litres of macrogol solution (Klean-Prep) on the last day of antibiotic treatment.</p> <p>PPI: Not stated.</p>	<p>Treatment arm: FMT + bowel lavage Overall cure rate: 94% (<math>n=15/16</math>). Cure with one infusion alone: 81% (<math>n=13/16</math>).</p> <p>Treatment arm: Vancomycin: Overall cure rate: 31% (<math>n=4/13</math>) patients at 10 weeks.</p> <p>Treatment arm: Vancomycin + bowel lavage: Overall cure rate: 23% (<math>n=3/13</math>) patients at 10 weeks.</p>	<p>Minor GI adverse events: 94% immediate diarrhoea, 31% abdominal pain with cramping, 19% belching - resolved within 3 hours. During follow-up, x3 patients had constipation (19%).</p> <p>Minor non-GI adverse events: Nil.</p> <p>Serious adverse events: Nil described.</p> <p>Deaths: None.</p>
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	<p>at a dose of 500mg three times per day).</p> <p>Total follow up period: After first infusion at 10 weeks; follow-up was extended to 10 weeks after the second infusion.</p> <p>Cochrane Collaboration risk of bias assessment: low risk of bias.</p>	<p><i>difficile</i>, and enteropathogenic bacteria.</p>	<p>Antimotility: Not stated.</p> <p>Prokinetics: Not stated.</p> <p>Time before CDI treatment was stopped before FMT: 24 hours.</p>		
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<p>Youngster <i>et al</i>, <i>Clinical infectious diseases</i>, 2014</p>	<p>Intervention: Colonoscopic FMT. Number of patients: 10. Female: male: 6:4. Age (mean/ standard deviation): Mean 50.4 (+/- 28.8) years.</p> <p>Intervention: Nasogastric FMT. Number of patients: 10. Female: male: 5: 5. Age (mean/ standard deviation): Mean 58.6(+/-19.6) years.</p> <p>Comorbidities: Not defined.</p> <p>CDI features: Relapsing or recurring (having at least 3 episodes of mild-to-moderate <i>CDI</i> or at least 2 episodes of severe <i>CDI</i> resulting in hospitalization and associated with significant morbidity.</p> <p>CDI diagnosis confirmation: Toxin; initial GDH enzyme-linked immunosorbent assay, followed by PCR only if the GDH test is positive or indeterminate.</p> <p>Pre-FMT antibiotics: Treatment failures of a 6- to 8-week taper with vancomycin (95% of patients) with or without an alternative antibiotic, including fidaxomicin (70% of participants).</p> <p>Total follow up period: 8 weeks follow-up for primary response.</p>	<p>Donors were healthy volunteer non-pregnant adults.</p> <p>Donors working in healthcare: No.</p> <p>Donor demographics: 18-50 years of age, on no medications, with a normal body mass index.</p> <p>Donor screening: questionnaire - initial screening using the American Association of Blood Banks donor questionnaire for exposure to infectious agents.</p> <p>Travel and antibiotic exclusion period: Excluded if antibiotic use within 6 months.</p> <p>Screening blood tests: Blood was screened for antibodies to hepatitis A, B, and C; HIV; and <i>Treponema pallidum</i> within 2 weeks of donations.</p> <p>Screening stool tests: Donor faeces were screened for enteric bacterial pathogens including rotavirus, <i>Listeria monocytogenes</i>, <i>Vibrio</i></p>	<p>Amount of stool per transplant / administered to patients: 90mls of thawed FMT (41g).</p> <p>Diluent used to prepare: Normal saline.</p> <p>Diluent used to store if frozen: 10% glycerol.</p> <p>Preparation methods: Homogenised using a commercial blender then passed through sieves.</p> <p>Time from preparation to transplant (fresh): N/A.</p> <p>Time period for storage (frozen): Inocula were stored frozen for up to 156 days, range, 29-156 days.</p> <p>Route administered: Upper GI (nasogastric) 10; lower GI (colonoscopy): 10; capsule: nil.</p> <p>Number of infusions: Colonoscopy: 8 patients - 1 infusion, 2 patients – 2 infusions; NG: 7 patients - 1 infusion; 3 patients – 2 infusions.</p> <p>Bowel purgative: For colonic route - 4 liters of PEG solution.</p> <p>PPI: 20mg of omeprazole orally for 48 hours prior to FMT.</p>	<p>Treatment arm: Overall Overall cure rate: 90% (<i>n</i>=18/20). Cure with one infusion alone: 70% (<i>n</i>=14/20).</p> <p>Treatment arm: Colonoscopy: Overall cure rate: 100% (<i>n</i>=10/10). Cure with one infusion alone: 80% (<i>n</i>=8/10).</p> <p>Treatment arm: Nasogastric: Overall cure rate: 80% (<i>n</i>=8/10). Cure with one infusion alone: 60% (<i>n</i>=6/10).</p>	<p>Minor GI adverse events: Mild abdominal discomfort and bloating in x4 patients (20%). X1 child treated colonoscopically had a transient fever of 38.8°C on day 2 that resolved spontaneously.</p> <p>Minor non-GI adverse events: Nil described.</p> <p>Serious adverse events: x1 new diagnosis of malignancy, x1 hospitalisation for Fournier gangrene (unrelated to FMT).</p> <p>Deaths: x2 deaths (unrelated to FMT).</p>
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	Cochrane Collaboration risk of bias assessment: uncertain risk of bias.	<i>cholerae</i> , <i>Escherichia coli</i> O157, ova and parasites (including general microscopy, acid-fast staining, and/or antigen testing for <i>Giardia</i> , <i>Cryptosporidium</i> , <i>Isospora</i> , and <i>Microsporidia</i> ), <i>C difficile</i> , and <i>Helicobacter pylori</i> antigen.	Antimotility: single dose of oral loperamide prior to procedure.  Prokinetics: Nil.  Time before CDI treatment was stopped before FMT: Patients were required to discontinue all antibiotics at least 48 hours prior to the procedure.		
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**C.3. Reviewed randomised studies of FMT for non-CDI indications**

Confidential: For Review Only

Paper	Study and patient characteristics	Donor characteristics	FMT characteristics	Outcomes	Adverse events
Moayyedi <i>et al</i> , <i>Gastroenterology</i> , 2015	<p>Intervention: FMT. Number of patients: 38. Female: male 20: 18. Age (mean +/-range)*: 42.2+/-15.0 years.</p> <p>Comparator: Water enema. Number of patients: 37. Female: male: 11: 26. Age (mean +/-range)*: 35.8 +/-12.1 years.</p> <p>Primary outcome: Remission at week 7, defined as full Mayo score &lt; 3 and complete healing of mucosa at flexible sigmoidoscopy (endoscopic Mayo score: 0).</p> <p>Secondary outcome: Clinical response (at least 3 point reduction in Mayo score), change in Mayo, IBD Questionnaire scores, EQ-5D scores.</p> <p>Inclusion criteria: &gt;18 years with UC - Mayo at least 4 with endoscopic subscore at least 1 (included patients with severe disease).</p> <p>Exclusions - antibiotics/ probiotics in past 30 days, concomitant <i>C difficile</i>/ other enteric pathogens, disease severity requiring hospitalisation, pregnancy, unable</p>	<p>Donors were unrelated volunteers - six donors used. Plus - one patient in active treatment arm had spouse as donor (treatment failure).</p> <p>Working in healthcare: Not specifically stated.</p> <p>Donor demographics: 18-60 years.</p> <p>Donor screening: Questionnaire – yes.</p> <p>Travel and antibiotic exclusion period: Retesting of stool whenever donor travelled outside North America. Excluded as donor if antibiotics within past 3 months. Screening repeated regardless every 6 months.</p> <p>Screening blood tests: HIV, hepatitis A IgM, HBsAg, hepatitis C antibody, syphilis, HTLV-1/-2.</p> <p>Screening stool tests: MC&amp;S, ova, cysts and parasites, <i>C difficile</i> toxin, VRE, MRSA.</p>	<p>Amount of stool per transplant / administered to patients: 8.3g of stool per enema</p> <p>Diluent used to prepare: 50g of stool mixed with 300ml of commercial bottled drinking water, then 50ml of mixture administered as enema.</p> <p>Diluent used to store if frozen: No glycerol. FMT administered either fresh, or stored at -20 degrees. 21 received frozen, 15 received fresh, 1 mixture of fresh and frozen.</p> <p>Preparation methods: Not anaerobic. Single donor per FMT.</p> <p>Time from preparation to transplant (fresh): Processing within 5hr of collection.</p> <p>Time period for storage (frozen): Not stated.</p> <p>Route administered and frequency: Upper GI: nil; lower GI: enema - weekly for 6 weeks. Aimed to retain for at least 20 mins (38); capsule: nil.</p> <p>Bowel purgative: No PEG.</p> <p>PPI: Not described.</p> <p>Antimotility: Not described.</p>	<p>FMT arm: Remission rates: 24% (n=9/38). Clinical response rates: 40% (n=15/38) had reduction in full Mayo score of at least 3 points. Quality of Life Assessment: Yes - IBDQ and EQ-5D not significantly different between groups.</p> <p>Water enema arm: Remission rates: 5% (n=2/37) (p=0.03) Clinical response rates: 24% (n=9/37) had reduction in full Mayo score of at least 3 points (p=0.16).</p>	<p>FMT arm: Minor GI adverse events: Two patients developed patchy inflam in the colon and also rectal abscess formation - resolved with antibiotics.</p> <p>Minor non-GI adverse events: None.</p> <p>Serious adverse events: x2 patients had diagnosis changed to Crohn's colitis, one was <i>C difficile</i> toxin positive at end of therapy.</p> <p>Deaths: None.</p> <p>Water enema arm: Minor GI adverse events: x1 patient developed patchy inflammation in the colon and also rectal abscess formation - resolved with antibiotics.</p> <p>Minor non-GI adverse events: None.</p> <p>Serious adverse events: x1 patient changed diagnosis from UC to Crohn's colitis; x1</p>

	<p>to give informed consent.</p> <p>Concomitant medications: Stable dose thiopurines, mesalamine, corticosteroids, and anti-TNF allowed as long as stable dose for at least 12 weeks (4 weeks for steroids).</p> <p>Total follow-up period: Up to 12 months.</p> <p>Cochrane Collaboration risk of bias assessment: low risk of bias.</p>		Prokinetics: Not described.		<p>admitted with hospital with active severe colitis and required colectomy.</p> <p>Deaths: None.</p>
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Rossen <i>et al</i> , <i>Gastroenterology</i> , 2015	<p>Intervention: Donor faeces. Number of patients: 23. Female: male: 12: 11. Age (median, (range)): 40 (33-56) years.</p> <p>Comparator: Autologous faeces. Number of patients: 25. Female: male: 14:11. Age (median, (range)): 41 (30 – 48) years.</p> <p>Primary outcome: Clinical remission (defined as a SCCAI score <math>\leq 2</math>) in combination with 1-point improvement on the combined Mayo endoscopic score of the sigmoid and rectum, as compared with baseline sigmoidoscopy, 12 weeks after the first treatment.</p> <p>Secondary outcome: Endpoints at 6 and 12 weeks were clinical response (defined as a reduction of 1.5 points on the Simple Clinical Colitis Activity Index (SCCAI), a validated disease activity index tool in ulcerative colitis), clinical remission (defined as a SCCAI of <math>\leq 2</math>), endoscopic response, change in median (Inflammatory Bowel Disease Questionnaire [IBDQ]) score from baseline to shortly after treatment (week 6), and adverse events.</p> <p>Inclusion criteria: enteric infection, use of biologics within 8 weeks or</p>	<p>Donors were healthy partners, relatives, or volunteers.</p> <p>Working in healthcare: Not stated</p> <p>Donor demographics: &gt;18 yrs</p> <p>Donor screening: Questionnaire - Dutch Red Cross Questionnaire addressing risk factors for potential transmissible diseases used for screening of blood donors in The Netherlands.</p> <p>Travel and antibiotic exclusion period: Excluded as donor if antibiotics within 8 weeks.</p> <p>Screening blood tests: CMV (IgG + IgM), EBV (IgG + IgM), hepatitis A (total antibody), hepatitis B (HBsAg), hepatitis C (hepatitis C virus antibody), HIV (1+2 antibodies/antigen), HTLV (I + II antibodies), <i>Entamoeba</i> (antibodies against <i>Entamoeba histolytica</i>), <i>Strongyloides</i> (<i>Strongyloides</i> ELISA).</p> <p>Screening stools: Multiplex PCR containing probes against enteral viruses (<i>rotavirus</i>, <i>norovirus</i>, <i>enterovirus parechovirus</i>, <i>sapovirus</i>, <i>adenovirus 40/41/52</i>, <i>astrovirus</i>), FT + TFT II: PCR op <i>Giardia</i>, <i>SSYC</i>, <i>Clostridium</i> toxin</p>	<p>Amount of stool per transplant / administered to patients: 120g</p> <p>Diluent used to prepare: Normal saline</p> <p>Diluent used to store if frozen: not stated</p> <p>Preparation methods: Not anaerobic</p> <p>Time from preparation to transplant (fresh): not stated</p> <p>Time period for storage (frozen): not stated</p> <p>Route administered and frequency: Upper GI: Nasoduodenal route. 2 infusions three weeks apart. Nil lower GI or capsule</p> <p>Bowel purgative: Macrogol before both infusions</p> <p>PPI: Not described</p> <p>Antimotility: Not described</p> <p>Prokinetics: Not described</p>	<p>Donor faeces arm: Remission rates: 30% (<math>n=7/23</math>) Clinical response rates: 47.8% (<math>n=11/23</math>) at 12 weeks. Quality of Life Assessment: IBDQ only calculated based on responders vs nonresponders.</p> <p>Autologous faeces arm: Remission rates: 20% (<math>n=5/25</math>), (<math>p=0.51</math>). Clinical response rates: 52% (<math>n=13/25</math>) at 12 weeks.</p>	<p>Minor GI adverse events: 78.3% (<math>n=18/23</math>) of donor stool and 64% (<math>n=16/25</math>) of autologous stool experienced side effects post FMT: transient borborygmus, diarrhoea, vomiting, fever.</p> <p>Minor non-GI adverse events: None.</p> <p>Serious adverse events: x4 overall (small bowel perforation – secondary to Crohn’s), CMV infection, abdominal pain, cervical carcinoma.</p> <p>Deaths: Nil.</p>
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	<p>methotrexate within 4 weeks</p> <p>Concomitant medications: stable doses of thiopurines, mesalamine, or corticosteroids 10 mg/day for the 8 weeks before inclusion.</p> <p>Total follow-up period: 12 weeks.</p> <p>Cochrane Collaboration risk of bias assessment: low risk of bias.</p>				
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Paramsothy <i>et al</i> , <i>Lancet</i> , 2017	<p>Intervention: FMT. Number of patients: 41. Female: male 19: 22. Age (median, (range)): 35.6 (27.8-48.9) years.</p> <p>Comparator: Placebo-isotonic saline with added colourant odourant and glycerol cryoprotectant (concentration 10%). Number of patients: 40. Female: male: 15: 25. Age (median, (range)): 35.4 (27.7-45.6) years.</p> <p>Primary outcome: Composite of steroid-free clinical remission and endoscopic remission or response at week 8, defined as a total Mayo score of 2 or less, with all Mayo subscores of 1 or less, and at least a 1 point reduction from baseline in the endoscopy subscore.</p> <p>Secondary outcome: Secondary outcomes were: steroid-free clinical remission (defined as combined Mayo subscores of 1 or less for rectal bleeding plus stool frequency); steroid-free clinical response (defined as either a decrease of 3 points or more on the Mayo score, a 50% or greater reduction from baseline in combined rectal bleeding plus stool frequency Mayo subscores, or both); steroid-free endoscopic</p>	<p>Donors were between 3-7 unrelated donors.</p> <p>Working in healthcare: No.</p> <p>Donor demographics: Not described.</p> <p>Donor screening: Questionnaire asked regarding:</p> <ul style="list-style-type: none"><li>· Known HIV, hepatitis B or hepatitis C infection</li><li>· Known exposure to HIV or viral hepatitis within the previous 12 months</li><li>· High risk sexual behavior (e.g. sexual contact with anyone with HIV/AIDS or viral hepatitis, men who have sex with men, sex for drugs or money)</li><li>· Use of illicit drugs</li><li>· Tattoo or body piercing within the preceding 6 months</li><li>· Incarceration or history of incarceration</li><li>· Known current communicable disease (e.g. upper respiratory tract infection)</li><li>· Risk factors for variant Creutzfeldt-Jakob disease</li><li>· Travel within last 2 weeks to areas of the world where diarrhoeal illnesses are endemic or risk of traveler's diarrhea is high</li><li>· History of or current inflammatory bowel disease (IBD)</li><li>· History of or current irritable</li></ul>	<p>Amount of stool per transplant / administered to patients: 37.5g of blended stool to isotonic saline; volume of each infusion was 150ml.</p> <p>Diluent used to prepare: isotonic saline with 10% glycerol cryoprecipitant.</p> <p>Diluent used to store if frozen: -80°C with glycerol cryoprotectant (concentration 10%).</p> <p>Preparation methods: Donors had to provide faeces within 4 hours of a bowel movement, which was inspected visually for suitability (formed stool, no blood or mucous). Donor stool homogenised for a given batch on each day in a biosafety cabinet in isotonic saline then filtered. Placebo infusions comprised isotonic saline; brown food colourant, odourant, and glycerol cryoprotectant (concentration 10%) was added to all study infusions (investigational and placebo). The volume of each infusion was 150 mL. Infusions were stored at -80°C until dispensation to patients at fortnightly study visits for home freezer storage at -20°C before daily administration.</p> <p>Time from preparation to transplant (fresh): Not described.</p>	<p>Donor FMT arm: Remission rates: 275 (n=11/41). Clinical response rates: 54% (n=22/41). Quality of Life Assessment: Not described.</p> <p>Placebo arm: Remission rates: 8% (n=3/40) (p=0.021). Clinical response rates: 23% (n=9/40) (p=0.04). Quality of Life Assessment: Not described.</p>	<p>FMT arm: Minor GI adverse events: abdominal pain x12 (29%), colitis x10 (24%), flatulence x10 (24%), bloating x8 (20%), nausea x2 (5%), elevated ALT x2 (5%), vomiting x2 (5%), enterocolitis x1 (2%), diarrhoea x1 (2%), reflux x1 (2%), haemorrhoids x1 (2%), elective surgical procedure x1 (2%).</p> <p>Minor non-GI adverse events: None.</p> <p>Serious adverse events: x2 (5%) - x1 clinical deterioration and colectomy, x1 needed intravenous intravenous steroids.</p> <p>Deaths: Nil.</p> <p>Placebo arm: Minor GI adverse events: abdominal pain x11 (28%), colitis x9 (23%), flatulence x8 (20%), bloating x11 (28%), nausea x5 (13%), vomiting x1 (3%), enterocolitis x3 (8%), anal fissure x1 (3%), faecal incontinence x1</p>
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	<p>response (defined as a Mayo endoscopy subscore of 1 or less, with a reduction of at least 1 point from baseline); steroid-free endoscopic remission (defined as a Mayo endoscopy subscore of 0); quality of life (assessed with the IBDQ); and safety (assessed by adverse events).</p> <p>Inclusion criteria: 1. 18-75 years; 2. UC for &gt;3 months; 3. UC of any extent except isolated proctitis &lt;5cm; 4. currently active mild-moderate UC as measured by a Mayo score of 4-10, endoscopy score must be greater or equal to 1 and a physician global assessment score of less than or equal to 2; 5. Written consent.</p> <p>Concomitant medications: Drugs permitted as long as the dose was stable preceding enrolment: oral 5-aminosalicylates (stable dose for 4 weeks); thiopurines and methotrexate (on medication for ≥90 days and dose stable for 4 weeks); and oral prednisolone (dose ≤20mg daily and stable for 2 weeks). During the study, patients remained on the same dose of 5-aminosalicylate, thiopurine, and methotrexate. For oral prednisolone, patients received a mandatory taper of up to 2.5 mg per week so that patients would be steroid-free by week 8.</p>	<p>bowel syndrome (IBS), chronic constipation, chronic diarrhea or other intrinsic gastrointestinal illness / condition</p> <ul style="list-style-type: none"> <li>· History of or current gastrointestinal malignancy or known polyposis or strong family history of colorectal cancer</li> <li>· History of major gastrointestinal surgery (e.g. gastric bypass, partial colectomy)</li> </ul> <p>Antimicrobials (antibiotics, antivirals, antifungals), probiotics or proton pump inhibitors (PPIs) within the preceding 3 months</p> <ul style="list-style-type: none"> <li>· Major immunosuppressive medications (e.g. calcineurin inhibitors, biological agents, exogenous glucocorticoids)</li> <li>· Systemic anti-neoplastic agents</li> <li>· Household members with active GI infection</li> </ul> <p>Systemic autoimmunity (e.g. multiple sclerosis, connective tissue disease)</p> <ul style="list-style-type: none"> <li>· Atopic disease (e.g. moderate - severe asthma, eosinophilic disorders of the gastrointestinal tract)</li> <li>· Metabolic syndrome, obesity (BMI &gt;30) or moderate to severe under-nutrition / malnutrition</li> <li>· Chronic pain syndromes (e.g. chronic fatigue syndrome, fibromyalgia) or neurologic / neurodevelopmental disorders</li> <li>· History of malignant illness or ongoing oncologic therapy</li> </ul>	<p>Time period for storage (frozen): Not described.</p> <p>Route administered and frequency: Upper GI: 0; lower GI: 5 enemas per week following colonoscopic delivery -5 days on, two days off for 8 weeks (40 enemas per patient); capsule: 0.</p> <p>Bowel purgative: Yes, but no details</p> <p>PPI: Not described</p> <p>Antimotility: Not described</p> <p>Prokinetics: Not described</p>		<p>(3%), elevated ALT x2 (5%).</p> <p>Minor non-GI adverse events: None.</p> <p>Serious adverse events: x1 (3%) - admitted to hospital (no details why).</p> <p>Deaths: Nil.</p>
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	<p>Cochrane Collaboration risk of bias assessment: low risk of bias.</p>	<p>Travel and antibiotic exclusion period: Excluded if travel within last 2 weeks to areas where diarrheal illnesses are endemic or risk of travelers diarrhea is high.</p> <p>Screening blood tests: Complete blood count, electrolytes, urea and creatinine, LFTS, ESR, CRP, HIV-1 and -2, hepatitis A IgM, hepatitis B SAg, hepatitis B core antibody (IgM and IgG) and surface antibody, hepatitis c antibody, rapid plasma reagin and/or fluorescent treponemal antibody-absorbed, HTLV-1 and HTLV-2.</p> <p>Screening stools: <i>C difficile</i> PCR, faecal MC&amp;S with routine bacterial culture for enteric pathogens, <i>Giardia</i> antigen, <i>Cryptosporidium</i> antigen, faecal ova/cysts/parasites including <i>Blastocystitis hominis</i> and <i>Dientamoeba fragilis</i>, and Norovirus.</p>			
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<p>Costello <i>et al</i>, <i>Journal of Crohn's and Colitis</i> (abstract), 2017</p>	<p>Intervention: Donor FMT. Number of patients: 38. Female: male: Not stated. Age (mean/median): Not stated.</p> <p>Comparator: Control - autologous FMT in saline. Number of patients: 35. Female: male: Not stated. Age (mean/median): Not stated.</p> <p>Primary outcome: Steroid-free remission of UC, as defined by total Mayo of 2 or less with an endoscopic Mayo score of 1 or less at week 8.</p> <p>Secondary outcome: Clinical response (at least 3 point reduction in Mayo score), clinical remission (i.e. SCCAI of 2 or less), endoscopic remission (Mayo 1 or less), and safety.</p> <p>Inclusion criteria: UC - Mayo 3-10 with endoscopic subscore at least 2.</p> <p>Concomitant medications: Stable dose of immunomodulator, 5-ASA, biological, tapering prednisolone.</p> <p>Cochrane Collaboration risk of bias assessment: uncertain risk of bias.</p>	<p>Donors were healthy volunteers.</p> <p>Working in healthcare: Not clear.</p> <p>Donor demographics: Not described.</p> <p>Donor screening: Questionnaire – yes but no details described.</p> <p>Travel and antibiotic exclusion period: Not described.</p> <p>Screening blood tests: Yes but not described .</p> <p>Screening stool tests: Yes but not described.</p>	<p>Amount of stool per transplant / administered to patients: 50g of stool for first FMT, 25g of stool in subsequent enemas.</p> <p>Diluent used to prepare: 65% saline.</p> <p>Diluent used to store if frozen: Yes - frozen with 10% glycerol.</p> <p>Preparation methods: Anaerobic prep, donor stool pooled from 3-4 donors.</p> <p>Time from preparation to transplant (fresh): N/A.</p> <p>Time period for storage (frozen): Not stated.</p> <p>Route administered and frequency: Upper GI: nil; lower GI: FMT via colonoscopy on day 0, followed by 2 enemas on day 7 (38); capsule: nil</p> <p>Bowel purgative: PEG before colonoscopy but not enema</p> <p>PPI: Not described</p> <p>Antimotility: Not described</p> <p>Prokinetics: Not described</p>	<p>Donor FMT arm: Remission rates: 32% (n=12/38) in steroid-free remission at week 8. Clinical response rates: 55% (n=21/38). Quality of Life Assessment: Not described.</p> <p>Autologous FMT arm: Remission rates: 9%. (n=3/35) in steroid-free remission at week 8 (p&lt;0.01). Clinical response rates: 20% (n=7/35) (p&lt;0.01). Quality of Life Assessment: Not described.</p>	<p>Donor FMT arm: Minor GI adverse events: Nil.</p> <p>Minor non-GI adverse events: Nil.</p> <p>Serious adverse events: Worsening colitis in x2 patients</p> <p>Deaths: Nil.</p> <p>Control - autologous FMT in saline arm. Minor GI adverse events: Nil.</p> <p>Minor non-GI adverse events: None.</p> <p>Serious adverse events: Worsening colitis in x2 placebo patients. x1 patient requiring colectomy, x1 pneumonia.</p> <p>Deaths: Nil.</p>
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Johnsen et al, <i>Lancet Gastroenterology and Hepatology</i> , 2017	Intervention: Donor FMT. Number of patients: 55. Female: male: 36: 19. Age (median, (range)): 44 (33-54) years.	Donors were two volunteers screened at start and at 7 months post donation.  Working in healthcare: Not stated.	Amount of stool per transplant / administered to patients: 50 to 80g of stool in 50mls.	Donor FMT arm: Remission rates: 66% (n=36/55) . Quality of Life Assessment: Not described.  Autologous FMT arm: Remission rates: 43% (n=12/28) (p=0.49). Quality of Life Assessment: Not described.	FMT arm: Minor GI adverse events: Self limiting intermittent abdominal pain x1, self limiting nausea and vertigo x1.  Minor non-GI adverse events: Nil.  Serious adverse events: Nil.  Deaths: Nil.
	Comparator: Control - autologous FMT . Number of patients: 28. Female: male: 19: 9. Age (median (range)): 45 (34-57) years.  Primary outcome: Symptom relief of more than 75 points assessed by IBS-SSS at 3 months after FMT.  Inclusion criteria: 18-75 yrs of age, IBS with diarrhoea or mixed IBS according to Rome III criteria. Exclusion criteria: participants with severe cardiac disease, pulmonary disease, or kidney failure, non-IBS type abdominal pain, immunodeficiency or on immunomodulating agents.  Cochrane Collaboration risk of bias assessment: low risk of bias	Donor demographics: Not described.  Donor screening: Questionnaire - new tattoos or piercings in the past 3 months; high-risk sexual behaviour; former imprisonment; or history of any of the following conditions: chronic diarrhoea, constipation, inflammatory bowel disease, IBS, colorectal polyps or cancer, immunosuppression, obesity, metabolic syndrome, atopic skin disease, or chronic fatigue.  Travel and antibiotic exclusion period: Excluded if antibiotics within past three months.  Screening blood tests: Glycated haemoglobin; and serology for HIV, <i>Treponema pallidum</i> , and hepatitis A, B, and C.  Screening stool tests: <i>Salmonella</i> spp, <i>Shigella</i> spp, <i>Campylobacter</i> spp, <i>Yersinia</i> spp, and toxin-producing <i>C difficile</i> ; faecal tests for <i>Helicobacter pylori</i> antigen,	Diluent used to prepare: 200ml isotonic saline and 50mls of 85% glycerol.  Diluent used to store if frozen: glycerol, only for autologous transplants.  Preparation methods: Aerobic, stool from both donors was mixed together.  Time from preparation to transplant (fresh): 7 hours.  Time period for storage (frozen): 2-4 weeks.  Route administered and frequency: upper GI: none; lower GI: single infusion of FMT via colonoscopy; nil capsule. Bowel purgative: Picoprep.  PPI: Not described.  Antimotility: Loperamide 8mg 2 hours before.  Prokinetics: Not described.		

viruses (norovirus, rotavirus,  
Sapovirus, adenovirus),  
and faecal calprotectin.

Bajaj et al, Hepatology, 2017	<p>Intervention: Donor FMT. Number of patients: 10. Female: male: 0: 10. Age (mean+/-standard deviation): 64.5 +/- 5.1 years. Aetiology (HCV / alcohol / HCV+alcohol / NAFLD / others): 2/4/2/2/0.</p> <p>Comparator: Standard of care (lactulose/ rifaximin). Number of patients: 10. Female: male: 0: 10. Age (mean+/-standard deviation): 62.9 +/- 9.8 years. Aetiology (HCV / alcohol / HCV+alcohol / NAFLD / others): 1/5/2/1/1.</p> <p>Primary outcome: Proportion of participants with FMT-related serious adverse events (SAEs) at day 150, a composite endpoint of death, hospitalisations, emergency room visits or transmissible infections, as defined by the FDA.</p> <p>Secondary outcomes: Changes in cognitive function at day 20, cirrhosis severity (MELD score, albumin), changes in liver function and white blood cell (WBC) count, development of all adverse events (AEs), and changes in microbiota composition and function in the FMT arm compared to standard of care arm.</p>	<p>Single donor only - identified based on highest relative abundances of <i>Lachnospiraceae</i> and <i>Ruminococcaceae</i> (16S rRNA gene sequencing analysis) among a universal stool donor bank (OpenBiome).</p> <p>Working in healthcare: Not stated.</p> <p>Donor demographics: Not described</p> <p>Donor screening: Based on OpenBiome screening. 178-point clinical assessment for infectious and microbiome-mediated diseases and 30 stool pathogen and serological tests before and after the stool is collected.</p> <p>Screening blood tests: HIV-1/-2 status, hepatitis A/B/C, <i>Treponema pallidum</i>, LFT, Complete Blood Count (CBC) (Includes differentials and platelets), HTLV-I/II antibody, with Reflex to Confirmatory Assay.</p> <p>Screening stool tests: <i>Clostridium difficile</i> Toxin B and PCR, <i>Cyclospora</i> and <i>Isospora</i> Examination, ova, cysts and parasites with <i>Giardia</i> Antigen EIA, <i>Salmonella</i>/ <i>Shigella</i>/ <i>Campylobacter</i> Culture, Shiga Toxin EIA with Reflex to <i>E. coli</i> O157 Culture and <i>Vibrio</i> Culture, <i>Cryptosporidium</i> Antigen EIA, <i>Helicobacter pylori</i> Antigen EIA,</p>	<p>Amount of stool per transplant / administered to patients: 37.5g of stool.</p> <p>Diluent used to prepare: 90mls glycerol saline buffer in total.</p> <p>Diluent used to store if frozen: glycerol.</p> <p>Preparation methods: Aerobic.</p> <p>Time from preparation to transplant (fresh): N/A - frozen.</p> <p>Time period for storage (frozen): not stated.</p> <p>Route administered and frequency: Upper GI: non; lower GI: Single infusion of FMT via enema.</p> <p>Bowel purgative: Picoprep.</p> <p>PPI: Not described.</p> <p>Antimotility: Loperamide 8mg 2 hrs before.</p> <p>Prokinetics: None.</p> <p>Others: Lactulose and rifaximin were continued for all patients throughout the trial. A 5-day broad-spectrum coverage regimen was used (metronidazole 500 mg orally three times daily, ciprofloxacin 500 mg orally twice-daily, and amoxicillin</p>	<p>FMT arm: Patients with SAEs at day 150: 20% (<i>n</i> =2/10) (<i>p</i>=0.02).</p> <p>Total SAEs at day 150: 20% (<i>n</i> =2/10) (<i>p</i>=0.01).</p> <p>Patients with altered mental status by day 150: 0% (<i>n</i> =0/10) (<i>p</i>=0.03).</p> <p>Total HE episodes at day 150: 0% (<i>n</i> =0/10) (<i>p</i>=0.03).</p> <p>Stroop OffTime+OnTime change (day 0 and day 20); positive indicates improvement: 29.1 +/- 27.9 (<i>p</i>=0.04) (N.B. Stroop OffTime+OnTime is a validated tool for objectively assessing for hepatic encephalopathy using a smartphone app).</p> <p>PHES score change (day 0 and day 20); negative indicates improvement - 3.1+/-2.1 (<i>p</i>=0.01).</p> <p>MELD score change (day 0 and day 35): 0.1+/-2.0 (<i>p</i>=0.78).</p> <p>Standard of care arm: Patients with SAEs at day 150: 80% (<i>n</i> =8/10).</p>	<p>FMT arm: Serious adverse events: x1 hospitalisation for acute kidney injury, and 1 was due to chest pain (all within 5 months post FMT).</p> <p>Deaths: Nil.</p> <p>Standard of care arm: Serious adverse events: x11 in total. x9 events linked to liver-related complications, of which x4 needed hospitalisation. x1 patient developed pneumonia and x1 developed gastroenteritis.</p> <p>Deaths: Nil.</p>
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<p>Inclusion criteria: &gt;18 yrs outpatients with cirrhosis and recurrent hepatic encephalopathy (HE) defined as at least two documented overt HE episodes requiring therapy.</p> <p>Exclusion criteria: MELD score &gt;17, on oral or intravenous antimicrobial agents besides nonabsorbable rifaximin, allergies to pretreatment antibiotics, immunosuppressive medications, positive C. difficile test, pregnancy, active infection, those with active alcohol abuse, and unable to provide informed consent</p> <p>Cochrane Collaboration risk of bias assessment: low risk of bias</p>	<p>Stool Norovirus EIA, Stool Rotavirus Antigen Detection, Adenovirus Antigen Detection, Gastroenteritis EIA, Vancomycin-resistant Enterococcus Culture, <i>Microsporidia</i> Exam.</p>	<p>500 mg orally three times daily). All antibiotics were discontinued at least 12 hours before FMT. This regime was not used in patients randomised to standard of care arm.</p>	<p>Total SAEs at day 150: 11.</p> <p>Patients with altered mental status day 150: 50% (n =5/10).</p> <p>Total HE eps day 150: 6</p> <p>Stroop OffTime+OnTime change (day 0 and day 20): - 43.5 +/- 95.7.</p> <p>PHES score change (day 0 and day 20): 0.0 +/- 3.1.</p> <p>MELD score change (day 0 and day 35): 0.2 +/- 2.7.</p> <p>N.B. no significant difference in serum albumin, AST, ALT, WBC or haemoglobin counts between the two groups.</p>	
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Tian <i>et al</i> , <i>PLoS ONE</i> , 2017	<p>Intervention: Donor FMT (one for six days in a row). Number of patients: 30. Female: male 19: 11. Age (mean+/-SD): 53.1 +/- 10.2 years.</p> <p>Comparator: Standard of care (education, behavioural strategies, oral laxatives; expressively told to avoid antibiotics). Macrogol permitted if no bowel movement for three days, and enema permitted if even this failed. Number of patients: 30. Female: male 21: 9. Age (mean+/-SD)*: 55.4 +/- 12.1 years.</p> <p>Primary outcome: At least three complete spontaneous bowel movements (CSBMs) per week during the 12 week follow-up.</p> <p>Secondary outcomes: 1) Proportion of patients with average increase of at least 1 CSBM per week; 2) Number of CSBMs per week; 3) Colonic transit time (assessed via abdominal x-ray/ radiopaque markers); 4) subjective stool consistency; 5) Wexner constipation scale.</p> <p>Inclusion criteria: ≥18 yrs outpatients with cirrhosis and recurrent hepatic encephalopathy (HE) defined as at last two</p>	<p>One universal donor used throughout (24 year old healthy university student).</p> <p>Working in healthcare: No.</p> <p>Donor demographics: As above.</p> <p>Donor screening: Similar to FDA blood screening.</p> <p>Screening blood tests: Full blood count, chemistry and iron profile, hepatitis A, B and C, HIV-1 and-2, CMV, EBV, HSV, VZV, and <i>Treponema pallidum</i>.</p> <p>Screening stool tests: <i>Yersinia spp</i>, <i>Salmonella spp</i>, <i>Shigella spp</i>, <i>Campylobacter jejuni</i>, <i>C difficile</i> toxin, helminths, ova, parasites, and <i>Helicobacter pylori</i>.</p>	<p>Amount of stool per transplant / administered to patients: 100g of stool.</p> <p>Diluent used to prepare: Either 500mls normal saline, or normal saline amended with glycerol to final concentration of 10%.</p> <p>Diluent used to store if frozen: Glycerol.</p> <p>Preparation methods: Not stated.</p> <p>Time from preparation to transplant (fresh): 2 hours.</p> <p>Time period for storage (frozen): 1-4 weeks.</p> <p>Route administered and frequency: Upper GI: all via nasojejunal tube (originally placed endoscopically); lower GI: nil.</p> <p>Bowel purgative: Not described.</p> <p>PPI: Not described.</p> <p>Antimotility: Not described.</p> <p>Prokinetics: None.</p>	<p>Donor FMT arm Meeting primary outcome: 37% (n=11/30) (p=0.04).</p> <p>Meeting second outcomes: At least one more CSBM per week: 53% (n=16/30) (p=0.009).</p> <p>Number of CSBMs per week: 3.2+/-1.4.</p> <p>Stool consistency score: 3.9+/-1.3.</p> <p>Colonic transit time (hours): 58.5+/-9.8.</p> <p>Wexner constipation score: 8.6+/-1.5.</p> <p>Quality of Life Assessment: Not described.</p> <p>Autologous FMT arm: Meeting primary outcome: 13% (n=4/30)</p> <p>Meeting second outcomes: At least one more CSBM per week: 20% (n=6/30).</p> <p>Number of CSBMs per week: 2.1+/-1.2.</p> <p>Stool consistency score: 2.4+/-1.1.</p>	<p>FMT arm: 50 in total (1 x sedation contraindications, x22 endoscopy-related respiratory difficulty, x12 nausea, x5 abdominal pain, x4 diarrhoea, x4 flatulence, x2 transient fever).</p> <p>Placebo arm: x4 in total (x0 sedation contraindications, x0 endoscopy-related respiratory difficulty, x0 nausea, x3 abdominal pain, x0 diarrhoea, x1 flatulence, x0 transient fever).</p>
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	<p>documented overt HE episodes requiring therapy.</p> <p>Exclusion criteria: At least 18 years, BMI of 18-25 kg/m<sup>2</sup>, and slow transit constipation defined as colonic transit time of &gt;48hr, and symptoms unresponsive to dietary modification, enemas or biofeedback in the previous six months.</p> <p>Cochrane Collaboration risk of bias assessment: low risk of bias.</p>			<p>Colonic transit time (hours): 73.6+/-8.7.</p> <p>Wexner constipation score: 12.7+/-2.5.</p> <p>Quality of Life Assessment: Not described.</p>	
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Vrieze et al, <i>Gastroenterology</i> , 2012	Intervention: Donor FMT Number of patients: 9. Female: male 0: 9. Age (mean+/-SD): 47 +/- 4 years.		Amount of stool per transplant / administered to patients: Not stated.		
	Comparator: Autologous FMT. Number of patients: 9. Female: male 0: 9. Age (mean+/-SD): 53 +/- 3 years.	Lean healthy Caucasian males (body mass index < 23 kg/m <sup>2</sup> .	Diluent used to prepare: 500mls of normal saline.	Donor FMT arm: Median rate of glucose disappearance, Rd: from 26.2 to 45.3 µmol/kg/min; <i>p</i> <0.05).	
	Primary outcome: Effect of lean donor gut microbiota infusion on insulin sensitivity after 6 weeks.	Working in healthcare: Not stated.	Diluent used to store if frozen: N/A.	Autologous FMT arm: Median rate of glucose disappearance, Rd: from 18.9 to 19.5 µmol/kg/min).	
	Secondary outcomes: Change in specific small- and large-gut microbiota as well as produced fecal short chain fatty acids	Donor demographics: As above.	Preparation methods: Faeces was covered with sterile saline (500 ml 0.9% NaCl) to reduce exposure to oxygen, transferred to a blender, and mixed for 10 minutes. The homogenized solution then was filtered twice through a clean metal sieve.	Quality of Life Assessment: Not described.	
	Inclusion criteria: Male Caucasian obese subjects with characteristics of the metabolic syndrome, specifically with a body mass index > 30 kg/m <sup>2</sup> , or waist circumference > 102 cm, and a fasting plasma glucose level > 5.6 mmol/L.	Donor screening: Questionnaires regarding diet and bowel habits, travel history, comorbidity including (family history of) diabetes mellitus, and lack of medication use.	Time from preparation to transplant (fresh): Same day.	Secondary outcomes: No change in the total numbers of fecal bacteria (allogenic, from 10.8 +/- 0.2 to 11.0 +/- 0.4 vs autologous, from 11.6 +/- 0.6 to 11.3 +/- 0.4 log <sub>10</sub> bacteria/g faeces, non significant [NS]). Fecal short-chain fatty acids decreased after allogenic gut microbiota infusion (median acetate from 49.5 to 37.6; <i>p</i> <0.05; butyrate, from 14.1 to 8.9; <i>p</i> < 0.05; and propionate, from 18.2 to 16.3 mmol/kg feces; NS).	No adverse events
	Exclusion criteria: History of cholecystectomy were excluded, as well as subjects who used any medication, probiotics, and/or antibiotics in the past 3 months.	Screening blood tests: Human immunodeficiency virus; human T-lymphotropic virus; hepatitis A, B, and C; cytomegalovirus; Epstein–Barr virus; <i>Strongyloides</i> ; and amoebiasis.	Time period for storage (frozen): N/A.		
	Cochrane Collaboration risk of bias assessment: low risk of bias.	Screening stool tests: Presence of parasites (eg, <i>Blastocystis hominis</i> or <i>Dientamoeba fragilis</i> ), <i>Clostridium difficile</i> , or other pathogenic bacteria ( <i>Shigella</i> , <i>Campylobacter</i> , <i>Yersinia</i> , <i>Salmonella</i> )	Route administered and frequency: Upper GI: all via nasoduodenal tube (originally placed endoscopically); lower GI: nil.		
			Bowel purgative: PEG solution.		
			PPI: Not described.		
			Antimotility: Not described.		
			Prokinetics: None.		

<p>Kootte et al, <i>Cell Metabolism</i>, 2017</p>	<p>Intervention: Donor FMT Number of patients: 26. Female: male 0: 26. Age (mean): 54 years.</p> <p>Comparator: Autologous FMT. Number of patients: 12. Female: male 0: 12. Age (mean): 54 years.</p> <p>Primary outcome: Change in intestinal microbiota composition upon FMT in relation to insulin sensitivity.</p> <p>Secondary outcomes: Post-prandial lipid, glucose excursions and plasma metabolites</p> <p>Inclusion criteria: All adult (age 21-69 years) Caucasian males, who had obesity (body mass index (BMI) &gt; 30 kg/m<sup>2</sup>), fulfilled the National Cholesterol Education Program (NCEP)-criteria for metabolic syndrome, were treatment-naïve and who were otherwise healthy.</p> <p>Exclusion criteria: History of recent weight loss, cardiovascular event, cholecystectomy and the use of any medication known to influence gut microbial composition in the last three months (including proton pump inhibitors, antibiotics and pre-/pro-/synbiotics) or treatments targeting metabolic diseases.</p>	<p>Lean healthy Caucasian males (body mass index &lt; 25 kg/m<sup>2</sup>.</p> <p>Working in healthcare: Not stated.</p> <p>Donor demographics: As above.</p> <p>Donor screening: Questionnaires regarding diet and bowel habits, travel history, comorbidity including (family history of) diabetes mellitus, and lack of medication use.</p> <p>Screening blood tests: Human immunodeficiency virus; human T-lymphotropic virus; hepatitis A, B, and C; cytomegalovirus; Epstein-Barr virus; <i>Strongyloides</i>; lues and amoebiasis</p> <p>Screening stool tests: Pathogenic parasites (e.g., <i>Blastocystis hominis</i>, <i>dientamoeba fragilis</i>, <i>giardia lamblia</i>), bacteria (<i>Shigella</i>, <i>Campylobacter</i>, <i>Yersinia</i>, <i>Salmonella</i>, enteropathogenic <i>E. coli</i> and <i>Clostridium difficile</i>) or viruses (noro-, rota-, astro-, adeno (40/41/52)-, entero-, parecho- and sapovirus).</p>	<p>Amount of stool per transplant / administered to patients: Not stated.</p> <p>Diluent used to prepare: 500mls of normal saline.</p> <p>Diluent used to store if frozen: N/A.</p> <p>Preparation methods: Faeces was covered with sterile saline (500 ml 0.9% NaCl) to reduce exposure to oxygen, transferred to a blender, and mixed for 10 minutes. The homogenized solution then was filtered twice through a clean metal sieve.</p> <p>Time from preparation to transplant (fresh): Same day.</p> <p>Time period for storage (frozen): N/A.</p> <p>Route administered and frequency: Upper GI: Single infusion all via nasoduodenal tube (originally placed endoscopically). A subgroup of patients receiving donor FMT had a second infusion; lower GI: nil.</p> <p>Bowel purgative: PEG solution.</p> <p>PPI: Not described.</p> <p>Antimotility: Not described.</p> <p>Prokinetics: None.</p>	<p>Donor FMT arm: improved peripheral insulin sensitivity at week 6 (from 25.8 to 28.8 µmol/kg/min, , <math>p &lt; 0.05</math>. This change was no longer significant at week 18 (including those that had a second infusion).</p> <p>Autologous FMT arm: FMT had no effect at week 6 (from 22.5 to 20.8 µmol/kg/min, NS)</p> <p>Quality of Life Assessment: Not described.</p> <p>Secondary outcomes: No significant changes in fecal butyrate levels (butyrate from 13 to 20 mmol/g faeces, <math>p = 0.096</math>). Fecal acetate levels, however, were significantly increased from 62 to 85] mmol/g feces (<math>p &lt; 0.05</math>) after allogenic FMT, whereas fecal propionate was borderline significantly altered (from 23 to 28 mmol/g faeces, <math>p = 0.062</math>).</p>	<p>No adverse events</p>
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	Cochrane Collaboration risk of bias assessment: low risk of bias.				
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Confidential: For Review Only

## Appendix D. Excluded clinical studies

### D.1. *Clostridium difficile* infection:

#### D.1.1. Studies excluded at Sift 2 by working group:

Paper:	Grounds for exclusion:
Allegretti JR, Allegretti AS, Phelps E, <i>et al.</i> Asymptomatic <i>Clostridium difficile</i> carriage rate post-fecal microbiota transplant is low: a prospective clinical and stool assessment. <i>Clin Microbiol Infect</i> 2017; doi: 10.1016/j.cmi.2017.10.022	Prospective case series of FMT for CDI, but insufficient patient data to fully populate data table (study primarily designed to evaluate <i>C. difficile</i> carriage post-FMT).
Aroniadis OC, Brandt LJ, Greenberg A, <i>et al.</i> Long-term follow-up study of fecal microbiota transplantation for severe and/or complicated <i>Clostridium difficile</i> infection: a multicenter experience. <i>J Clin Gastroenterol</i> 2016;50(5):398-402.	Case series of FMT for CDI, but insufficient patient data to fully populate data table.
Cammarota G, Ianiro G, Masucci L, <i>et al.</i> OC.12.9 Fecal microbiota transplantation for recurrent <i>C. difficile</i> infection: a 2-year experience from a European referral centre. <i>Dig Liver Dis</i> 2016;48 S2:e118.	Case series of FMT for CDI, but abstract only.
Dutta SK, Girortra M, Garg S, <i>et al.</i> Efficacy of combined jejunal and colonic fecal microbiota transplantation for recurrent <i>Clostridium difficile</i> infection. <i>Clin Gastroenterol Hepatol</i> 2014;12(9):1572-1576.	Prospective case series of FMT for CDI, but heterogenous primary endpoint (combination of clinical symptoms and <i>C difficile</i> toxin, but assessed between 1-3 months after FMT).
Ganc AJ, Ganc RL, Reimao SM, <i>et al.</i> Fecal microbiota transplant by push enteroscopy to treat diarrhea caused by <i>Clostridium difficile</i> . <i>Einstein</i> 2015;13(2):338-339.	Case series of FMT for CDI, but insufficient patient data to fully populate data table.
Ganc A, Ganc R, Frisoli Jr A, <i>et al.</i> Fecal transplantation – an original per-oral endoscopic technique with a pediatric colonoscope. <i>J Gastroenterol Hepatol</i> 2013;28 S3:115	Case series of FMT for CDI, but abstract only.
Jorup-Ronstrom C, Hakanson A, Sandell S, <i>et al.</i> Fecal transplant against relapsing <i>Clostridium difficile</i> -associated diarrhea in 32 patients. <i>Scand J Gastroenterol</i> 2012;47(5):548-552.	Case series of 'FMT' for CDI, but bacteriotherapy rather than true FMT.
Kao D, Roach B, Beck P, <i>et al.</i> A dual center, randomized trial comparing colonoscopy and oral capsule delivered fecal microbiota transplantation in the treatment of recurrent <i>Clostridium difficile</i> infection: preliminary results. <i>Am J Gastroenterol</i> 2015;110:S553.	Abstract of RCT of capsulised vs colonoscopic FMT for CDI, but same trial/ data set reported in more developed stage at later date <sup>48</sup> , so this abstract excluded.
Mah XJ, Paramsothy R, Lo-Cao E, <i>et al.</i> Faecal microbiota transplant (FMT) for recurrent and life	Case series of FMT for CDI, but abstract only.

threatening <i>Clostridium difficile</i> infection. <i>J Gastroenterol Hepatol</i> 2016;31:167-168.	
Mandali A, Ward A, Tauxe W, <i>et al.</i> Fecal transplant is as effective and safe in immunocompromised as non-immunocompromised patients for <i>Clostridium difficile</i> . <i>Int J Colorectal Dis</i> 2016;31(5):1059-1060.	Case series of FMT for CDI, but insufficient patient data to fully populate data table.
Oprita R, Bratu M, Oprita B, <i>et al.</i> Fecal transplantation – the new, inexpensive, safe, and rapidly effective approach in the treatment of gastrointestinal tract disease. <i>J Med Life</i> 2016;9(2):160-162.	Prospective case series of FMT for CDI or UC, but insufficient patient data to fully populate data table.
Ott SJ, Waetzig GH, Rehman A, <i>et al.</i> Efficacy of sterile fecal filtrate transfer for treating patients with <i>Clostridium difficile</i> infection. <i>Gastroenterology</i> 2017;152(4):799-811.	Case series of 'FMT' for CDI, but only five patients. Furthermore, sterile faecal filtrate rather than true FMT.
Orenstein R, Dubberke E, Hardi R, <i>et al.</i> Safety and durability of RBX2660 (microbiota suspension) for recurrent <i>Clostridium difficile</i> infection: results of the PUNCH CD study. <i>Clin Infect Dis</i> 2016;62(5):596-602.	Prospective case series of FMT for CDI, but using 'microbiota suspension' derived from stool rather than conventional FMT.
Ray A, Jones C, Shannon B, <i>et al.</i> Does the donor matter? Results from PUNCH CD 2: a randomized controlled trial of a microbiota-based drug for recurrent <i>Clostridium difficile</i> infection. <i>Am J Gastro</i> 2016;111:S65-S66.	Abstract of RCT of treatment for CDI, but 'microbiota suspension' rather than true FMT.
Ray A, Smith R, Breaux J. Fecal microbiota transplantation for <i>Clostridium difficile</i> infection: the Ochsner experience. <i>Ochsner Journal</i> 2014;14(4):538-544.	Case series of FMT for CDI, but heterogenous primary end point.
Rupali P, Mittal C, Deol A, <i>et al.</i> Fecal microbiota transplantation for <i>Clostridium difficile</i> infection in immunocompromised hosts: one easy strategy, one giant success. <i>Transplantation</i> 2014;98:687-688.	Case series of FMT for CDI, but abstract only.
Russell GH, Kaplan JL, Youngster I, <i>et al.</i> Fecal transplant for recurrent <i>Clostridium difficile</i> infection in children with and without inflammatory bowel disease. <i>J Pediatric Gastroenterol Nut</i> 2014;58(5):588-592.	Case series of FMT for CDI, but all children, and presented as separate cases rather than as group of 10 recipients.
Tauxe WM, Haydek JP, Rebolledo PA, <i>et al.</i> Fecal microbiota transplant for <i>Clostridium difficile</i> infection in older adults. <i>Ther Adv Gastroenterol</i> 2016;9(3):273-281.	Case series of FMT for CDI, but heterogenous primary end point.
True E, Tsoraidis S, Wang H, <i>et al.</i> Predictors of failure with fecal microbiota therapy for recurrent <i>Clostridium difficile</i> colitis. <i>Dis Colon Rectum</i> 2014;57(5):e99-e100.	Case series of FMT for CDI, but abstract only.
Tvede M, Tinggaard M, Helms M. Rectal bacteriotherapy for recurrent <i>Clostridium difficile</i> -associated diarrhoea: results from a case series of 55	Case series of 'FMT' for CDI, but bacteriotherapy rather than true FMT.

patients in Denmark 2000-2012. *Clin Micro Infect* 2015;21(1):48-53.

#### D.1.2. Abstracts not fulfilling selection criteria:

Borody TJ, Wettstein A, Nowak A, Finlayson S, Leis S. Fecal microbiota transplantation (FMT) eradicates clostridium difficile infection (CDI) in inflammatory bowel disease (IBD). *United Eur Gastroenterol J*. 2013;1(PG-A57):A57.

D.N. S, Seril DN, Shen B. Clostridium difficile infection in patients with ileal pouches. *Am J Gastroenterol*. B. Shen, Department of Gastroenterology/Hepatology-A31, Digestive Disease Institute, Cleveland Clinic Foundation, 9500 Euclid Avenue, Cleveland, OH 44195, United States. E-mail: shenb@ccf.org: Nature Publishing Group (Houndmills, Basingstoke, Hampshire RG21 6XS, United Kingdom); 2014;109(7):941–7.

Ganc AJ, Ganc RL. Fecal microbiota transplantation, by means of push enteroscopy. A novel endoscopic technique, for the treatment of chronic diarrhea associated with clostridium difficile-a pilot study. *Gastrointest Endosc*. 2014;1(PG-AB380-AB381):AB380-AB381.

Garg S, Fricke WF, Girotra M, Dutta A, Von Rosenvinge EC, Dutta S. Recurrent clostridium difficile infection: A longitudinal study of alterations in fecal microbiome in patients-donor pairs before and after fecal microbiota therapy. *Gastroenterology*. 2013;1(PG-S184-S185):S184–5.

Garg S, Fricke WF, Girotra M, Von Rosenvinge EC, Dutta A, Dutta SK. Emerging role of fecal microbiota therapy in the treatment of recurrent clostridium difficile infection in children. *Gastroenterology*. 2013;1(PG-S45):S45.

Garg S, Song Y, Han MAT, Girotra M, Fricke WF, Dutta S. Post-infectious irritable bowel syndrome in patients undergoing fecal microbiota transplantation for recurrent clostridium difficile colitis. *Gastroenterology*. 2014;1(PG-S83-S84):S83–4.

Girotra M, Bartlett J, Koerner K, Dutta S. Combined jejunal and colonic fecal bacteriotherapy in patients with recurrent clostridium difficile infection (RCDI). *Am J Gastroenterol*. 2011;106(PG-S162-S163):S162–3.

Girotra M, Dutta A, Koerner K, Bodner B, Dutta SK. Recurrent clostridium difficile infection (RCDI) in geriatric patients: A long-term follow up of simultaneous jejunal and colonic administration of fecal bacteriotherapy (FT). *Gastroenterology*. 2012;1(PG-S130):S130.

Goyal A, Chu A, Calabro K, Firek B, Bush B, Morowitz M. Safety and efficacy of fecal microbiota transplant in children with inflammatory bowel disease. *J Pediatr Gastroenterol Nutr*. 2016;63(PG-S212):S212.

Goyal A, Kufen A, Jackson Z, Morowitz M. A study of fecal microbiota transplantation in pediatric patients with inflammatory bowel disease. *Inflamm Bowel Dis*. 2016;22:S74.

Graham D, Attumi T, Opekun A, Metcalf G, Muzny D, Hyde E, et al. Triple bacteroides fecal replacement therapy for relapsing clostridium difficile diarrhea (fecal transplantation sans feces). *Am J Gastroenterol*. 2013;108(PG-S170):S170.



- Greenberg A, Aroniadis O, Shelton C, Brandt L. Long-term follow-up study of fecal microbiota transplantation (FMT) for inflammatory bowel disease (IBD). *Am J Gastroenterol*. 2013;108(PG-S540):S540.
- Greenwald D, Patel T, Barto A. Fecal microbiota transplant for treatment of refractory *C. Difficile* colitis: Long-term follow-up of 58 patients. *Am J Gastroenterol*. 2014;109(PG-S679):S679.
- Greig J, Swope LK, Calvin H. Shaking up *clostridium difficile* infections: Implementation of a fecal microbiota transplant program. *Am J Infect Control*. 2014;1(PG-S4-S5):S4–5.
- Grzesiowski P, Hermann A, Dubaniewicz A, Kasprzyk J, Pawlik D, Zak-Pulawska Z. Effectiveness of FMT in recurrent *Clostridium difficile* infection. *Antimicrob Resist Infect Control Conf 3rd Int Conf Prev Infect Control ICPIIC*. 2015;4(no pagination PG-).
- Gupta S, He SM, Noordhof C, Allen-Vercoe E, Petrof EO. Minimalist defined gut microbial ecosystem demonstrates protection against *clostridium difficile* toxin-mediated effects in vitro via toxin degradation. *Gastroenterology*. 2016;1(PG-S544):S544.
- Haran M, Tsang T, Kupfer Y, Tessler S. Intravenous immunoglobulins in severe *clostridium difficile* colitis. *Chest Conf CHEST*. 2011;140(4 MEETING ABSTRACT PG-). *Gastroenterol*. 2016;9(2 PG-229-239):229–39.
- Harrison MJ, Burke D, Fleming C, McCarthy M, Shortt C, O’Callaghan G, et al. *Clostridium difficile* in adult cystic fibrosis (CF): Prevalence, ribotyping and toxigenic capability. A prospective study. *J Cyst Fibrosis Conf 36th Eur Cyst Fibros Conf Lisbon Port Conf Start*. 12(pp S6 PG-).
- Holvoet T, Boelens J, Joossens M, Raes J, De Vos M, De Looze D. Fecal microbiota transplantation in irritable bowel syndrome with bloating: Results from a prospective pilot study. *Gastroenterology*. 2015;1(PG-S963-S964):S963–4.
- Holzwanger EA, Kaufman D, Foley A, Pellish R. Fecal microbiota transplantation via colonoscope: A single-center experience. *Am J Gastroenterol*. 2016;111(PG-S1232):S1232.
- Hourigan S, Ann Chen L, Grigoryan Z, Laroche G, Weidner M, Sears CL, et al. Microbiome changes associated with sustained eradication of *clostridium difficile* after fecal microbiota transplantation in children with and without inflammatory bowel disease. *Gastroenterology*. 2015;1(PG-S45):S45.
- Hubble L, Joshua S, Glover PH, Trivedi A, Pfanner TP. Colonoscopic vs. Upper endoscopic placement of fecal microbiota transplant for recurrent *clostridium difficile* infection: A retrospective review. *Gastroenterology*. 2015;1(PG-S728):S728.
- Ihara S, Hirata Y, Serizawa T, Suzuki N, Kinoshita H, Nakagawa H, et al. Transforming growth factor-beta signaling on dendritic cells regulates bacterial communities and gut homeostasis. *Gastroenterology*. 2014;1(PG-S-113):S-113.
- Ihunnah C, Khoruts A, Fischer M, Afzali A, Aroniadis O, Barto A, et al. Fecal microbiota transplantation (FMT) for treatment of *clostridium difficile* infection (CDI) in immunocompromised patients ACG governors award for excellence in clinical research. *Am J Gastroenterol*. 2013;108(PG-S179-S180):S179–80.
- Ishikawa D, Osada T, Haga K, Kodani T, Shibuya T, Watanabe S. Combination therapy of fresh faecal microbial transplantation and antibiotics for ulcerative colitis. *J Crohn’s Colitis*. 2016;10(PG-S335-S336):S335–6.

- Jain A, Parian AM, Dudley-Brown S, Lazarev M. Fecal microbiota transplantation is safe and effective for treatment of recurrent clostridium difficile infection in inflammatory bowel disease patients. *Gastroenterology*. 2015;1(PG-S869):S869.
- Jamot S, Kelly CR, Shah S. Won the battle, lost the war: Crohn's flare after fecal microbiota transplant (FMT) for recurrent C. Difficile infection. *Am J Gastroenterol*. 2016;111(PG-S833-S834):S833-4.
- Kassam Z, Hundal R, Marshall JK, Lee CH. Fecal transplantation via retention enema is effective for recurrent or refractory clostridium difficile-associated diarrhea. *Gastroenterology*. 2010;1(PG-S207-S208):S207-8.
- Kellermayer R, Hollister EB, Nagy-Szakal D, Ihekweazu FD, Haynes A, Pitashny M, et al. Special considerations for fecal microbiota transplantation in pediatric recurrent clostridium difficile infection. *Gastroenterology*. 2015;1(PG-S961-S962):S961-2.
- Khanna S, Kashyap P, Rainey J, Loftus E, Pardi D. Outcomes from fecal microbiota transplantation in adults with C. difficile infection and inflammatory bowel disease. *Am J Gastroenterol*. 2013;108(PG-S508):S508.
- Khanna S, Weatherly R, Kammer P, Pardi D. Management and outcomes of patients with failed fecal microbiota transplantation for recurrent clostridium difficile infection. *Am J Gastroenterol*. 2015;110(PG-S580):S580.
- Khanna S, Weatherly RM, Kammer PP, Loftus E V, Pardi DS. Long-term follow-up after fecal microbiota transplantation for C. Difficile infection in inflammatory bowel disease patients. *Gastroenterology*. 2015;1(PG-S726):S726.
- Khoruts A, Hamilton MJ, Weingarden A, Sadowsky MJ. Treatment of C. difficile by fecal transplantation. *Gastrointest Endosc*. 2012;1(PG-AB329):AB329.
- Khoruts A, Rank KM, Viskocil K, Newman KM. Diagnostic value of colonoscopy in patients receiving fecal microbiota transplantation in treatment of refractory clostridium difficile infection. *Gastroenterology*. 2015;1(PG-S729):S729.
- Kukkadapu T, Chintalapally R, Daram S. Clostridium difficile infection in adult patients with acute myeloid leukemia: Incidence, recurrence, and outcomes. *Am J Gastroenterol*. 2015;110(PG-S590):S590.
- Kump PK, Grochenig HP, Spindelbock W, Hoffmann CM, Gorkiewicz G, Wenzl H, et al. Preliminary clinical results of repeatedly fecal microbiota transplantation (FMT) in chronic active ulcerative colitis. *United Eur Gastroenterol J*. 2013;1(PG-A57):A57.
- Kump PK, Wurm P, Grochenig HP, Reiter L, Hoffmann KM, Spindelboeck W, et al. Impact of antibiotic treatment before faecal microbiota transplantation (FMT) in chronic active ulcerative colitis. *United Eur Gastroenterol J*. 2015;1(PG-A437):A437.
- LaBarbera F, Jackson W, Surace L. FMT in our ASC: Successful fecal microbiota transplantation for recurrent clostridium difficile infections in an ambulatory surgical center. *Am J Gastroenterol*. 2015;110(PG-S559-S560):S559-60.
- Lan N, Stocchi L, Remzi FH, Shen B. Fecal microbiota transplantation for recurrent clostridium difficile infection in patients with ileal pouches. *Gastroenterology*. 2016;1(PG-S542):S542.



- Landy J, Al-Hassi HO, Mann ER, Peake ST, McLaughlin SD, Ciclitira PJ, et al. A prospective controlled pilot study of fecal microbiota transplantation for chronic refractory pouchitis. *Gastroenterology*. 2013;1(PG-S897):S897.
- Landy J, Omar H, Ronde E, Mann E, Peake S, McLaughlin S, et al. A prospective controlled pilot study of faecal microbiota transplantation for chronic refractory pouchitis. *J Crohn's Colitis*. 2013;7:S247–8.
- Licht E, Maltz C. The potential role of lactulose in the prevention of clostridium difficile diarrhea. *Am J Gastroenterol*. 2012;107(PG-S203-S204):S203–4.
- Lin E, Jaworski A, Furnari V, Wong C, Bull M, Chapman B, et al. Twelve week storage trial of microbial viability in lyophilized and frozen fecal microbiota preparations. *Gastroenterology*. 2015;1(PG-S962):S962.
- Long Miao C, Mowery A, Khara H, Shellenberger M, Komar M. C. difficile enteritis after total proctocolectomy successfully treated with fecal transplant. *Am J Gastroenterol*. 2014;109(PG-S442):S442.
- Mandalia A, Ward A, Kraft CS, Dhere TA. Outcomes for route and immunocompromised status do not significantly differ in fecal microbiota transplant for recurrent clostridium difficile. *Gastroenterology*. 2014;1(PG-S252-S253):S252–3.
- Martin D, Munoz R, Yoder K, Allegretti JR, Smith M, Kassam Z. Assessing the landscape of fecal microbiota transplantation programs for recurrent clostridium difficile infection: A survey of existing practices among healthcare centers using an international public stool bank. *Gastroenterology*. 2016;1(PG-S238):S238.
- Mehta SR, Kelly CR, Kao D, Dimitry J, Martin T, Allegretti JR, et al. Inpatient status, severe clostridium difficile infection and immunocompromised state predict failure despite multiple fecal microbiota transplants: A multicenter study. *Gastroenterology*. 2016;1(PG-S745-S746):S745–6.
- Meighani A, Ramesh M, Salgia R. Successful outcomes of fecal microbiota transplantation in patients with chronic liver disease. *Hepatology*. 2016;63 (1 Supplement 1)(PG-1016A-1017A):1016A–1017A.
- Mellow M, Kanatzar A, Brandt L, Aroniadis O, Kelly C, Park T, et al. Longterm follow-up of colonoscopic Fecal Microbiota Transplant (FMT) for recurrent C. difficile infection (RCDI). *Am J Gastroenterol*. 2011;106(PG-S149-S150):S149–50.
- Mellow M, Kanatzar A. Colonoscopic fecal bacteriotherapy in the treatment of recurrent clostridium difficile infection-results and follow-up. *Am J Gastroenterol*. 2010;105(PG-S135):S135.
- Mellow M, Kohli V, Jalil S, Jabbour N. Persistent clostridium difficile infection in a patient with decompensated liver disease: “double transplant” saves a life! *Am J Gastroenterol*. 2012;107(PG-S461):S461.
- Mendelson AH, Rifkin S, Shay J, Razvi MA, Lee LA. *Gastroenterology*. 2017;152(5 Supplement 1):S949-S950.
- Mikamo H. Treatment for Clostridium difficile infections. *Int J Antimicrob Agents*. 2013;42(PG-S16):S16.
- Miller CB, Dellon E, Isaacs K, Gangarosa L. Fecal bacteriotherapy via colonoscopy as rescue therapy for refractory and recurrent clostridium difficile - Associated diarrhea. *Am J Gastroenterol*. 2010;105(PG-S323):S323.

- Mintz M, Monzur F, Chowdhury T, Rowehl L, Grewal S, Li E, et al. Comparing fecal microbial transplant outcomes in patients with recurrent clostridium difficile or ulcerative colitis. *Inflamm Bowel Dis*. 2016;22(PG-S31):S31.
- Misra B, Ramesh M, Sobcinski MK. Evaluation of health-related quality of life in patients treated with RBX2660 (Microbiota Suspension) for Recurrent C. Difficile Infection. *Am J Gastroenterol*. 2014;109(PG-S188):S188.
- Mitchell SW, Jaworski A, Bull M, Wong C, Furnari V, Chapman B, et al. Lyophilized fecal microbiota transplantation and cryoprotectants for viable bacteria preservation. *Gastroenterology*. 2016;1(PG-S542-S543):S542–3.
- Mittal C, John A, Hart BR, Miller N, Meighani A, Ramesh M. Fecal microbiota transplantation for recurrent and/or refractory clostridium difficile infection: A large retrospective study of failure rates, predictors of failure and outcomes. *Gastroenterology*. 2015;1(PG-S723-S724):S723–4.
- Monzur F, Mintz M, Tian X, Grewal S, Khair S, Rowehl L, et al. Microbiome analysis and fecal microbiota transplant outcomes in clostridium difficile and ulcerative colitis patients. *Am J Gastroenterol*. 2016;111(PG-S321):S321.
- Newton D, Hewlett A. Fecal biotherapy as treatment for recurrent clostridium difficile infection in immunocompromised patients. *Am J Gastroenterol*. 2013;108(PG-S178):S178.
- Niccum BA, Stein DJ, Wang P, Cohn SM, Hays RA. Zinc deficiency: A possible contributor to long-term FMT failure in recurrent clostridium difficile infection. *Am J Gastroenterol*. 2016;111(PG-S92):S92.
- Norin E. Experience with cultivated microbiota transplant: ongoing treatment of Clostridium difficile patients in Sweden. *Microb Ecol Heal Dis*. 2015;26(PG-27638):27638.
- O'Brien K, Osman M, Eysenbach L, Stoltzner Z, Day R, Norgaard KS, et al. Clinical efficacy of fecal microbiota transplantation for recurrent clostridium difficile infection from an international public stool bank: Results from a 1,406 patient multi-center cohort. *Gastroenterology*. 2016;1(PG-S539-S540):S539–40.
- O'Brien K, Petimar J, Ling K, Omas Gurry T, Ladha A, Day R, et al. Nutritional composition of stool donors' diets relative to that of the U.S. population: Results from 44 donors from an international stool bank for fecal microbiota transplantation. *Am J Gastroenterol*. 2016;111(PG-S447-S448):S447–8.
- Olefson SH, Jackson M, Kelly C. Clostridium difficile: The spectrum of diagnoses in patients referred for fecal microbiota transplant. *Gastroenterology*. 2015;1(PG-S727):S727.
- Oprita R, Kostov A, Musat F. Clostridium difficile-associated diarrhea, a new challenge. *Eur J Intern Med*. 2013;24(PG-e73):e73.
- Ordway S, Harris N, Wong R. Skinning the cat twice: Refractory CDI in an solid organ transplant patient requiring 2 fecal microbiota transplants. *Am J Gastroenterol*. 2015;110(PG-S171):S171.
- Osman M, Dubois N, Gangireddy V, Amaratunga K, Allegretti JR, Kassam Z. The great mimic: Food-borne illness masquerading as an infectious adverse event following fecal microbiota transplantation. *Am J Gastroenterol*. 2016;111(PG-S592):S592.

- Osman M, Khoiri A, Stoltzner Z, Koelsch E, O'Brien K, Ling K, et al. Clinical effectiveness and safety of fecal microbiota transplantation in children for clostridium difficile infection: Results from 9 pediatric centers in the United States 2016 ACG presidential poster award. *Am J Gastroenterol*. 2016;111(PG-S452):S452.
- Ott SJ, Waetzig GH, Rehman A, Moltzau-Anderson J, Bharti R, Grasis JA, et al. Efficacy of Sterile Fecal Filtrate Transfer for Treating Patients With Clostridium difficile Infection. *Gastroenterology*. 2016;17(PG-17):17.
- Paramsothy S, Borody T, Lin E, Finlayson S, Walsh A, Samuel D, et al. Obstacles to donor recruitment for faecal microbiota transplantation: Experiences from the focus study. *Am J Gastroenterol*. 2014;109(PG-S188):S188.
- Paramsothy S, Borody T, Lin E, Finlayson S, Walsh A, Samuel D, et al. Obstacles to donor recruitment for faecal microbiota transplantation-Experiences from the FOCUS study. *J Gastroenterol Hepatol*. 2014;29(PG-135):135.
- Parekh R, Ramesh MS, Tang J. Lymphocytic colitis in patients with recurrent clostridium difficile colitis: Case series. *Am J Gastroenterol*. 2016;111(PG-S1308):S1308.
- Park L, Tzimas D, Price J, Mone A, Hirsh J, Poles M, et al. Perceptions of fecal microbiota transplantation: Factors that predict acceptance: A preliminary analysis. *Am J Gastroenterol*. 2014;109(PG-S206):S206.
- Patel LN, Schairer J, Shen B. Fecal transplantation therapy for Clostridium difficile-associated pouchitis. *Int J Colorectal Dis*. 2014;29(2 PG-263-264):263–4.
- Patel LN, Schairer J, Shen B. Fecal transplantation therapy for Clostridium difficile-associated pouchitis. *Int J Colorectal Dis*. 2014;29(2 PG-263-264):263–4.
- Patel P, Goyal A. Comparative analysis of the efficacy of fecal transplantation in pediatric inflammatory bowel disease patients with and without clostridium difficile infection. *Inflamm Bowel Dis*. 2016;22(PG-S68-S69):S68–9.
- Patel P, Goyal A. Comparative analysis of the efficacy of fecal transplantation in pediatric inflammatory bowel disease patients with and without clostridium difficile infection. *Inflamm Bowel Dis*. 2016;22(PG-S68-S69):S68–9.
- Patel S, Kelly C, Colombel JF, Atreja A. Comparative cost analysis of fecal microbiota transplant and antibiotic treatment for recurrent clostridium difficile infection. *Am J Gastroenterol*. 2013;108(PG-S169-S170):S169–70.
- Patel S, Kelly C, Colombel JF, Atreja A. Comparative cost analysis of fecal microbiota transplant and antibiotic treatment for recurrent clostridium difficile infection. *Am J Gastroenterol*. 2013;108(PG-S169-S170):S169–70.
- Patel SS, Grinspan A, Colombel JF, Atreja A. Cost effectiveness analysis of fecal microbiota transplant and antibiotic treatment for recurrent clostridium difficile infection. *Gastroenterology*. 2014;1(PG-S190-S191):S190–1.

- Patel SS, Grinspan A, Colombel JF, Atreja A. Cost effectiveness analysis of fecal microbiota transplant and antibiotic treatment for recurrent clostridium difficile infection. *Gastroenterology*. 2014;1(PG-S190-S191):S190–1.
- Pinn D, Aroniadis O, Brandt L. Follow-up study of fecal microbiota transplantation (FMT) for the treatment of refractory irritable bowel syndrome (IBS). *Am J Gastroenterol*. 2013;108(PG-S563):S563.
- Potakamuri L, Turnbough L, Maheshwari A, Kantsevov S, Ofosu A, Thuluvath P, et al. Effectiveness of fecal microbiota transplantation for the treatment of recurrent clostridium difficile infection: Community hospital experience. *Am J Gastroenterol*. 2013;108(PG-S175):S175.
- Pyo-Twist A, Brumbaugh D, Fidanza SJ, Montero C, Dolan S, Hughes S, et al. Preliminary outcomes of a registered nurse driven fecal microbiota transplantation (FMT) procedure to treat clostridium difficile (c.diff) infection in pediatrics. *J Pediatr Gastroenterol Nutr*. 2016;63(PG-S154):S154.
- Quraishi MN, McCune V, Iqbal TH, Pathmakanthan S, Struthers JK, Moran E, et al. Faecal microbiota transplantation via nasogastric route for the treatment of recurrent and antibiotic refractory Clostridium Difficile infection: The UK experience. *J Crohn's Colitis*. 2015;9(PG-S323-S324):S323–4.
- Quraishi MN, McCune VL, Iqbal T, Pathmakanthan S, Struthers JK, Moran E, et al. Faecal microbiota transplantation via nasogastric route for the treatment of recurrent and antibiotic refractory clostridium difficile infection: The UK experience. *Gastroenterology*. 2015;1(PG-S641-S642):S641–2.
- Quraishi MN, Segal J, Mullish B, McCune V, Hawkey P, Colville A, et al. National survey of practice of faecal microbiota transplantation for clostridium difficile infection in the United Kingdom. *Gut*. 2016;65(PG-A23-A24):A23–4.
- Quraishi N, McMillan M, Widlak M, Nell L, Quraishi K, Pathmakanthan S, et al. Patient perception towards faecal microbiota transplantation for treatment of inflammatory bowel disease. *United Eur Gastroenterol J*. 2014;1(PG-A383):A383.
- Quraishi N, McMillan M, Widlak M, Nell L, Quraishi K, Pathmakanthan S, et al. Patient perception towards faecal microbiota transplantation for treatment of Inflammatory Bowel Disease. *J Crohn's Colitis*. 2015;9(PG-S252):S252.
- Ramesh M, Misra B, Ray A, Smith R, Sobcinski MK. RBX2660 (Microbiota Suspension) for Recurrent C. Difficile infection: 60-day interim analysis of the PUNCH CD phase 2 safety study. *Am J Gastroenterol*. 2014;109(PG-S188):S188.
- Ray A, Hardi R, Ramesh M, Sobcinski MK. Enema administration of RBX2660 (microbiota suspension) for Recurrent C. difficile infection: Lessons learned from the PUNCH CD Study. *Am J Gastroenterol*. 2014;109(PG-S192-S193):S192–3.
- Razik R, Osman M, Lieberman A, Dubois N, Allegretti JR, Smith M, et al. Characterizing patients who fail fecal microbiota transplantation for clostridium difficile infection: Results from a 135-patient, multi-center, non-responder cohort. *Am J Gastroenterol*. 2016;111(PG-S66):S66.

- Rezk AN, Stewart D, West S, Miao C, Khara HS, Komar M. Outcomes, safety and predictors of failure of fecal microbiota transplantation for refractory clostridium difficile infection. *Am J Gastroenterol*. 2016;111(PG-S82-S83):S82–3.
- Roediger R, Grinspan A. Safety and efficacy of fecal microbiota transplantation for clostridium difficile in a cohort of patients with a severe infection and/or IBD. *Am J Gastroenterol*. 2016;111(PG-S446):S446.
- Sadowsky MJ, Weingarden A, Khoruts A, Gonzalez A, Vazquez-Baeza Y, Weiss S, et al. Short and long term changes in bacterial composition following fecal microbiota transplantation for CDI visualized in movie format. *Gastroenterology*. 2014;1(PG-S-838):S-838.
- Sbahi H, Di Palma JA. Faecal microbiota transplantation: applications and limitations in treating gastrointestinal disorders. *BMJ Open Gastroenterol*. 2016;3(1 PG-e000087):e000087.
- Scaldaferri F, Pecere S, Bruno G, Ianiro G, Laterza L, Gerardi V, et al. An Open-label, pilot study to assess feasibility and safety of fecal microbiota transplantation in patients with mild-moderate ulcerative colitis: Preliminary results. *J Crohn's Colitis*. 2015;9(PG-S278):S278.
- Scaldaferri F, Pecere S, Bruno G, Ianiro G, Laterza L, Gerardi V, et al. An open-label, pilot study to assess feasibility and safety of fecal microbiota transplantation in patients with mild-moderate ulcerative colitis: Preliminary results. *Gastroenterology*. 2015;1(PG-S870):S870.
- Scaldaferri F, Pecere S, Lopetuso LR, Ianiro G, Laterza L, Schiavoni E, et al. An open-label, pilot study to assess feasibility and safety of fecal microbiota transplantation in patients with mild-moderate ulcerative colitis: Preliminary results. *United Eur Gastroenterol J*. 2015;1(PG-A257):A257.
- Shah R, Robinson L, Herrera HR, Swaroop PP. Human probiotic infusion (HPI) in ulcerative colitis-'patient's perceptions and predictors of efficacy'. *Gastroenterology*. 2012;1(PG-S253):S253.
- Shogbesan O, Poudel D, Jehangir A, Fadahunsi O, Shogbesan G, Donato A. Fecal microbiota transplantation for clostridium difficile infections in immunocompromised patients: A systematic review. *Am J Gastroenterol*. 2016;111(PG-S79):S79.
- Singh T, Yu S, Gangireddy V, Rao S. Diarrhea after fecal microbiota transplantation and usefulness of commercial stool donor for C. difficile infection. *Am J Gastroenterol*. 2015;110(PG-S589-S590):S589–90.
- Staley C, Kelly CR, Brandt LJ, Khoruts A, Sadowsky MJ. Characterization of fecal microbiota in response to heterologous versus autologous (placebo) fecal microbial transplantation: Results from a dual-center, randomized, placebo-controlled trial. *Gastroenterology*. 2016;1(PG-S542):S542.
- Swanson S, Herman M, Vindigni S, Broussard E. Application of a predictive model for early failure of FMT (fecal microbiota transplant). *Am J Gastroenterol*. 2016;111(PG-S82):S82.
- Tafesh Z, O'Neil S, Crawford Jr C V. Fecal microbiota transfer as rescue therapy: Is there a role in severe C. difficile infection? *Am J Gastroenterol*. 2015;110(PG-S185):S185.
- Tariq R, Weatherly RM, Kammer PP, Pardi D, Khanna S. Experience and outcomes from a specialized clostridium difficile clinical practice. *Gastroenterology*. 2016;1(PG-S746-S747):S746–7.

- Van Beurden YH, De Groot PF, Van Nood E, Nieuwdorp M, Keller JJ, Goorhuis A. Complications and long term follow-up of fecal microbiota transplantation for treatment of recurrent clostridium difficile infection. *Gastroenterology*. 2016;1(PG-S544):S544.
- Vermeire S, Joossens M, Verbeke K, Hildebrand F, Machiels K, Van den Broeck K, et al. Pilot Study on the Safety and Efficacy of Faecal Microbiota Transplantation in Refractory Crohn's Disease. *Gastroenterology*. 2012;142(5):S-360.
- Vitek P, Zela O, Mikoviny Kajrlikova I, Kuchar J, Chalupa J. Age is the main risk factor of mortality among patients with clostridium difficile infection. *United Eur Gastroenterol J*. 2014;1(PG-A543):A543.
- Wang PT, Fashandi AZ, Martin AN, Hays RA. Comparing fecal microbiota transplantation to total abdominal colectomy and loop ileostomy in severe and complicated clostridium difficile infections. *Am J Gastroenterol*. 2016;111(PG-S94-S95):S94-5.
- Wang PT, Schall SE, Doran AE, Tuskey AG, Hays RA. Healthy pregnancy in a newly diagnosed crohn's patient treated with fecal microbiota transplant for recurrent clostridium difficile infections. *Am J Gastroenterol*. 2016;111(PG-S595-S596):S595-6.
- Wang Y, Shen B. Fecal microbiota transplantation in refractory clostridium difficile pouchitis. *Inflamm Bowel Dis*. 2016;22(PG-S11-S12):S11-2.
- Watson JB, Habr F, Kelly C. First reported complication of fecal microbiota transplant: Ulcerative colitis flare after FMT for relapsing clostridium difficile infection. *Gastroenterology*. 2012;1(PG-S540):S540.
- Weingarden A, Hamilton MJ, Sadowsky MJ, Khoruts A. Changes in bacterial composition following fecal microbiota transplantation for severe clostridium difficile infection. *Gastroenterology*. 2013;1(PG-S829):S829.
- Wieczorek T, Macholz M, Bethge A, Neumann F, Schreiter K, Lindner M, et al. Fecal microbiome therapy in relapsing clostridium difficile infection-long-term results. *Int J Infect Dis*. 2016;45(PG-347):347.
- Wilcox GM. Early experience with a Fecal Bacteriotherapy (FB) program for recurrent and c-difficile infection (CDI). *Gastroenterology*. 2011;1(PG-S361):S361.
- Zhou E, Kumar V, Mansoor MS, Feuerstadt P. Pseudomembranes are infrequently seen in patients undergoing fecal microbiota transplant (FMT) for recurrent C. Difficile infection (CDI). *Am J Gastroenterol*. 2016;111(PG-S68):S68.
- Zhu J, Roach B, Kao D. Successful eradication of recurrent clostridium difficile infection (rCDI) of small bowel with frozen encapsulated fecal microbiota transplantation (FMT) in a patient with crohn's disease and ileostomy. *Can J Gastroenterol Hepatol Conf*. 2016;(pagination PG-)

### **D.1.3. Case series not fulfilling selection criteria**

- Alhmoud T, Gavin M. An unusual complication after a fecal microbiota transplant via colonoscopy. *Am J Gastroenterol*. 2014;109(PG-S424):S424.



- Allegretti JR, Day R, Kassam Z, Smith M. Empiric treatment of suspected recurrent clostridium difficile infection with vancomycin may interfere with evaluation for fecal microbiota transplantation. *Am J Gastroenterol*. 2016;111(PG-S87):S87.
- Allegretti JR, Hamilton MJ, Korzenik JR, Chan WW. Factors associated with C. Difficile negative gastrointestinal symptoms after intestinal microbiome restoration. *Gastroenterology*. 2015;1(PG-S643):S643.
- Allegretti JR, Korzenik JR, Hamilton MJ. Intestinal microbiome restoration for recurrent clostridium difficile infection in patients with concurrent inflammatory bowel disease. *Gastroenterology*. 2015;1(PG-S869):S869.
- Allegretti JR, Phelps E, Xu H, Kassam Z, Fischer M. Redefining cure in clostridium difficile infection: Clinical assessment 4 weeks after fecal microbiota transplantation is predictive of standard 8-week cure endpoint. *Am J Gastroenterol*. 2016;111(PG-S56):S56.
- Allegretti JR, Storm M, Smith M, Kelly CR, Kearney S, Perrotta A, et al. Strain-level analysis of microbial engraftment associated with resolution of recurrent clostridium difficile following fecal microbiota transplantation. *Gastroenterology*. 2016;1(PG-S540-S541):S540–1.
- Anand R, Sinha A, Sivaraman A, Hasan S, Garg S, Dutta S. Quality of life index in patients with recurrent clostridium difficile colitis following fecal microbiota transplantation: Long-term outcome. *Am J Gastroenterol*. 2015;110(PG-S568):S568.
- Andrews J, Costello S. The emerging role of faecal microbiota transplantation. *Med Today*. 2014;15(7 PG-62-64):62–4.
- Angelberger S, Reinisch W, Makristathis A, Lichtenberger C, Dejaco C, Papay P, et al. Temporal bacterial community dynamics vary among ulcerative colitis patients after fecal microbiota transplantation. *Am J Gastroenterol*. 2013;108(10 PG-1620-1630):1620–30.
- Angelberger S, Reinisch W, Makristathis A, Lichtenberger C, Dejaco C, Papay P, et al. Temporal bacterial community dynamics vary among ulcerative colitis patients after faecal microbiota transplantation. *J Crohn's Colitis*. 2013;7(PG-S291):S291.
- Arkkila P, Mattila E, Anttila VJ. [Fecal transfusion as treatment of Clostridium difficile infection]. [Finnish]. *Duodecim*. 2013;129(16 PG-1671-1679):1671–9.
- Arkkila P, Mattila E, Anttila VJ. [Fecal transfusion as treatment of Clostridium difficile infection]. *Duodecim*. 2013;129(16 PG-1671-9):1671–9.
- Asonuma K, Kuroki Y, Ino S, Hanamura S, Takano Y, Yamamura E, et al. Severe refractory Clostridium difficile infection with good response to fecal microbiota transplantation: A case report. [Japanese]. *J Japanese Soc Gastroenterol*. 2016;113(1 PG-55-62):55–62.
- Atkins KA, Kao D. Potential cost savings associated with timely fecal microbiota transplantation (FMT) for recurrent clostridium difficile infection (RCDI). *Gastroenterology*. 2014;1(PG-S-252):S-252.

- Balzola F, Cullen G, Hoentjen F, Ho GT, Russell R. Safety, tolerability, and clinical response after fecal transplantation in children and young adults with ulcerative colitis. *Inflamm Bowel Dis Monit*. 2013;13(4 PG-167):167.
- Bansal S, Serban R, Kemal N, Casey K, Dunnigan K, Kurchin A. Fecal microbiota transplant for recurrent clostridium difficile infection at a teaching hospital in upstate New York: Our experience. *Am J Gastroenterol*. 2013;108(PG-S383-S384):S383–4.
- Bartlett M, Alsafadi A. The outcomes of using fresh parental stool versus frozen anonymous-donor stool in pediatric fecal microbiota transplant. *J Pediatr Gastroenterol Nutr*. 2016;63(PG-S314):S314.
- Borody TJ, Mitchell SW, Wong C, Jaworski A. Encapsulated lyophilized fecal microbiota therapy for the treatment of clostridium difficile infection. *Am J Gastroenterol*. 2016;111(PG-S409):S409.
- Borody TJ, Warren EF, Leis S, Surace R, Ashman O. Treatment of ulcerative colitis using fecal bacteriotherapy. *J Clin Gastroenterol*. 2003;37(1):42–7.
- Brandt L, Aroniadis O. Long-term follow-up study of fecal microbiota transplantation (FMT) for ulcerative colitis (UC). *Am J Gastroenterol*. 2012;107(PG-S657):S657.
- Buers M, Quatrara B, Niccum B, Vance S, Hays RA. All-cause hospital admissions decreased after fecal microbiota transplantation for recurrent clostridium difficile infection. *Am J Gastroenterol*. 2016;111(PG-S91-S92):S91–2.
- Burns LJ, Dubois N, Smith MB, Mendolia GM, Burgess J, Edelstein C, et al. Donor recruitment and eligibility for fecal microbiota transplantation: Results from an international public stool bank. *Gastroenterology*. 2015;1(PG-S96-S97):S96–7.
- Cammarota G, Ianiro G, Gasbarrini A, Masucci L, Sanguinetti M. Faecal transplantation for Clostridium difficile infection. Three cases treated in Italy. *Dig Liver Dis*. 2014;46(5 PG-475):475.
- Cheng YW, Xu H, Rogers N, Sagi S, Bohm M, Fischer M. Sequential fecal microbiota transplant protocol: A promising alternative to colectomy in severe and severe/complicated clostridium difficile. *Am J Gastroenterol*. 2016;111(PG-S59-S60):S59–60.
- Cherem JH, Ulloa IH. Home fecal transplantation in elderly women. *Gac Med Mex*. 2014;150(1 PG-106-107):106–7.
- Chetan M, Benjamin H, Ajin J, Alireza M, Nichole M, Mayur R. Fecal transplant for recurrent and/or refractory clostridium difficile infection in patients with inflammatory bowel disease. *Inflamm Bowel Dis*. 2014;20(PG-S72):S72.
- Costello S, La Brooy J, Tucker E, Holloway R, Schoeman M, Andrews JM. Establishment of a fecal microbiota transplant service for the treatment of recurrent Clostridium difficile colitis in the Australian public hospital setting. *J Gastroenterol Hepatol*. 2014;29(PG-134):134.
- Damman C, Brittnacher M, Hayden H, Radey M, Hager K, Miller S, et al. Single colonoscopically administered fecal microbiota transplant for ulcerative colitis-a pilot study to determine therapeutic benefit and graft stability. *Gastroenterology*. 2014;1(PG-S-460):S-460.



- Dimitry J, Keshteli A, Kao D. Independent predictors of failure of fecal microbiota transplantation (FMT) for recurrent or refractory clostridium difficile infection. *Can J Gastroenterol Hepatol Conf*. 2016;(pagination PG-).
- Dimitry J, Keshteli AH, Kao D. Su1746 Predictors of Failure of Fecal Microbiota Transplantation (FMT) in the Management of Recurrent Clostridium difficile Infection. *Gastroenterology*. 2016 Apr;150(4):S543.
- Doran A, Vance S, Warren C, Kolling G, Chaplain A, Archbald-Pannone L, et al. Microscopic colitis in recurrent C. difficile infection may resolve spontaneously after fmt. *Am J Gastroenterol*. 2015;110(PG-S584):S584.
- El-Halabi M, Cheng YW, Rogers N, Sagi S, Bohm M, Xu H, et al. Changes in mortality, colectomy, and length of hospital stay after implementation of inpatient fecal microbiota transplantation program for severe clostridium difficile infection. *Am J Gastroenterol*. 2016;111(PG-S67):S67.
- Elliott R. Stool transplant for recurrent clostridium difficile infection using designated screened donors in a community hospital. *Am J Gastroenterol*. 2016;111(PG-S71):S71.
- Elliott RJ, Njenga M, Ladha A, Warren K, Blackler D, Stoltzner Z, et al. Stool processing speed and storage duration do not impact clinical effectiveness of fecal microbiota transplantation across 1,924 clostridium difficile infection patients. *Am J Gastroenterol*. 2016;111(PG-S57):S57.
- Emanuelsson F, Claesson BEB, Ljungström L, Tvede M, Ung K-A. Faecal microbiota transplantation and bacteriotherapy for recurrent Clostridium difficile infection: A retrospective evaluation of 31 patients. *Scand J Infect Dis*. 2014 Feb 20;46(2):89–97.
- Falconer S, Moss E, Andermann T, Systrom H, Mahabamunuge J, Hohmann E, et al. Fecal microbiota transplant is a potentially safe and effective treatment for clostridium difficile infection in hematopoietic stem cell recipients. *Biol Blood Marrow Transplant*. 2016;1(PG-S53-S54):S53–4.
- Fischer M, Bittar M, Papa E, Kassam Z, Smith M. Can you cause IBD with fecal transplantation? 31-patient case series of fecal transplantation using stool from a donor who later developed Crohn's disease. *Am J Gastroenterol*. 2016;111(PG-S294-S295):S294–5.
- Fischer M, Cook G, Rogers N, Sipe B, Vuppalachchi R. Rescue therapy with fecal microbiota transplantation in hospitalized patients with severe and severe-complicated clostridium difficile infection. *Am J Gastroenterol*. 2014;109(PG-S195):S195.
- Fischer M, Kelly C, Kao D, Kuchipudi A, Jafri SM, Blumenkehl M, et al. Outcomes of fecal microbiota transplantation for C. Difficile infection in patients with inflammatory bowel disease. *Am J Gastroenterol*. 2014;109(PG-S487):S487.
- Fischer M, Mehta S, Martin T, Cook G, Phelps E, Sipe B, et al. Predictors of failure after fecal microbiota transplantation (FMT) for the therapy of clostridium difficile infection (CDI). *Am J Gastroenterol*. 2015;110(PG-S582):S582.
- Fischer M, Phelps E, Bolla R, Storm M, Allegretti JR. Long-term risk of clostridium difficile infection recurrence with or without antibiotic exposure following successful fecal microbiota transplant. *Gastroenterology*. 2016;1(PG-S23):S23.

- Fischer M, Rex DK, Cook GK. Fecal microbiota transplantation for recurrent clostridium difficile in patients with prolonged immunosuppression. *United Eur Gastroenterol J*. 2013;1(PG-A380):A380.
- Frank J, Hogenauer C, Grochenig HP, Hoffmann KM, Reicht G, Wenzl HH, et al. Safety of fecal microbiota transplantation in patients with chronic colitis and immunosuppressive treatment. *J Crohn's Colitis*. 2015;9(PG-S245-S246):S245–6.
- Freeman AE, Roberts SC, Chey WD, Kao JY, Rao K. New onset functional gi disorders following fecal microbiota transplant for recurrent clostridium difficile infection-prevalence and risk factors. *Gastroenterology*. 2016;1(PG-S495):S495.
- Gallegos-Orozco JF, Paskvan-Gawryletz CD, Gurudu SR, Orenstein R. Successful colonoscopic fecal transplant for severe acute Clostridium difficile pseudomembranous colitis. *Rev Gastroenterol Mex*. 2012;77(1 PG-40-2):40–2.
- Gupta A, Khanna S. Ipilimumab-associated colitis or refractory Clostridium difficile infection? *BMJ Case Rep*. 2015;2015 (no pagination)(A1084 PG-).
- Gweon TG, Kim J, Lim CH, Park JM, Lee DG, Lee IS, et al. Fecal Microbiota Transplantation Using Upper Gastrointestinal Tract for the Treatment of Refractory or Severe Complicated Clostridium difficile Infection in Elderly Patients in Poor Medical Condition: The First Study in an Asian Country. *Gastroenterol Res Pract*. 2016;2016 (no pagination)(2687605 PG-).
- Gweon TG, Lee KJ, Kang D, Park SS, Kim KH, Seong H, et al. A case of toxic megacolon caused by Clostridium difficile infection and treated with fecal microbiota transplantation. *Gut Liver*. 2015;9(2 PG-247-250):247–50.
- Gweon TG, Lee KJ, Kang DH, Park SS, Kim KH, Seong HJ, et al. A case of toxic megacolon caused by clostridium difficile infection and treated with fecal microbiota transplantation. *Gut Liver*. 2015;9(2 PG-247-50):247–50.
- Hourigan SK, Chen LA, Grigoryan Z, Laroche G, Weidner M, Sears CL, et al. Microbiome changes associated with sustained eradication of Clostridium difficile after single faecal microbiota transplantation in children with and without inflammatory bowel disease. *Aliment Pharmacol Ther*. 2015;42(6 PG-741-752):741–52.
- IrreGaertner W, Madoff R, Mellgren A, Kwaan M, Melton G. Impact of postoperative clostridium difficile infection after colon and rectal operations. *Color Dis*. 2014;16(PG-145):145.
- Karolewska-Bochenek K, Lazowska-Przeorek I, Banaszkiwicz A, Gawronska A, Kotowska M, Dziekiewicz M, et al. Fecal microbiota transplantation for CMV infection in pediatric patients with IBD. *J Pediatr Gastroenterol Nutr*. 2016;62(PG-147):147.
- Karolewska-Bochenek K, Lazowska-Przeorek I, Grzesiowski P, Banaszkiwicz A, Albrecht P, Gawronska A, et al. Fecal microbiota transplantation in refractory pediatric UC - Preliminary data. *J Crohn's Colitis*. 2015;9(PG-S294):S294.
- Kelly C, De Leon L. Successful treatment of recurrent clostridium difficile infection with donor stool administered at colonoscopy: A case series. *Am J Gastroenterol*. 2010;105(PG-S135):S135.

- Kump PK, Gröchenig H-P, Lackner S, Trajanoski S, Reicht G, Hoffmann KM, et al. Alteration of intestinal dysbiosis by fecal microbiota transplantation does not induce remission in patients with chronic active ulcerative colitis. *Inflamm Bowel Dis*. 2013;19(10):2155–65.
- Laszlo M, Ciobanu L, Andreica V, Pascu O. Fecal transplantation indications in ulcerative colitis. Preliminary study. *Clujul Med*. 2016;89(2 PG-224-8):224–8.
- Le L, El-Nachef N. Fecal microbiota transplantation in solid organ transplant and hematopoietic stem cell transplant patients with recurrent or refractory *Clostridium difficile* infection: A case series. *Am J Gastroenterol*. 2016;111(PG-S615):S615.
- Link A, Lachmund T, Schulz C, Weigt J, Malfertheiner P. Endoscopic peroral jejunal fecal microbiota transplantation. *Dig Liver Dis*. 2016;48(11 PG-1336-1339):1336–9.
- Lofland D, Josephat F, Partin S. Fecal transplant for recurrent *Clostridium difficile* infection. *Clin Lab Sci*. 2013;26(3 PG-131-135):131–5.
- Petrof EO, Gloor GB, Vanner SJ, Weese SJ, Carter D, Daigneault MC, et al. Stool substitute transplant therapy for the eradication of *Clostridium difficile* infection: “RePOOPulating” the gut. *Microbiome*. 2013 Jan;1(1):3.
- Pierog A, Mencin A, Reilly NR. Fecal microbiota transplantation in children with recurrent *Clostridium difficile* infection. *Pediatr Infect Dis J*. 2014;33(11 PG-1198-1200):1198–200.
- Ponte A, Pinho R, Mota M, Silva J, Vieira N, Oliveira R, et al. Initial experience with fecal microbiota transplantation in *Clostridium difficile* infection - transplant protocol and preliminary results. *Rev Esp Enfermedades Dig*. 2015;107(7 PG-402-407):402–7.
- Ray A, Jones C. Does the donor matter? Donor vs patient effects in the outcome of a next-generation microbiota-based drug trial for recurrent *Clostridium difficile* infection. *Future Microbiol*. 2016;11(PG-611-6):611–6.
- Rebello D, Yen E, Lio P, Kelly CR. Unexpected benefits: Hair growth in two alopecia patients after fecal microbiota transplant. *Am J Gastroenterol*. 2016;111(PG-S623-S624):S623–4.
- Satokari R, Mattila E, Kainulainen V, Arkkila PE. Simple faecal preparation and efficacy of frozen inoculum in faecal microbiota transplantation for recurrent *Clostridium difficile* infection - an observational cohort study. *Aliment Pharmacol Ther*. 2015;41(1):46–53.
- Schulz-Stubner S, Textor Z, Anetseder M. Fecal Microbiota Therapy as Rescue Therapy for Life-Threatening *Clostridium difficile* Infection in the Critically Ill: A Small Case Series. *Infect Control Hosp Epidemiol*. 2016;37(9 PG-1129-1131):1129–31.
- Silverman MS, Davis I, Pillai DR. Success of Self-Administered Home Fecal Transplantation for Chronic *Clostridium difficile* Infection. *Clin Gastroenterol Hepatol*. 2010;8(5 PG-471-473):471–3.
- Veling N. A novel approach in the treatment of *Clostridium difficile*: A case study. *J Spinal Cord Med*. 2014;37(4)(PG-441-442):441–2.

Vigvari S, Nemes Z, Vincze A, Solt J, Sipos D, Feiszt Z, et al. Faecal microbiota transplantation in Clostridium difficile infections. *Infect Dis (Auckl)*. 2015;47(2 PG-114-116):114–6.

Webb BJ, Brunner A, Ford CD, Gazdik MA, Petersen FB, Hoda D. Fecal microbiota transplantation for recurrent Clostridium difficile infection in hematopoietic stem cell transplant recipients. *Transpl Infect Dis*. 2016;18(4 PG-628-633):628–33.

Weidner M, Hourigan S, Ling K, O'Brien K, Oliva-Hemker M. Fecal microbiota transplantation using banked donor stool is effective in the treatment of recurrent clostridium difficile infection in children. *J Pediatr Gastroenterol Nutr*. 2016;63(PG-S143-S144):S143–4.

#### **D.1.4. Case reports**

Abeyesundere RL. A ward outbreak of Clostridium difficile enterocolitis. *J Infect*. 1982;5(3 PG-277-282):277–82.

Adamski JK, Jaschke BB, Uusitalo-Seppala RS, Moilanen KJV, Pehkonen AV, Weigl W. Routine Treatment-Resistant Clostridium difficile Infection during Recovery from Myxedema. *Case Reports in Gastroenterology*. 2017:740-6.

Agrawal M, Aroniadis OC, Brandt LJ, Kelly C, Freeman S, Surawicz C, et al. A long-term follow-up study of the efficacy and safety of fecal microbiota transplant (FMT) for Recurrent/Severe/Complicated C. Difficile Infection (CDI) in the elderly. *Gastroenterology*. 2014;1(PG-S42-S43):S42–3.

Al-Nassir WN, Sethi AK, Li Y, Pultz MJ, Riggs MM, Donskey CJ. Both oral metronidazole and oral vancomycin promote persistent overgrowth of vancomycin-resistant enterococci during treatment of Clostridium difficile-associated disease. *Antimicrob Agents Chemother*. 2008;52(7 PG-2403-2406):2403–6.

Alang N, Kelly CR. Weight gain after fecal microbiota transplantation. *Open Forum Infect Dis*. 2015;2 (1) (no pagination)(ofv004 PG-).

Alonso CD, Kamboj M. Clostridium difficile Infection (CDI) in solid organ and hematopoietic stem cell transplant recipients. *Curr Infect Dis Rep*. 2014;16 (8) (no pagination)(414 PG-).

Alrabaa S, Noel PR, Wills T. Clostridium difficile infection: What you need to know. *Consultant*. 2013;53(6 PG-389-395):389–95.

Alsakka M, Sharabash N, Alktaifi A, Salih M, German M. Successful fecal microbiota transplantation (FMT) for recurrent clostridium difficile infection (CDI) after subtotal colectomy. *Am J Gastroenterol*. 2013;108(PG-S365-S366):S365–6.

Anand R, Song Y, Sinha A, Hasan S, Sivaraman A, Garg S, et al. Effect of aging on the fecal microbiome in healthy donors for fecal microbiota transplant. *Gastroenterology*. 2015;1(PG-S719):S719.

Andrews R, Gavin M. Post-infectious IBS following recurrent/ relapsing C. difficile associated diarrhea (CDAD). *J Investig Med*. 2016;64 (1)(PG-244):244.

- Ang P, Cheong WK, Khoo KS. Pseudomembranous colitis in a patient treated with paclitaxel for carcinoma of the breast: A case report. *Ann Acad Med Singapore*. 2000;29(1 PG-132-134):132–4.
- Aratari A, Cammarota G, Papi C. Fecal microbiota transplantation for recurrent *C. difficile* infection in a patient with chronic refractory ulcerative colitis. *J Crohns Colitis*. 2015;9(4 PG-367):367.
- Asonuma K, Kuroki Y, Ino S, Hanamura S, Takano Y, Yamamura E, et al. Severe refractory *Clostridium difficile* infection with good response to fecal microbiota transplantation: a case report. *Nippon Shokakibyo Gakkai Zasshi - Japanese J Gastroenterol*. 2016;113(1 PG-55-62):55–62.
- Bamba S, Nishida A, Imaeda H, Inatomi O, Sasaki M, Sugimoto M, et al. Successful treatment by fecal microbiota transplantation for Japanese patients with refractory *Clostridium difficile* infection: A prospective case series. *Journal of Microbiology, Immunology and Infection*. 2017.
- Bartosz C, Marino D, DeCross A. A highly illustrative case report detailing the profound subjective and objective response of severe pseudomembranous colitis (from *clostridium difficile*) to fecal transplant. *Am J Gastroenterol*. 2016;111(PG-S620):S620.
- Bartosz C, Marino D, DeCross A. A highly illustrative case report detailing the profound subjective and objective response of severe pseudomembranous colitis (from *clostridium difficile*) to fecal transplant. *Am J Gastroenterol*. 2016;111(PG-S620):S620.
- Berro ZZ, Hamdan RH, Dandache IH, Saab MN, Karnib HH, Younes MH. Fecal microbiota transplantation for severe *clostridium difficile* infection after left ventricular assist device implantation: A case control study and concise review on the local and regional therapies. *BMC Infect Dis*. 2016;16 (1) (no pagination)(234 PG-).
- Binkovitz LA, Allen E, Bloom D, Long F, Hammond S, Buonomo C, et al. Atypical presentation of *Clostridium difficile* colitis in patients with cystic fibrosis. *Am J Roentgenol*. 1999;172(2 PG-517-521):517–21.
- Brechmann T, Swol J, Knop-Hammad V, Willert J, Aach M, Cruciger O, et al. Complicated fecal microbiota transplantation in a tetraplegic patient with severe *Clostridium difficile* infection. *World J Gastroenterol*. 2015;21(12 PG-3736-3740):3736–40.
- Broecker F, Klumpp J, Schuppler M, Russo G, Biedermann L, Hombach M, et al. Long-term changes of bacterial and viral compositions in the intestine of a recovered *Clostridium difficile* patient after fecal microbiota transplantation. *Cold Spring Harb Mol Case Stud*. 2016;2(1 PG-a000448):a000448.
- Broecker F, Kube M, Klumpp J, Schuppler M, Biedermann L, Hecht J, et al. Analysis of the intestinal microbiome of a recovered *clostridium difficile* patient after fecal transplantation. *Digestion*. 2013;88(4 PG-243-251):243–51.
- Cammarota G, Ianiro G, Masucci L, Pecere S, Bibbo S, Scaldaferri F, et al. Fecal microbiota transplantation for recurrent *C. difficile* infection: A 2-year experience from a European referral centre. *United Eur Gastroenterol J*. 2015;1(PG-A131):A131.
- Chang B, Rezaie A. Post-fecal microbiota transplantation (FMT) constipation and abdominal distention due to methane-predominant bacterial overgrowth contracted from the donor. *Am J Gastroenterol*. 2016;111(PG-S807-S808):S807–8.

- Chao HC, Yu WL. Treatment failure of fecal microbiota transplant for pseudomembranous colitis due to coexistent cytomegalovirus colitis. *J Microbiol Immunol Infect*. 2016;49(4 PG-617-618):617–8.
- Cho S, Spencer E, Hirten R, Grinspan A, Dubinsky M. High recurrence rate after fecal microbiota transplant for recurrent *clostridium difficile* infection in pediatric inflammatory bowel disease. *Journal of Pediatric Gastroenterology and Nutrition*. 2017;65 (Supplement 2):S279-S80.
- Chu A, Michail S. Pediatric recurrent *c difficile* infections-a sign of undiagnosed gi disease. *Journal of Pediatric Gastroenterology and Nutrition*. 2017;65 (Supplement 2):S70.
- Colleen K, Hassan Z, Stacy K. New diagnosis of crohn's colitis 6 weeks after fecal microbiota transplantation (FMT). *Inflamm Bowel Dis*. 2014;20(PG-S21):S21.
- Collins DC. Pseudomembranous enterocolitis. Further observations on the value of donor fecal enemata as an adjunct in the treatment of pseudomembranous enterocolitis. *Am J Proctol*. 1960;2(PG-389-91):389–91.
- Costello SP, Chung A, Andrews JM, Fraser RJ. Fecal microbiota transplant for *clostridium difficile* colitis-induced toxic megacolon. *Am J Gastroenterol*. 2015;110(5 PG-775-777):775–7.
- Davidovics ZH, Vance K, Etienne N, Hyams JS. Fecal Transplantation Successfully Treats Recurrent D-Lactic Acidosis in a Child With Short Bowel Syndrome. *Jpen J Parenter Enter Nutr*. 2015;29(PG-29):29.
- De Castro CG, Ganc AJ, Ganc RL, Petrolli MS, Hamerschlack N. Fecal microbiota transplant after hematopoietic SCT: Report of a successful case. *Bone Marrow Transplant*. 2015;50(1 PG-145):145.
- Diamond C, McNeilly T. Faecal microbiota transplantation for *clostridium difficile* - a local perspective. *Ulster Medical Journal*. 2017;86(2):108-10.
- Didesch MM, Averill A, Oh-Park M. Peripheral Neuropathy After Fecal Microbiota Transplantation for *Clostridium difficile* Infection: A Case Report. *PM R*. 2016;8(8 PG-813-816):813–6.
- Didesch MM, Averill A, Oh-Park M. Peripheral neuropathy after fecal transplantation for *clostridium difficile* infection: A case report. *PM R*. 2014;1(PG-S237-S238):S237–8.
- Duke PS, Fardy J. Recurrent *Clostridium difficile* infection treated with home fecal transplantation: A case report. *J Med Case Rep*. 2014;8 (1) (no pagination)(393 PG-).
- Dumitru IM, Dumitru E, Resul G, Curtali L, Paris S, Rugina S. Concomitant CMV and *Clostridium difficile* colitis in an immunocompetent patient treated with Ganciclovir and fecal transplantation. *J Gastrointest Liver Dis*. 2014;23(2 PG-221-222):221–2.
- Duplessis CA, You D, Johnson M, Speziale A. Efficacious outcome employing fecal bacteriotherapy in severe Crohn's colitis complicated by refractory *Clostridium difficile* infection. *Infection*. 2012;40(4 PG-469-472):469–72.
- Ehlermann P, Dosch AO, Katus HA. Donor fecal transfer for recurrent *Clostridium difficile*-associated diarrhea in heart transplantation. *J Hear Lung Transplant*. 2014;33(5 PG-551-553):551–3.



- Enriquez R, Borrás-Blasco J, Sirvent AE, Padilla S, Navarro-Ruiz A, Solavera J, et al. Imipenem-induced *Clostridium difficile* diarrhea in a patient with chronic renal failure. *Saudi J Kidney Dis Transpl*. 2011;22(3 PG-541-543):541–3.
- Espinoza R, Quera R, Meyer L, Rivera D. [Fecal microbiota transplantation: first case report in Chile and review]. *Rev Chil Infectol*. 2014;31(4 PG-477-82):477–82.
- Floe A, Leutscher P. [Recurrent *Clostridium difficile* infection treated with faecal microbiota transplantation]. *Ugeskr Laeger*. 2014;176(4 PG-17):17.
- Freeman S, Mao E, Shah S, Kelly C. A case of recurrent *clostridium difficile* enteritis treated with fecal microbiota transplant. *Am J Gastroenterol*. 2014;109(PG-S328):S328.
- Garcia-Fernandez S, Morosini MI, Cobo M, Foruny JR, Lopez-Sanroman A, Cobo J, et al. Gut eradication of VIM-1 producing ST9 *Klebsiella oxytoca* after fecal microbiota transplantation for diarrhea caused by a *Clostridium difficile* hypervirulent R027 strain. *Diagn Microbiol Infect Dis*. 2016;86(4 PG-470-471):470–1.
- Garg S, Walia R, Girotra M, Gjokopulli A, Mirza Y, Cuffari C, et al. A novel treatment for recurrent *clostridium difficile* infection in a 20-month-old. *Am J Gastroenterol*. 2012;107(PG-S556):S556.
- Gathe JC, Diejomaoh EM, Mayberry CC, Clemmons JB. Fecal Transplantation for *Clostridium Difficile* - “all Stool May Not Be Created Equal.” *J Int Assoc Provid AIDS Care*. 2016;15(2 PG-107-108):107–8.
- Goeser F, Schlabe S, Ruiner CE, Kramer L, Strassburg CP, Spengler U. Non-invasive fecal microbiota transplantation for recurrent *Clostridium difficile* infection in a patient presenting with hypertensive disorder post interventionem. *Z Gastroenterol*. 2016;54(10 PG-1143-1146):1143–6.
- Gundling F, Tiller M, Agha A, Schepp W, Iesalnieks I. Successful autologous fecal transplantation for chronic diversion colitis. *Tech Coloproctol*. 2015;19(1 PG-51-52):51–2.
- Jang MO, An JH, Jung SI, Park KH. Refractory *Clostridium difficile* Infection Cured With Fecal Microbiota Transplantation in Vancomycin-Resistant *Enterococcus* Colonized Patient. *Intest Res*. 2015;13(1 PG-80-4):80–4.
- Kakkar E, Othman M. Fecal transplant in recurrent *clostridium difficile* enteritis. *Journal of General Internal Medicine*. 2017;32 (2 Supplement 1):S499.
- Kao D, Madsen K. Fecal microbiota transplantation (FMT) in the treatment of inflammatory bowel disease (IBD): A case report acg/astrazeneca clinical vignette award. *Am J Gastroenterol*. 2013;108:S415–6.
- Karlsson KA. Faecal transplantation for the treatment of recurrent *clostridium difficile* associated diarrhoea. *South African Gastroenterol Rev*. 2012;10(2 PG-19):19.
- Kelly CR, Olefson S, Jackson M. A challenging case of diarrhea after fecal microbiota transplant. *Am J Gastroenterol*. 2015;110(PG-S156-S157):S156–7.
- Kim JE, Gweon TG, Yeo CD, Cho YS, Kim GJ, Kim JY, et al. A case of *Clostridium difficile* infection complicated by acute respiratory distress syndrome treated with fecal microbiota transplantation. *World J Gastroenterol*. 2014;20(35 PG-12687-12690):12687–90.

- Kleger A, Schnell J, Essig A, Wagner M, Bommer M, Seufferlein T, et al. Fecal transplant in refractory *Clostridium difficile* colitis. *Dtsch Arztebl Int.* 2013;110(7 PG-108-15):108–15.
- Konturek P, Haziri D, Helfritsch H, Hess T, Heymann S, Harsch I. Successful Therapy of Severe Pseudomembranous *Clostridium difficile* Colitis using Combination of Fecal Microbiota Therapy and Fidaxomicin. *Med Princ Pract.* 2016;15(PG-).
- Kurtz M, Morgan M. Concomitant *Clostridium difficile* colitis and cytomegalovirus colitis in an immunocompetent elderly female. *BMJ Case Rep.* 2012;(no pagination)(1379 PG-).
- Laster J, Sultan M, Mattar M. Fecal microbiota transplantation in refractory *clostridium difficile* infection in children: Case report and review of the literature. *Am J Gastroenterol.* 2015;110(PG-S399):S399.
- Lingala S. Fecal microbiota transplantation in critically ill patient with severe *clostridium difficile* colitis. *Gastroenterology.* 2014;1(PG-S-251):S-251.
- Loke P, Heine RG, McWilliam V, Cameron DJ, Tang ML, Allen KJ. Fecal microbial transplantation in a pediatric case of recurrent *Clostridium difficile* infection and specific antibody deficiency. *Pediatr Allergy Immunol.* 2016;27(8 PG-872-874):872–4.
- Loke P, Heine RG, McWilliam V, Cameron DJS, Tang MLK, Allen KJ. Fecal microbial transplantation in a pediatric case of recurrent *Clostridium difficile* infection and specific antibody deficiency. *Pediatr Allergy Immunol.* 2016;27(8 PG-872-874):872–4.
- Mandalia A, Kraft CS, Dhere T. Diverticulitis after fecal microbiota transplant for *C. difficile* infection. *Am J Gastroenterol.* 2014;109(12 PG-1956-1957):1956–7.
- Marcos LA, Gersh A, Blanchard K, Foil S, Mallini B, Farrell SE, et al. Fecal transplantation to treat initial severe *Clostridium difficile* infection with sepsis. *J Miss State Med Assoc.* 2015;56(2 PG-38-40):38–40.
- Matsushita M, Watanabe O, Nakamura M, Yamamura T, Funasaka K, Ohno E, et al. Two cases of fecal microbiota transplantation in patients with recurrent *clostridium difficile* infection. *J Gastroenterol Hepatol.* 2016;31(PG-217):217.
- Midani D, Criner G, Clauss H, Smith MS, Ehrlich AC. Fecal microbiota transplant as a bridge to organ transplant: An alternative indication for treatment of *C. Difficile* in a critically ill patient. *Am J Gastroenterol.* 2016;111(PG-S616):S616.
- Million M, Hocquart M, Seghboyan JM, Griffiths K, Halfon P, Lagier JC, et al. Faecal microbiota transplantation as salvage therapy for fulminant *Clostridium difficile* infections. *Int J Antimicrob Agents.* 2015;46(2 PG-227-228):227–8.
- Mohamed A, Hogan N, Moloney M. Faecal transplant in the management of Crohn's colitis with persistent *clostridium difficile* infection: A case report. *Ir J Med Sci.* 2015;184 (6 Supplement 1)(PG-S236):S236.
- Morales SJ, Medvedev S, Lee A, Mattar M. A case of successful treatment of refractory *clostridium difficile* colitis with fecal microbiota transplantation in a critically ill patient. *Am J Gastroenterol.* 2015;110(PG-S146):S146.



- Navalkele BD, Lerner SA. Intravenous tigecycline facilitates cure of severe *Clostridium difficile* infection (CDI) after failure of standard therapy: A case report and literature review of tigecycline use in CDI. *Open Forum Infect Dis*. 2016;3 (2) (no pagination)(ofw094 PG-).
- Neelakanta A, Moudgal V, Upadhyay N, Valenstein P, Gunaratnam NT. Title: Successful treatment of refractory *clostridium difficile* infection(CDI) with intestinal microbiota transplant (IMT) in two patients with inflammatory bowel disease (IBD) and its effects on IBD. *Gastroenterology*. 2012;1)(PG-S395):S395.
- Neemann K, Eichele DD, Smith PW, Bociek R, Akhtari M, Freifeld A. Fecal microbiota transplantation for fulminant *Clostridium difficile* infection in an allogeneic stem cell transplant patient. *Transpl Infect Dis*. 2012;14(6 PG-E161-5):E161-5.
- Neemann K, Eichele DDD, Smith PPW, Bociek R, Akhtari M, Freifeld A. Fecal microbiota transplantation for fulminant *Clostridium difficile* infection in an allogeneic stem cell transplant patient. *Transpl Infect Dis*. 2012;14(6 PG-E161-E165):E161–5.
- Oppfeldt AM, Dahlerup JF, Christensen LA, Hvas CL. Faecal microbiota transplantation for recurring *Clostridium difficile* infection in a patient with Crohn's disease and ileorectal anastomosis. *BMJ Case Rep*. 2016;(PG-).
- Persky SE, Brandt LJ. Treatment of recurrent *Clostridium difficile*-associated diarrhea by administration of donated stool directly through a colonoscope. *Am J Gastroenterol*. 2000;95(11 PG-3283-3285):3283–5.
- Pitts K, Shah N, Uppal D, Hays RA. Fecal microbiota transplantation in a patient with liver cirrhosis: Changing the intestinal microbiota in a high-risk group. *Am J Gastroenterol*. 2015;110(PG-S358):S358.
- Popa D, Laszlo M, Ciobanu L, Ucenic E, Mihalache M, Pascu O. Self-Administered home series fecal “minitransplants” for recurrent *Clostridium difficile* infection on a rectal remnant. *J Gastrointest Liver Dis*. 2015;24(4 PG-531-533):531–3.
- Porr CI. Uncontrolled asthma - Case presentation. *Allergy Eur J Allergy Clin Immunol*. 2014;69(PG-585):585.
- Porter RJ, Fogg C. Faecal microbiota transplantation for *Clostridium difficile* infection in the United Kingdom. *Clin Microbiol Infect*. 2015 Jun;21(6):578–82.
- Porter RJ. Pulsed faecal microbiota transplantation for recalcitrant recurrent *Clostridium difficile* infection. *Clin Microbiol Infect*. 2015;21(3 PG-e23-e24):e23–4.
- Quera R, Espinoza R, Estay C, Rivera D. Bacteremia as an adverse event of fecal microbiota transplantation in a patient with Crohn's disease and recurrent *Clostridium difficile* infection. *J Crohn's Colitis*. 2014;8(3 PG-252-253):252–3.
- Raghunathan VM, Sheng I, Lim SH. Intestinal dysbiosis and allogeneic hematopoietic progenitor cell transplantation. *J Transl Med*. 2016;14 (1) (no pagination)(335 PG-).
- Rahman O, Farooq H, Mahmood SB, Khalid MB, Kapoor R, Wack M. *Clostridium difficile* enteritis and proctitis: Novel multimodality treatment regimen postcolectomy. *Crit Care Med*. 2016;44 (12 Supplement 1)(PG-520):520.

- Ramay FH, Amoroso A, Von Rosenvinge EC, Saharia K. Fecal microbiota transplantation for treatment of severe, recurrent, and refractory *Clostridium difficile* infection in a severely immunocompromised patient. *Infect Dis Clin Pract*. 2016;24(4 PG-237-240):237–40.
- Robin C, Paul M, Nebbad B, Beckerich F, Lepeule R, Ait Ammar N, et al. Fecal microbiota transplantation after allogeneic HSCT for curing recurrent *Clostridium difficile* infection: Why using the stem cell donor again? *Bone Marrow Transplant*. 2016;51(PG-S199-S200):S199–200.
- Russell G, Kaplan J, Ferraro M, Michelow IC. Fecal bacteriotherapy for relapsing *Clostridium difficile* infection in a child: A proposed treatment protocol. *Pediatrics*. 2010;126(1 PG-e239-e242):e239–42.
- Saeedi BJ, Morison DG, Kraft CS, Dhere T. Fecal Microbiota Transplant for *Clostridium difficile* Infection in a Pregnant Patient. *Obstet Gynecol*. 2017;129(3):507–9.
- Satokari R, Fuentes S, Mattila E, Jalanka J, de Vos WM, Arkkila P. Fecal transplantation treatment of antibiotic-induced, noninfectious colitis and long-term microbiota follow-up. *Case Rep Med*. 2014;2014(PG-913867):913867.
- Satokari R, Fuentes S, Mattila E, Jalanka J, De Vos WM, Arkkila P. Fecal transplantation treatment of antibiotic-induced. *Case Rep Med*. 2014;2014 (no pagination)(913867 PG-).
- Schunemann M, Oette M. Fecal microbiota transplantation for *Clostridium difficile*-associated colitis in a severely immunocompromized critically ill AIDS patient: A case report. *Aids*. 2014;28(5 PG-798-799):798–9.
- Seth AK, Rawal P, Bagga R, Jain P. Successful colonoscopic fecal microbiota transplantation for active ulcerative colitis: First report from India. *Indian J Gastroenterol*. 2016;35(5 PG-393-395):393–5.
- Seth AK, Rawal P, Bagga R. Successful stool transplantation for severe ulcerative colitis: First report from India. *Indian J Gastroenterol*. 2015;1(PG-A21-A22):A21–2.
- Shin JY, Ko EJ, Lee SH, Shin JB, Kim SI, Kwon KS, et al. Refractory pseudomembranous colitis that was treated successfully with colonoscopic fecal microbial transplantation. *Intest Res*. 2016;14(1 PG-83-8):83–8.
- Singh P, Udeh B, Dalton J, Udeh C, Hata J. Cost-effectiveness of 6 treatments for primary *Clostridium difficile* infection in an ICU population. *Crit Care Med*. 2014;1(PG-A1474):A1474.
- Singh S, Jing E, Stollman N. Self-Limited Sepsis Syndrome Following Fecal Microbiota Therapy for Refractory *C. difficile* Infection. *Dig Dis Sci*. 2016;61(9 PG-2488-2491):2488–91.
- Smith S. Intestinal microbiota transplantation: A case of Crohn's colitis with superimposed *Clostridium difficile* infection. *West Indian Med J*. 2013;62(7 PG-675-677):675–7.
- Sonpal N, Datta S, Mammen A, Haber G. The stool strikes back: Fecal transplantation for the treatment of *Clostridium difficile* infection. *Am J Gastroenterol*. 2015;110(PG-S159):S159.
- Soota K, Telfah M, Ramesh N, Pereira M, Lingutla D. Treatment of recurrent *Clostridium difficile* infection with combined jejunal and colonic fecal microbiota transplant. *Am J Gastroenterol*. 2013;108(PG-S398):S398.

- Stanley E, McNamara D. "Non-Resolving *C. difficile* infection cured by transplant." *Ir J Med Sci.* 2015;1(PG-S342):S342.
- Stein D, Rizvi S, Modiri AN, Fang T, Naik AS. Two case reports of toxic megacolon from *clostridium difficile* infection successfully treated with fecal microbiota therapy. *Gastroenterology.* 2015;1(PG-S645):S645.
- Stollman N, Smith M, Giovanelli A, Mendolia G, Burns L, Didyk E, et al. Frozen encapsulated stool in recurrent *clostridium difficile*: Exploring the role of pills in the treatment hierarchy of fecal microbiota transplant nonresponders. *Am J Gastroenterol.* 2015;110(4 PG-600-601):600–1.
- Stollman N, Surawicz C. Fecal transplant for *Clostridium difficile*. *Arch Intern Med.* 2012;172(10 PG-825):825.
- Stripling J, Kumar R, Baddley JW, Nellore A, Dixon P, Howard D, et al. Loss of vancomycin-resistant enterococcus fecal dominance in an organ transplant patient with *Clostridium difficile* colitis after fecal microbiota transplant. *Open Forum Infect Dis.* 2015;2 (2) (no pagination)(ofv078 PG-).
- Stysly B, Kukkadapu T, Singh E. *Clostridium difficile* in ulcerative colitis complicated by underlying aplastic anemia. *Am J Gastroenterol.* 2014;109(PG-S443):S443.
- Sun W, Arunachalam A, Siddique S, Zandman D. Multi-organism bacteremia after fecal microbiota transplantation for relapsing *clostridium difficile* infection. *Am J Gastroenterol.* 2014;109(PG-S420):S420.
- Syed R, Rahim U, Humphrey F, Ray A. Fecal microbiota transplant for severe complicated *clostridium difficile* infection via a loop ileostomy: A novel administration route. *Am J Gastroenterol.* 2015;110(PG-S142):S142.
- Tafesh Z, O'Neil S, Crawford Jr C V. Frozen universal stool for fecal microbiota transfer (FMT) in recurrent *C. difficile* infection. *Am J Gastroenterol.* 2015;110(PG-S588):S588.
- Tanaka T, Kato H, Fujimoto T. Successful fecal microbiota transplantation as an initial therapy for *Clostridium difficile* infection on an outpatient basis. *Intern Med.* 2016;55(8 PG-999-1000):999–1000.
- Tariq R, Smyrk T, Pardi DS, Tremaine WJ, Khanna S. New-onset microscopic colitis in an ulcerative colitis patient after fecal microbiota transplantation. *Am J Gastroenterol.* 2016;111(5 PG-751-752):751–2.
- Tian H, Ding C, Gong J, Wei Y, McFarland L V, Li N. Freeze-dried, capsulized fecal microbiota transplantation for relapsing *clostridium difficile* infection. *J Clin Gastroenterol.* 2015;49(6 PG-537-538):537–8.
- Trubiano JA, Gardiner B, Kwong JC, Ward P, Testro AG, Charles PG. Faecal microbiota transplantation for severe *Clostridium difficile* infection in the intensive care unit. *Eur J Gastroenterol Hepatol.* 2013;25(2 PG-255-7):255–7.
- Trubiano JA, Gardiner B, Kwong JC, Ward P, Testro AG, Charles PGP. Faecal microbiota transplantation for severe *clostridium difficile* infection in the intensive care unit. *Eur J Gastroenterol Hepatol.* 2013;25(2 PG-255-257):255–7.
- Trubiano JA, George A, Barnett J, Siwan M, Heriot A, Prince HM, et al. A different kind of "allogeneic transplant": Successful fecal microbiota transplant for recurrent and refractory *Clostridium difficile* infection in a patient with relapsed aggressive B-cell lymphoma. *Leuk Lymphoma.* 2015;56(2 PG-512-514):512–4.

Walia R, Garg S, Song Y, Girotra M, Cuffari C, Fricke WF, et al. Efficacy of fecal microbiota transplantation in 2 children with recurrent *Clostridium difficile* infection and its impact on their growth and gut microbiome. *J Pediatr Gastroenterol Nutr*. 2014;59(5 PG-565-570):565–70.

Wang J, Xiao Y, Lin K, Song F, Ge T, Zhang T. Pediatric severe pseudomembranous enteritis treated with fecal microbiota transplantation in a 13-month-old infant. *Biomed Reports*. 2015;3(2 PG-173-175):173–5.

Wang PT, Fashandi AZ, Hays RA. Comparison of laparoscopic loop ileostomy and fecal microbiota transplantation in a patient with two episodes of severe and complicated *clostridium difficile* infection: A case report. *Am J Gastroenterol*. 2016;111(PG-S624):S624.

Wonderlick JS, D'Agostino R. Fecal microbiota transplantation via fluoroscopy-guided nasojejunal catheter placement: indications, technique, and the role of radiology. *Abdom Radiol*. 2016;41(10 PG-2020-2025):2020–5.

You D, Johnson M, Duplessis C, Speziale A. Successful Use of Fecal Bacteriotherapy in Severe Crohn's Colitis and Refractory *Clostridium difficile* Infection. *Am J Gastroenterol*. 2011;106(PG-S315):S315.

You DM, Franzos MA, Holman RP. Successful treatment of fulminant *Clostridium difficile* infection with fecal bacteriotherapy. *Ann Intern Med*. 2008;148(8 PG-632-633):632–3.

Youssef MA, Gavin M. Fecal microbiota transplant: A case report in an immunosuppressed patient with crohn's disease and recurrent *clostridium difficile* infection. *Gastroenterology*. 2013;1(PG-S626):S626.

Yu S, Abdelkarim A, Nawras A, Hinch BT, Mbaso C, Valavoor S, et al. Fecal transplant for treatment of toxic megacolon associated with *clostridium difficile* colitis in a patient with duchenne muscular dystrophy. *Am J Ther*. 2016;23(2 PG-e609-e613):e609–13.

Zainah H, Silverman A. Fecal Bacteriotherapy: A Case Report in an Immunosuppressed Patient with Ulcerative Colitis and Recurrent *Clostridium difficile* Infection. *Case Reports Infect Dis*. 2012;2012(PG-810943):810943.

#### D.1.5. Non-English language:

*Chinese* Li N, Tian H, Ma C, Ding C, Ge X, Gu L, Zhang X, Yang B, Hua Y, Zhu Y, Zhou Y. Efficacy analysis of fecal microbiota transplantation in the treatment of 406 cases with gastrointestinal disorders [Chinese]. *Zhonghua Wei Chang Wai Ke Za Zhi*. 2017;20(1):40-46.

*Chinese* Wang Y, Yang B, Ye Y, Li Z, Kang W. Therapeutic effects and the possible mechanism of fecal transplantation on rats with *Clostridium difficile*-associated pseudomembranous colitis. [Chinese]. *Chinese J Microbiol Immunol*. 2015;35(8 PG-582-586):582–6.

*Chinese* Yang Y, Wang Z. Advances in study on fecal microbiota transplantation. [Chinese]. *Chinese J Gastroenterol*. 2014;19(1 PG-1-5):1–5.

*Czech* Polak P, Freibergerovala M, Husa P, Jurankova J, Svacinka R, Mikesova L, et al. Fecal bacteriotherapy for the treatment of recurrent *Clostridium difficile* colitis used in the Clinic of Infectious Diseases of the

- University Hospital Brno in 2010-2014 - a prospective study. [Czech]. *Epidemiol Mikrobiol Imunol Cas Spol pro Epidemiol a Mikrobiol Ces Lek Spol JE*. 2015;64(4 PG-232-235):232-5.
- Czech* Polak P, Freibergova M, Husa P, Jurankova J, Svacinka R, Mikesova L, et al. [Fecal bacteriotherapy for the treatment of recurrent *Clostridium difficile* colitis used in the Clinic of Infectious Diseases of the University Hospital Brno in 2010-2014 - a prospective study]. *Epidemiol Mikrobiol Imunol*. 2015;64(4 PG-232-5):232-5.
- Czech* Polak P, Freibergova M, Jurankova J, Kocourkova H, Mikesova L, Svacinka R, et al. First experiences with faecal bacteriotherapy in the treatment of relapsing pseudomembranous colitis due to *Clostridium difficile*. [Czech]. *Klin Mikrobiol Infekc Lek*. 2011;17(6 PG-214-217):214-7.
- Czech* Polak P, Husa P, Freibergova M. Colitis due to *Clostridium difficile* in broader context. [Czech]. *Interni Med pro Praxi*. 2014;16(6 PG-241-244):241-4.
- Dutch* Holvoet T, Van De Wiele T, Boelens J, Raes J, Hindryckx P, De Vos M, et al. Fecal transplantation: Overview of the indications. [Dutch]. *Tijdschr Geneesk*. 2014;70(6 PG-289-297):289-97.
- Dutch* Nieuwdorp M, Van Nood E, Speelman P, Van Heukelem HA, Jansen JM, Visser CE, et al. Treatment of recurrent *Clostridium difficile*-associated diarrhoea with a suspension of donor faeces. [Dutch]. *Ned Tijdschr Geneesk*. 2008;152(35 PG-1927-1932):1927-32.
- Dutch* Nood E, Keller JJ, Kuijper EJ, Speelman P. New treatment options for infections with. [Dutch]. *Ned Tijdschr Geneesk*. 2014;158(3 PG-).
- Dutch* Terveer EM, van Beurden YH, Kuijper EJ, Keller JJ. [Fecal microbiota transplantation, a novel therapy for recurrent *Clostridium difficile* infection]. *Ned Tijdschr Tandheelkd*. 2016;123(9 PG-406-9):406-9.
- Dutch* van Nood E, Keller JJ, Kuijper EJ, Speelman P. [New treatment options for infections with *Clostridium difficile*]. [Dutch]. *Ned Tijdschr Geneesk*. 2013;157(48 PG-A6580):A6580.
- Finnish* Harkonen N. [Reccurent pseudomembranous colitis treated with the donor feces]. *Duodecim*. 1996;112(19 PG-1803-4):1803-4.
- French* Giger A, Barberini L, Bruchez P, Castioni J, Claude F, Rochat MC, et al. General internal medicine in hospital practice: The year 2013 put into perspective by residents. [French]. *Rev Med Suisse*. 2014;10(414 PG-164-170):164-70.
- French* Kohn M, Robin C, Beckerich F, Cordonnier C. *Clostridium difficile* infections and blood disease: What should I know? Infections a *Clostridium difficile* et hemopathies: que faut-il savoir?. [French]. *Hematologie*. 2015;21(1 PG-18-27):18-27.
- French* Lagier JC, Raoult D. [Fecal microbiota transplantation: indications and perspectives]. *M S-Medecine Sci*. 2016;32(11 PG-991-997):991-7.
- French* Lagier JC. Faecal microbiota transplantation: From practice to legislation before considering industrialization. *Clin Microbiol Infect*. 2014;20(11 PG-1112-1118):1112-8.

- French* Megerlin F, Fouassier E, Lopert R, Bourlioux P. Faecal microbiota transplantation: A sui generis biological drug, not a tissue. *Ann Pharm Fr.* 2014;72(4 PG-217-220):217–20.
- French* Megerlin F, Fouassier E. [Faecal microbiota transplantation in France: what applicable law?]. *Ann Pharm Fr.* 2014;72(5 PG-363-74):363–74.
- French* Megerlin F, Fouassier E. Faecal microbiota transplantation in France: What applicable law?. [French]. *Ann Pharm Fr.* 2014;72(5 PG-363-374):363–74.
- French* Rozier P, Fraisse T, Lauda M, Priner M, Forestier E, Paccalin M. Clostridium difficile in geriatrics. [French]. *Cah l'Annee Gerontol.* 2014;6(3 PG-107-113):107–13.
- French* Rozier P, Fraisse T, Lauda M, Priner M, Forestier E, Paccalin M. Clostridium difficile in geriatrics. [French]. *Cah l'Annee Gerontol.* 2014;6(3 PG-107-113):107–13.
- French* Seksik P. Clostridium difficile associated colitis. [French]. *Hepato-Gastro Oncol Dig.* 2016;23(8 PG-775-784):775–84.
- French* Sokol H, Galperine T, Kapel N, Bourlioux P, Seksik P, Barbut F, et al. Fecal microbiota transplantation for treatment of relapsing clostridium difficile infection: Guidelines for clinical practice. [French]. *Hepato-Gastro Oncol Dig.* 2015;22(4 PG-278-290):278–90.
- French* Surawicz CM, Alexander J. Treatment of refractory and recurrent Clostridium difficile infection. *Nat Rev Gastroenterol Hepatol.* 2011;19(PG-).
- French* Surawicz CM. [The microbiota and infectious diarrhea]. [French]. *Gastroenterol Clin Biol.* 2010;34 Suppl 1(PG-S29-36):S29–36.
- French* Surawicz CM. [The microbiota and infectious diarrhea]. *Gastroenterol Clin Biol.* 2010;34 Suppl 1(PG-S29-36):S29–36.
- French* Surawicz CM. Clostridium difficile infection: risk factors, diagnosis and management. *Curr Treat Options Gastroenterol.* 2015;13(1 PG-121-9):121–9.
- French* Terrier MCZ, Frossard JL, Simonet ML. Recurrent Clostridium difficile infections : The importance of the intestinal microbiota. [French]. *Rev Med Suisse.* 2013;9(402 PG-1898-1904):1898–904.
- French* Tissot F, Maillard MH. Clostridium difficile infections: Update on new European recommendations. [French]. *Rev Med Suisse.* 2014;10(427 PG-913-919):913–9.
- French* Voide C, Asner S, Giulieri S, Cavassini M, Merz L, Tissot F, et al. Infectious diseases. [French]. *Rev Med Suisse.* 2014;10(412-413-61–65):61–5.
- French* Werner CC. Fecal transplantation in the treatment of Clostridium difficile infections. [French]. *Rev Med Suisse.* 2013;9(373 PG-388-389):388–9.
- German* eufferlein T, Kleger A, Nitschmann S. [Recurrent Clostridium difficile infection. Treatment with duodenal infusion of donor feces]. *Internist.* 2014;55(4 PG-455-9):455–9.



- German Hagel S, Stallmach A, Vehreschild M, Angeli W, Bachmann O, Gross M, et al. Fecal microbiota transplant in patients with recurrent *Clostridium difficile* infection - A retrospective multicenter observational study from the MicroTrans registry. [German]. *Dtsch Arztebl Int.* 2016;113(35-36-583-589):583-9.
- German Hibbeler B. *Clostridium difficile*: Fecal bacteriotherapy as an option. *Dtsch Arztebl Int.* 2016;113(5 PG-A185):A185.
- German Liebhardt E, Seufferlein T, Wagner M. Fecal microbiota transplantation for *Clostridium difficile* infection. [German]. *Arzneimitteltherapie.* 2016;34(9 PG-285-291):285-91.
- German Lubbert C, Weis S. [Drug therapy of infectious diarrhea: part 1: acute diarrhea]. *Internist.* 2013;54(11 PG-1383-92):1383-92.
- German Lubbert C, Weis S. Drug therapy of infectious diarrhea: Part 1: Acute diarrhea. [German]. *Internist.* 2013;54(11 PG-1383-1392):1383-92.
- German Lubbert C. Fecal microbiota transplantation (FMT): Indications for treatment and future perspectives. [German]. *Diabetologe.* 2016;12(6 PG-409-419):409-19.
- German Menzel J. Fecal transplantation for refractory chronic *Clostridium difficile* infection. [German]. *Gastroenterologe.* 2013;8(4 PG-336-337):336-7.
- German Ramsauer B, König C, Sabelhaus T, Ockenga J, Otte JM. Fecal microbiota transplantation in relapsing *clostridium difficile* colitis. [German]. *MMW-Fortschritte der Medizin.* 2016;158(PG-17-20):17-20.
- German Rohrenbach J, Matthess A, Maier R, Von Bunau R. Treatment of children with *E. coli* strain Nissle 1917. Results of a prospective data collection with 668 patients. [German]. *Padiatr Prax.* 2009;73(4 PG-645-652):645-52.
- German Rohrenbach J, Matthess A, Maier R, Von Bunau R. Treatment of children with *E. coli* strain Nissle 1917. Results of a prospective data collection with 668 patients. [German]. *Padiatr Prax.* 2009;73(4 PG-645-652):645-52.
- German Rosien U, Hagel S, Gotz M. Stool transplant for recurrent *Clostridium difficile* infection. [German]. *Gastroenterologe.* 2015;10(2 PG-122-126):122-6.
- German Rosien U, Hagel S, Gotz M. Stool transplant for recurrent *Clostridium difficile* infection. [German]. *Gastroenterologe.* 2015;10(2 PG-122-126):122-6.
- German Salzberger B, Rauscher C. [The microbiome of the gut in critically ill patients]. *Med Klin Intensivmed Notfmed.* 2015;110(7 PG-521-5):521-5.
- German Salzberger B, Rauscher C. The microbiome of the gut in critically ill patients. [German]. *Medizinische Klin - Intensivmed und Notfallmedizin.* 2015;110(7 PG-521-525):521-5.
- German Schmelz R, Hampe J. [Fecal microbiota transplantation: when and for whom?]. *Dtsch Medizinische Wochenschrift.* 2014;139(23 PG-1237-9):1237-9.

- German Schmitz F. Fecal transplantation is highly effective in the treatment of recurrent *Clostridium difficile* infection. [German]. *Gastroenterologe*. 2013;8(1 PG-54-55):54–5.
- German Seufferlein T, Kleger A, Nitschmann S. Recurrent *Clostridium difficile* infection. Treatment with duodenal infusion of donor feces. [German]. *Internist*. 2014;55(4 PG-455-459):455–9.
- German Stallmach A. [*Clostridium difficile* infection : What is currently available for treatment?]. *Internist*. 2016;57(12 PG-1182-1190):1182–90.
- German Stallmach A. *Clostridium difficile* infection: What is currently available for treatment?. [German]. *Internist*. 2016;57(12 PG-1182-1190):1182–90.
- German Storr M, Starostzik C. [Stool transplantation: also an option for irritable bowel syndrome?]. *MMW Fortschr Med*. 2014;156(12 PG-16):16.
- German Storr M. [Donor stool now available in capsules]. *MMW Fortschr Med*. 2015;157 Suppl 1(PG-32):32.
- German Trautmann M. Fecal transplantation in *Clostridium difficile* colitis: New studies about a long-known therapeutic option. [German]. *Krankenhauspharmazie*. 2013;34(8 PG-414-415):414–5.
- German U R, S H, M G. Erratum to: Stool transplant for recurrent *Clostridium difficile* infection [ *Gastroenterologe*, (2015), DOI:10.1007/s11377-014-0962-8]. [German]. *Gastroenterologe*. 2015;10(2 PG-110):110.
- German von Muller L. [New aspects on *Clostridium difficile* infection]. *Dtsch Medizinische Wochenschrift*. 2016;141(16 PG-1144-7):1144–7.
- German Von Muller L. New aspects on *Clostridium difficile* infection. [German]. *Dtsch Medizinische Wochenschrift*. 2016;141(16 PG-1144-1147):1144–7.
- German Weis S, John E, Lippmann N, Mossner J, Lubbert C. [*Clostridium difficile* infection (CDI) in the course of time - an issue only for the internist?]. *Zentralbl Chir*. 2014;139(4 PG-460-8):460–8.
- German Zoller V, Laguna AL, Prazeres Da Costa O, Buch T, Goke B, Storr M. [Fecal microbiota transfer (FMT) in a patient with refractory irritable bowel syndrome]. *Dtsch Medizinische Wochenschrift*. 2015;140(16 PG-1232-6):1232–6.
- German Zoller V, Laguna AL, Prazeres Da Costa O, Buch T, Goke B, Storr M. Fecal microbiota transfer (FMT) in a patient with refractory irritable bowel syndrome. [German]. *Dtsch Medizinische Wochenschrift*. 2015;140(16 PG-1232-1236):1232–6.
- Greek Mentis AFA, Gypas F, Mentis AF. Human enteric microbiome: Its role in health and disease. [Greek]. *Arch Hell Med*. 2013;30(3 PG-272-288):272–88.
- Hebrew Israeli E, Shoenfeld Y. [Harnessing nature for treating infectious and autoimmune diseases: good and bad bacteria]. [Hebrew]. *Harefuah*. 2013;152(4 PG-188-189, 249):188–189,249.
- Hebrew Israeli E, Shoenfeld Y. [Harnessing nature for treating infectious and autoimmune diseases: good and bad bacteria]. *Harefuah*. 2013;152(4 PG-188-9, 249):188–189,249.



- Hebrew Maharshak N. [Use of fecal microbial transplantations for disease states in Israel]. Harefuah. 2015;154(3 PG-152-4, 213):152–154,213.
- Hebrew Maharshak N. Use of fecal microbial transplantations for disease states in Israel. [Hebrew]. Harefuah. 2015;154(3 PG-152-4, 213):152–154,213.
- Hungarian Kovacs G. [To the Editors, regarding feces transplantation]. [Hungarian]. Orv Hetil. 2013;154(11 PG-434-435):434–5.
- Hungarian Kovacs G. [To the Editors, regarding feces transplantation]. Orv Hetil. 2013;154(11 PG-434-5):434–5.
- Hungarian Nagy GG, Varvolgyi C, Balogh Z, Orosi P, Paragh G. [Detailed methodological recommendations for the treatment of Clostridium difficile-associated diarrhea with faecal transplantation]. Orv Hetil. 2013;154(1 PG-10-9):10–9.
- Hungarian Nagy GG, Varvolgyi C, Balogh Z, Orosi P, Paragh G. Detailed methodological recommendations for the treatment of Clostridium difficile-associated diarrhea with faecal transplantation. [Hungarian]. Orv Hetil. 2013;154(1 PG-10-19):10–9.
- Hungarian Nagy GG, Varvolgyi C, Paragh G. [Successful treatment of life-threatening, treatment resistant Clostridium difficile infection associated pseudomembranous colitis with faecal transplantation]. Orv Hetil. 2012;153(52 PG-2077-83):2077–83.
- Hungarian Szabolcs V, Zsuzsanna N, Aron V, Jen S, David S, Zsafia F, et al. Experience with fecal microbiota transplantation in the treatment of clostridium difficile infection. [Hungarian]. Orv Hetil. 2014;155(44 PG-1758-1762):1758–62.
- Hungarian Vigvari S, Nemes Z, Vincze A, Solt J, Sipos D, Feiszt Z, et al. [Experience with fecal transplantation in the treatment of Clostridium difficile infection]. Orv Hetil. 2014;155(44 PG-1758-62):1758–62.
- Hungarian Nagy GG, Varvolgyi C, Paragh G. [Successful treatment of life-threatening, treatment resistant Clostridium difficile infection associated pseudomembranous colitis with faecal transplantation]. [Hungarian]. Orv Hetil. 2012;153(52 PG-2077-2083):2077–83.
- Italian Russello G, Brovarone F, Bardaro M, Carretto E. Treating Clostridium difficile infection with faecal transplantation: Donor microbiological testing. Infez Med. 2014;22(1 PG-5-10):5–10.
- Japanese akamur I, Kunihiro M, Kato H. Bacteremia due to Clostridium difficile. [Japanese]. Kansenshogaku zasshi. 2004;The Journal of the Japanese Association for Infectious Diseases. 78(12 PG-1026-1030):1026–30.
- Japanese Nakamur I, Kunihiro M, Kato H. [Bacteremia due to Clostridium difficile]. Kansenshogaku Zasshi - J Japanese Assoc Infect Dis. 2004;78(12 PG-1026-30):1026–30.
- Japanese Ohkusa T, Koido S. [Gut Microbiota and Internal Diseases: Update Information. Topics: II. Fecal microbiota transplantation and its clinical application]. Nippon Naika Gakkai Zasshi - J Japanese Soc Intern Med. 2015;104(1 PG-42-7):42–7.

- Japanese Osada T, Ishikawa D, Watanabe S. [Fecal microbiota transplantation therapy for patients with gastrointestinal tract diseases]. *Nippon Shokakibyo Gakkai Zasshi - Japanese J Gastroenterol*. 2015;112(11 PG-1973-81):1973–81.
- Japanese Suzuki K, Kitahara T, Yokota A. Clinical studies of acute hemorrhagic colitis associated with antibiotic therapy. 3. Fecal bacterial flora and fecal short chain fatty acids. [Japanese]. *IRYO - Japanese J Natl Med Serv*. 1984;38(6 PG-570-576+543-544):570–576+543.
- Korean Kim SW. [Treatment of refractory or recurrent *Clostridium difficile* infection]. *Korean J Gastroenterol Sohwagi Hakhoe Chi*. 2012;60(2 PG-71-8):71–8.
- Korean Kim SW. [Treatment of refractory or recurrent *Clostridium difficile* infection]. [Korean]. *Korean J Gastroenterol*. 2012;60(2 PG-71-78):71–8.
- Korean Ko JS. [The intestinal microbiota and human disease]. [Korean]. *Korean J Gastroenterol*. 2013;62(2 PG-85-91):85–91.
- Korean Ko JS. [The intestinal microbiota and human disease]. *Korean J Gastroenterol Sohwagi Hakhoe Chi*. 2013;62(2 PG-85-91):85–91.
- Norwegian Lund-Tonnesen S, Berstad A, Schreiner A, Midtvedt T. [*Clostridium difficile*-associated diarrhea treated with homologous feces]. *Tidsskr Den Nor Laegeforening*. 1998;118(7 PG-1027-30):1027–30.
- Norwegian Lund-Tonnesen S, Berstad A, Schreiner A, Midtvedt T. *Clostridium difficile*-associated diarrhoea treated with homologous faeces. [Norwegian]. *Tidsskr Den Nor Laegeforening*. 1998;118(7 PG-1027-1030):1027–30.
- Polish Malopolska M, Fol M. [Intestinal microbiota transplantation for the treatment of *Clostridium difficile* infection]. *Med Dosw Mikrobiol*. 2015;67(3-4-207–19):207–19.
- Polish Malopolska M, Fol M. Intestinal microbiota transplantation for the treatment of *Clostridium difficile* infection. [Polish]. *Med Dosw Mikrobiol*. 2015;67(3-4-207–219):207–19.
- Polish Piekarska M, Wandalowicz AD, Miigoc H. [*Clostridium difficile* infection--diagnostics, prevention and treatment]. *Pol Merkur Lek*. 2014;36(214 PG-278-82):278–82.
- Polish Piekarska M, Wandalowicz AD, Miigoc H. [*Clostridium difficile* infection--diagnostics, prevention and treatment]. [Polish]. *Pol Merkur Lekarski*. 2014;36(214 PG-278-282):278–82.
- Polish Rebizak E, Sierant K, Labuzek K, Okopien B. [Fecal transplantation the future therapy?]. *Pol Merkur Lek*. 2015;39(230 PG-73-6):73–6.
- Serbian Suljagic V, Djordjevic D, Lazic S, Mijovic B. [Epidemiological characteristics of nosocomial diarrhea caused by *Clostridium difficile* in a tertiary level hospital in Serbia]. *Srp Arh Celok Lek*. 2013;141(7-8-482–9):482–9.
- Serbian Suljagic V, Djordjevic D, Lazic S, Mijovic B. Epidemiological characteristics of nosocomial diarrhea caused by *Clostridium difficile* in a tertiary level hospital in Serbia. [Serbian]. *Srp Arh Celok Lek*. 2013;141(7-8-482–489):482–9.

- Slovakian Sturdik I, Hlavaty T, Payer J. [Fecal microbiota transplantation]. *Vnitr Lek.* 2016;62(2 PG-147-51):147–51.
- Slovakian Sturdik I, Hlavaty T, Payer J. Fecal microbiota transplantation. [Slovak]. *Vnitr Lek.* 2016;62(2 PG-147-151):147–51.
- Spanish Garcia-Garcia-de-Paredes A, Rodriguez-de-Santiago E, Aguilera-Castro L, Ferre-Aracil C, Lopez-Sanroman A. Fecal microbiota transplantation. [Spanish]. *Gastroenterol Hepatol.* 2015;38(3 PG-123-134):123–34.
- Spanish Halabe Cherem J, Hoyo Ulloa I. [Successful home-made fecal transplant for an elderly woman]. *Gac Med Mex.* 2014;150(1 PG-106-7):106–7.
- Spanish Hernandez-Rocha C, Pidal P, Ajenjo MC, Quera R, Quintanilla M, Lubascher J, et al. [Chilean consensus of prevention, diagnosis and treatment of *Clostridium difficile*-associated diarrhea]. *Rev Chil Infectol.* 2016;33(1 PG-98-118):98–118.
- Spanish Montejano Sanchez R. Duodenal infusion of donor feces for recurrent *Clostridium difficile*. [Spanish]. *Rev Clin Esp.* 2013;213(4 PG-213):213.
- Spanish Moscoso F, Simian D, Rivera D, Acuna G, Quera R. [Fecal microbiota transplantation in recurrent *Clostridium difficile* infection. Report of one case]. *Rev Med Chil.* 2015;143(4 PG-531-5):531–5.
- Spanish Moscoso F, Simian D, Rivera D, Acuna G, Quera R. Fecal microbiota transplantation in recurrent *Clostridium difficile* infection. Report of one case. [Spanish]. *Rev Med Chil.* 2015;143(4 PG-531-535):531–5.
- Spanish Munez E, Ramos A, Banos I, Cuervas-Mons V. [Fecal transplantation for the treatment of relapsing diarrhea associated with *Clostridium difficile* infection in a liver transplantation patient]. *Med Clin (Barc).* 2016;146(1 PG-e3-4):e3–4.
- Spanish Pareja-Sierra T. [Diarrhea associated with *Clostridium difficile* in the elderly: new perspectives]. *Rev Esp Geriatr Gerontol.* 2014;49(4 PG-188-93):188–93.
- Spanish Pareja-Sierra T. Diarrhea associated with *Clostridium difficile* in the elderly: New perspectives. [Spanish]. *Rev Esp Geriatr Gerontol.* 2014;49(4 PG-188-193):188–93.
- Spanish: Munez E, Ramos A, Banos I, Cuervas-Mons V. Fecal transplantation for the treatment of relapsing diarrhea associated with *Clostridium difficile* infection in a liver transplantation patient. [Spanish]. *Med Clin (Barc).* 2016;146(1 PG-e3-e4):e3–4.
- Swiss Giger A, Barberini L, Bruchez P, Castioni J, Claude F, Cosma Rochat M, et al. [General internal medicine in hospital practice: the year 2013 put into perspective by residents]. *Rev Med Suisse.* 2014;10(414 PG-164, 166-70):164,166–170.

#### D.1.6. Basic sciences:

- Basic scilhekweazu F, Fofanova T, Nagy-Szakal D, Hulten K, Queliza K, Opekun A, et al. Complex and defined bacteriotherapy can inhibit acute colitis in mice. *J Pediatr Gastroenterol Nutr*. 2016;63(PG-S277-S278):S277–8.
- Grehan MJ, Borody TJ, Leis SM, Campbell J, Mitchell H, Wettstein A. Durable alteration of the colonic microbiota by the administration of donor fecal flora. *J Clin Gastroenterol*. 2010;44(8 PG-551-561):551–61.
- Gu S, Chen Y, Zhang X, Lu H, Lv T, Shen P, et al. Identification of key taxa that favor intestinal colonization of *Clostridium difficile* in an adult Chinese population. *Microbes Infect*. 2016;18(1 PG-30-38):30–8.
- Halpin AL, de Man TJ, Kraft CS, Perry KA, Chan AW, Lieu S, et al. Intestinal microbiome disruption in patients in a long-term acute care hospital: A case for development of microbiome disruption indices to improve infection prevention. *Am J Infect Control*. 2016;44(7 PG-830-6):830–6.
- Halpin AL, de Man TJB, Kraft CS, Perry KA, Chan AW, Lieu S, et al. Intestinal microbiome disruption in patients in a long-term acute care hospital: A case for development of microbiome disruption indices to improve infection prevention. *Am J Infect Control*. 2016;44(7 PG-830-836):830–6.
- Hamilton MJ, Weingarden AR, Unno T, Khoruts A, Sadowsky MJ. High-throughput DNA sequence analysis reveals stable engraftment of gut microbiota following transplantation of previously frozen fecal bacteria. *Gut Microbes*. 2013;4(2):125–35.
- Hecker MT, Obrenovich ME, Cadnum JL, Jencson AL, Jain AK, Ho E, et al. Fecal microbiota transplantation by freeze-dried oral capsules for recurrent *clostridium difficile* infection. *Open Forum Infect Dis*. 2016;3 (2) (no pagination)(ofw091 PG-).
- Hevia A, Delgado S, Margolles A, Sanchez B. Application of density gradient for the isolation of the fecal microbial stool component and the potential use thereof. *Sci Rep*. 2015;5(PG-16807):16807.
- Jalanka J, Mattila E, Jouhten H, Hartman J, de Vos WM, Arkkila P, et al. Long-term effects on luminal and mucosal microbiota and commonly acquired taxa in faecal microbiota transplantation for recurrent *Clostridium difficile* infection. *BMC Med*. 2016;14 (1) (no pagination)(155 PG-).
- Khanna S, Montassier E, Patel R, Kammer PP, Knights D, Pardi D, et al. Gut microbiome signatures at the time of primary *clostridium difficile* infection predict recurrence. *Gastroenterology*. 2016;1(PG-S23):S23.
- Khanna S, Montassier E, Schmidt B, Lynch D, Bernard C, Lekatz H, et al. Gut microbiota changes as predictors of treatment failure in primary *clostridium difficile* infection. *Am J Gastroenterol*. 2015;110(PG-S578):S578.
- Khanna S, Vazquez-Baeza Y, Gonzalez A, Weiss S, Schmidt B, Muniz-Pedrogo D, et al. Changes in microbial ecology after fecal microbiota transplantation for recurrent *C. Difficile* infection depends on underlying inflammatory bowel disease. *Am J Gastroenterol*. 2016;111(PG-S75):S75.
- Khoruts A, Dicksved J, Jansson JK, Sadowsky MJ. Changes in the composition of the human fecal microbiome after bacteriotherapy for recurrent *Clostridium difficile*-associated diarrhea. *J Clin Gastroenterol*. 2010;44(5):354–60.

- Kumar R, Maynard CL, Eipers P, Goldsmith KT, Ptacek T, Grubbs JA, et al. Colonization potential to reconstitute a microbe community in patients detected early after fecal microbe transplant for recurrent *C. difficile*. *BMC Microbiol.* 2016;16 (1) (no pagination)(5 PG-).
- Kumar V, Zhou E, Mansoor MS, Feuerstadt P. Treatment of initial recurrence of *C. Difficile* infection (CDI) with vancomycin may not prevent eventual need for fecal microbial transplantation (FMT). *Am J Gastroenterol.* 2016;111(PG-S93):S93.
- Landy J, Walker AW, Li J V, Al-Hassi HO, Ronde E, English NR, et al. Variable alterations of the microbiota, without metabolic or immunological change, following faecal microbiota transplantation in patients with chronic pouchitis. *Sci Rep. England;* 2015;5:12955.
- Li SS, Zhu A, Benes V, Costea PI, Hercog R, Hildebrand F, et al. Durable coexistence of donor and recipient strains after fecal microbiota transplantation. *Science (80- ).* 2016;352(6285 PG-586-589):586–9.
- Lichtman JS, Ferreyra JA, Ng KM, Smits SA, Sonnenburg JL, Elias JE. Host-Microbiota Interactions in the Pathogenesis of Antibiotic-Associated Diseases. *Cell Rep.* 2016;14(5 PG-1049-1061):1049–61.
- Likotrafiti E, Manderson KS, Fava F, Tuohy KM, Gibson GR, Rastall RA. Molecular identification and anti-pathogenic activities of putative probiotic bacteria isolated from faeces of healthy elderly individuals. *Microb Ecol Health Dis.* 2004;16(2-3-105–112):105–12.
- Lofgren ET, Moehring RW, Anderson DJ, Weber DJ, Fefferman NH. A mathematical model to evaluate the routine use of fecal microbiota transplantation to prevent incident and recurrent *clostridium difficile* infection. *Infect Control Hosp Epidemiol.* 2014;35(1 PG-18-27):18–27.
- Low DE, Shahinas D, Silverman M, Sittler T, Chiu C, Kim P, et al. Toward an understanding of changes in diversity associated with fecal microbiome transplantation based on 16s rRNA gene deep sequencing. *MBio.* 2012;3(5 PG-).
- Luna R, Pitashny M, Runge J, Shang Y, Hollister E, Nagy-Szakal D, et al. Microbiome characterization as a diagnostic tool in fecal microbiome transplantation. *J Mol Diagnostics.* 2013;15 (6)(PG-874-875):874–5.
- Millan B, Hotte N, Mathieu O, Burguiere P, Tompkins TA, Kao D, et al. Effects of fecal microbial transplantation on the gut resistome in patients with recurrent *clostridium difficile* infection. *Gastroenterology.* 2015;1)(PG-S120):S120.
- Millan B, Park H, Hotte N, Fedorak R, Kao D, Madsen K. Antibiotics and bowel preparation enhance the ability of fecal microbial transplantation to reshape the gut microbiota in IL-10<sup>-/-</sup> mice. *Can J Gastroenterol Hepatol Conf.* 2016;(pagination PG-).
- Millan B, Park H, Hotte N, Mathieu O, Burguiere P, Tompkins TA, et al. Fecal Microbial Transplants Reduce Antibiotic-resistant Genes in Patients with Recurrent *Clostridium difficile* Infection. *Clin Infect Dis.* 2016;62(12 PG-1479-1486):1479–86.
- Moelling K, Broecker F. Fecal microbiota transplantation to fight *Clostridium difficile* infections and other intestinal diseases. *Bacteriophage.* 2016;18(PG-).

- Murdoch DA, Gibbs S, Price CGA, Easmon S, Franklin J, Lister TA, et al. Effect of ceftazidime and gentamicin on the oropharyngeal and faecal flora of patients with haematological malignancies. *J Antimicrob Chemother.* 1990;26(3 PG-419-428):419–28.
- Nord CE, Kager L, Philipson A, Stiernstedt G. Impact of imipenem/cilastatin therapy on faecal flora. *Eur J Clin Microbiol.* 1984;3(5 PG-475-477):475–7.
- Peer X, An G. Agent-based model of fecal microbial transplant effect on bile acid metabolism on suppressing clostridium difficile infection: An example of agent-based modeling of intestinal bacterial infection. *J Pharmacokinet Pharmacodyn.* 2014;41(5 PG-493-507):493–507.
- Schenck LP, Hirota S, Armstrong G, MacDonald J, Beck P. Investigating the effect of antibiotics on gut microbiota components and subsequent Clostridium difficile infection. *FASEB Journal Conf Exp Biol.* 2014;28(1 SUPPL. 1 PG-).
- Schenck LP, Hirota SA, Armstrong GD, MacDonald JA, Beck PL. Elucidating intestinal microbiota components that play a protective or deleterious role during clostridium difficile infections. *Gastroenterology.* 2013;1(PG-S185):S185.
- Seekatz AM, Aas J, Gessert CE, Rubin TA, Saman DM, Bakken JS, et al. Recovery of the gut microbiome following fecal microbiota transplantation. *MBio.* 2014;5(3):e00893-14.
- Seekatz AM, Rao K, Santhosh K, Young VB. Dynamics of the fecal microbiome in patients with recurrent and nonrecurrent Clostridium difficile infection. *Genome Med.* 2016;8 (1) (no pagination)(47 PG-).
- Seekatz AM, Theriot CM, Molloy CT, Wozniak KL, Bergin IL, Young VB. Fecal microbiota transplantation eliminates Clostridium difficile in a murine model of relapsing disease. *Infect Immun.* 2015;83(10 PG-3838-3846):3838–46.
- Shanahan F. Separating the microbiome from the hyperbolome. *Genome Med.* 2015;7 (1) (no pagination)(17 PG-).
- Shankar V, Hamilton MJ, Khoruts A, Kilburn A, Unno T, Paliy O, et al. Species and genus level resolution analysis of gut microbiota in Clostridium difficile patients following fecal microbiota transplantation. *Microbiome.* 2014;2 (1) (no pagination)(13 PG-).
- Soma G, Inagawa H. Methods to Prevent or Treat Refractory Diseases by Focusing on Intestinal Microbes Using LPS and Macrophages. *Anticancer Res.* 2015;35(8 PG-4393-6):4393–6.
- Song Y, Garg S, Girotra M, Maddox C, Von Rosenvinge EC, Dutta A, et al. Microbiota dynamics in patients treated with fecal microbiota transplantation for recurrent Clostridium difficile infection. *PLoS One.* 2013;8 (11) (no pagination)(e81330 PG-).
- Song Y, Garg S, Girotra M, Maddox C, Von Rosenvinge EC. Correction: Microbiota dynamics in patients treated with fecal microbiota transplantation for recurrent Clostridium difficile infection (PLoS ONE (2013) 8, 11 (e81330) DOI: 10.1371/journal.pone.0081330). *PLoS One.* 2014;9 (7) (no pagination)(e104471 PG-).
- Spinler JK, Brown A, Ross CL, Boonma P, Conner ME, Savidge TC. Administration of probiotic kefir to mice with Clostridium difficile infection exacerbates disease. *Anaerobe.* 2016;40(PG-54-7):54–7.



- Tian Z, Liu J, Liao M, Li W, Zou J, Han X, et al. Beneficial Effects of Fecal Microbiota Transplantation on Ulcerative Colitis in Mice. *Dig Dis Sci*. 2016;61(8 PG-2262-2271):2262–71.
- Vermeire S, Joossens M, Verbeke K, Wang J, Machiels K, Sabino J, et al. Donor Species Richness Determines Faecal Microbiota Transplantation Success in Inflammatory Bowel Disease. *J Crohns Colitis*. 2016;10(4 PG-387-94):387–94.
- Vermeire S, Joossens M, Verbeke K, Wang J, Machiels K, Sabino J, et al. Donor Species Richness Determines Faecal Microbiota Transplantation Success in Inflammatory Bowel Disease. *J Crohn's Colitis*. 2016;10(4 PG-387-394):387–94.
- Villafuerte-Galvez JA, Patel IJ, Xu H, Yang X, Chen X, Kelly CP. Elevated serum IL-27 concentrations predict adverse outcomes in clostridium difficile associated diarrhea. *Gastroenterology*. 2014;1(PG-S253-S254):S253–4.
- Weingarden A, Gonzalez A, Vazquez-Baeza Y, Weiss S, Humphry G, Berg-Lyons D, et al. Dynamic changes in short- and long-term bacterial composition following fecal microbiota transplantation for recurrent *Clostridium difficile* infection. *Microbiome*. 2015;3 (1) (no pagination)(10 PG-).
- Weingarden AR, Chen C, Bobr A, Yao D, Lu Y, Nelson VM, et al. Microbiota transplantation restores normal fecal bile acid composition in recurrent *Clostridium difficile* infection. *AJP Gastrointest Liver Physiol*. 2014 Feb 15;306(4):G310–9.
- Weingarden AR, Chen C, Zhang N, Graiziger CT, Dosa PI, Steer CJ, et al. Ursodeoxycholic acid inhibits *clostridium difficile* spore germination and vegetative growth, and prevents the recurrence of ileal pouchitis associated with the infection. *J Clin Gastroenterol*. 2016;50(8 PG-624-630):624–30.
- Weingarden AR, Dosa PI, DeWinter E, Steer CJ, Shaughnessy MK, Johnson JR, et al. Changes in colonic bile acid composition following fecal microbiota transplantation are sufficient to control *Clostridium difficile* germination and growth. *PLoS One*. 2016;11 (1) (no pagination)(e0147210 PG-).
- Weingarden AR, Hamilton MJ, Sadowsky MJ, Khoruts A. Resolution of severe *clostridium difficile* infection following sequential fecal microbiota transplantation. *J Clin Gastroenterol*. 2013;47(8 PG-735-737):735–7.

#### D.1.7. Narrative reviews

- Actis GC. The gut microbiome. *Inflamm Allergy - Drug Targets*. 2014;13(4 PG-217-223):217–23.
- Adamu BO, Lawley TD. Bacteriotherapy for the treatment of intestinal dysbiosis caused by *Clostridium difficile* infection. *Curr Opin Microbiol*. 2013;16(5 PG-596-601):596–601.
- Agito MD, Atreja A, Rizk MK. Fecal microbiota transplantation for recurrent *C difficile* infection: Ready for prime time? *Cleve Clin J Med*. 2013;80(2 PG-101-108):101–8.
- Allegretti JR, Hamilton MJ, J.R. A, M.J. H, Allegretti JR, Hamilton MJ. Restoring the gut microbiome for the treatment of inflammatory bowel diseases. *World J Gastroenterol*. M. J. Hamilton, Division of

- Gastroenterology, Brigham and Women's Hospital and Harvard Medical School, 75 Francis St, Boston, MA 02115, United States. E-mail: mjhamilton@partners.org: WJG Press; 2014;20(13):3468–74.
- Allegretti JR, Korzenik JR, Hamilton MJ. Fecal microbiota transplantation via colonoscopy for recurrent *C. difficile* Infection. J Vis Exp. MyJoVE Corporation; 2014 Dec;(94).
- Allegretti JR, Phelps E, Xu H, Kassam Z, Fischer M. Redefining cure in clostridium difficile infection: Clinical assessment 4 weeks after fecal microbiota transplantation is predictive of standard 8-week cure endpoint. Am J Gastroenterol. 2016;111(PG-S56):S56.
- Allen-Vercoe E, Petrof EO. Artificial stool transplantation: Progress towards a safer, more effective and acceptable alternative. Expert Rev Gastroenterol Hepatol. 2013;7(4 PG-291-293):291–3.
- Allen-Vercoe E, Reid G, Viner N, Gloor GB, Hota S, Kim P, et al. A Canadian Working Group report on fecal microbial therapy: microbial ecosystems therapeutics. Can J Gastroenterol. 2012;26(7 PG-457-462):457–62.
- Almeida R, Gerbaba T, Petrof EO. Recurrent *Clostridium difficile* infection and the microbiome. J Gastroenterol. 2016;51(1 PG-1-10):1–10.
- Amirtha T. MICROBIOME RESEARCH. Banking on stool despite an uncertain future. Science (80- ). 2016;352(6291 PG-1261-2):1261–2.
- Anand R, Girotra M, Garg S, Dutta S. Safety and efficacy of fecal microbiota transplantation (FMT) for recurrent clostridium difficile infection (RCDI) in septuagenarians, octogenarians, and nonagenarians: A single-center experience. Am J Gastroenterol. 2014;109(PG-S195):S195.
- Anderson JL, Edney RJ, Whelan K. Systematic review: Faecal microbiota transplantation in the management of inflammatory bowel disease. Aliment Pharmacol Ther. 2012;36(6 PG-503-516):503–16.
- Anonymous. Donor faeces for recurrent *Clostridium difficile* diarrhoea? BMJ. 2013;346 (no pagination)(f376 PG-).
- Anonymous. Faecal microbiota transplantation. Drug Ther Bull. 2014;52(12 PG-141-144):141–4.
- Anonymous. Faecal microbiota transplantation. Drug Ther Bull. 2014;52(12 PG-141-144):141–4.
- Anonymous. Fecal microbiota therapy for *Clostridium difficile* infection: A health technology assessment. Ont Health Technol Assess Ser. 2016;16(17 PG-).
- Anonymous. Fecal microbiota transplantation for treating recurrent *Clostridium difficile* infection. OR Manager. 2012;28(8 PG-15-18):15–8.
- Anonymous. Fecal microbiota transplantation for treating recurrent *Clostridium difficile* infection. Manag Care. 2013;22(6 PG-18-19):18–9.
- Anonymous. Fecal microbiota transplantation for treatment of recurrent *C. difficile* infection. Clin Privil White Pap. 2013;(246 PG-1-15):1–15.



- 1  
2 Anonymous. Fecal microbiota transplantation: Where is it leading? *Gastroenterol Hepatol*. 2014;10(5 PG-  
3 307-309):307–9.  
4  
5  
6 Anonymous. Fecal microbiota transplantation: Where is it leading? *Gastroenterol Hepatol*. 2014;10(5 PG-  
7 307-309):307–9.  
8  
9  
10 Anonymous. Infection: FMT: a safe treatment for *Clostridium difficile* infection in immunocompromised  
11 patients. *Nat Rev Gastroenterol Hepatol*. 2014;1(PG-).  
12  
13 Anonymous. More on faecal microbiota transplantation. *Drug Ther Bull*. 2015;53(7 PG-76-77):76–7.  
14  
15 Anonymous. More on faecal microbiota transplantation. *Drug Ther Bull*. 2015;53(7 PG-76-77):76–7.  
16  
17  
18 Anonymous. Novel therapy for *C. difficile* infections. Infusions of donated feces may help those with  
19 recurrent infections. *Harv Health Lett*. 2011;36(12 PG-7):7.  
20  
21  
22 Anonymous. Probiotics are beneficial in *Clostridium difficile* infection: Healthy microbiota by probiotics or  
23 fecal transplantation prevent diarrhea. [Dutch]. *Pharm Weekbl Wet Platf*. 2014;149(10 PG-).  
24  
25 Anonymous. Solving a *C.difficile* problem. If antibiotics fail, a stool transplant can help cure a severe  
26 infection. *Johns Hopkins Med Lett Health After 50*. 2014;25(13 PG-3):3.  
27  
28  
29 Anonymous. Solving a *C.difficile* problem. If antibiotics fail, a stool transplant can help cure a severe  
30 infection. *Johns Hopkins Med Lett Health After 50*. 2014;25(13 PG-3):3.  
31  
32  
33 Anonymous. Therapy: FMT effective in patients with severe and/or complicated CDI. *Nat Rev Gastroenterol*  
34 *Hepatol*. 2015;21(PG-).  
35  
36 Anonymous. Therapy: FMT effective in patients with severe and/or complicated CDI. *Nat Rev Gastroenterol*  
37 *Hepatol*. 2015;21(PG-).  
38  
39  
40 Anonymous. Treat *Clostridium difficile* infection based on its severity and number of previous episodes.  
41 *Drugs Ther Perspect*. 2012;28(2 PG-10-13):10–3.  
42  
43  
44 Antonopoulos DA, Chang EB. Transplanting a microbial organ: The good, the bad, and the unknown. *MBio*.  
45 2016;7 (3) (no pagination)(e00572-16 PG-).  
46  
47 Apostolescu C, Moroti R, Molagic V, Gheorghite V, Telepan D, Popoiu M, et al. Gut microbiota and its complex  
48 role. The experience of the national institute for infectious diseases Prof. Dr. Matei Bals in fecal  
49 bacteriotherapy for *Clostridium difficile* infection. *BMC Infect Dis Conf 9th Ed Sci Days Natl Inst Infect Dis*  
50 “Prof Dr Matei Bals” Rom Conf Start. 2013;13(no pagination PG-).  
51  
52  
53 Aroniadis OC, Brandt LJ, Greenberg A, Borody TJ, Kelly C, Mellow M, et al. Long-term follow-up study of fecal  
54 microbiota transplantation (FMT) for severe or complicated *clostridium difficile* infection (CDI).  
55 *Gastroenterology*. 2013;1(PG-S185):S185.  
56  
57  
58 Aroniadis OC, Brandt LJ. Fecal microbiota transplantation: past, present and future. *Curr Opin Gastroenterol*.  
59 2013;29(1):79–84.  
60

- 1  
2 Aroniadis OC, Brandt LJ. Fecal microbiota transplantation: past, present and future. *Curr Opin Gastroenterol*.  
3 2013;29(1):79–84.  
4  
5  
6 Aroniadis OC, Brandt LJ. Intestinal microbiota and the efficacy of fecal microbiota transplantation in  
7 gastrointestinal disease. *Gastroenterol Hepatol*. 2014;10(4 PG-230-237):230–7.  
8  
9  
10 Austin M, Mellow M, Tierney WM. Fecal microbiota transplantation in the treatment of clostridium difficile  
11 infections. *Am J Med*. 2014;127(6 PG-479-483):479–83.  
12  
13 Avery L, Hasan M. Fecal bacteriotherapy for clostridium difficile infections - its time has come. *Clin Microbiol*  
14 *Newsl*. 2013;35(15 PG-119-124):119–24.  
15  
16  
17 Badger VO, Ledebore NA, Graham MB, Edmiston CE. Clostridium difficile: Epidemiology, pathogenesis,  
18 management, and prevention of a recalcitrant healthcare-associated pathogen. *J Parenter Enter Nutr*.  
19 2012;36(6 PG-645-662):645–62.  
20  
21  
22 Bafeta A, Yavchitz A, Riveros C, Batista R, Ravaud P. Methods and Reporting Studies Assessing Fecal  
23 Microbiota Transplantation: A Systematic Review. *Ann Intern Med*. 2017;167(1):34–9.  
24  
25  
26 Bagdasarian, N.; Rao, K.; Malani, P. N. Diagnosis and treatment of clostridium difficile in adults: A  
27 systematic review. *Journal of the American Medical Association* 27 Jan 2015;313(4):398–408  
28  
29 Bakken JS. Fecal bacteriotherapy for recurrent Clostridium difficile infection. *Anaerobe*. 2009;15(6):285–9.  
30  
31  
32 Bakken JS. Feces transplantation for recurrent Clostridium difficile infection: US experience and  
33 recommendations. *Microb Ecol Heal Dis*. 2015;26(PG-27657):27657.  
34  
35  
36 Bakken JS. Staggered and tapered antibiotic withdrawal with administration of kefir for recurrent Clostridium  
37 difficile infection. *Clin Infect Dis*. 2014;59(6 PG-858-861):858–61.  
38  
39 Balzola F, Cullen G, Ho GT, Russell R. Fecal microbiota transplantation: Indication, methods, evidence and  
40 further directions: Commentary. *Inflamm Bowel Dis Monit*. 2014;14(2 PG-56-57):56–7.  
41  
42 Barbut F, Collignon A, Butel MJ, Bourlioux P. [Fecal microbiota transplantation: review]. *Ann Pharm Fr*.  
43 2015;73(1 PG-13-21):13–21.  
44  
45  
46 Barbut F, Collignon A, Butel MJ, Bourlioux P. Fecal microbiota transplantation: Review. [French]. *Ann Pharm*  
47 *Fr*. 2015;73(1 PG-13-21):13–21.  
48  
49  
50 Barbut F, Guery B, Eckert C. How to treat Clostridium difficile infections in 2014?. [French]. *Reanimation*.  
51 2014;23(3 PG-284-297):284–97.  
52  
53  
54 Barbut F. Alleviating the burden of CDI: Current and emerging treatment options. *Int J Antimicrob Agents*.  
55 2013;42(PG-S11):S11.  
56  
57 Barnes D, Park KT. Donor Considerations in Fecal Microbiota Transplantation. *Current Gastroenterology*  
58 *Reports*. 2017;19 (3) (no pagination)(10).  
59  
60

- Baron TH, Kozarek RA. Fecal microbiota transplant: We know its history, but can we predict its future? *Mayo Clin Proc.* 2013;88(8 PG-782-785):782–5.
- Bartnicka A, Szachta P, Galecka M. Faecal microbiota transplant - prospects and safety. *Pomeranian J life Sci.* 2015;61(3 PG-282-286):282–6.
- Batista R, Kapel N, Megerlin F, Chaumeil JC, Barbut F, Bourlioux P, et al. [Fecal microbiota transplantation in recurrent *Clostridium difficile* infections. Framework and pharmaceutical preparation aspects]. *Ann Pharm Fr.* 2015;73(5 PG-323-31):323–31.
- Batista R, Kapel N, Megerlin F, Chaumeil JC, Barbut F, Bourlioux P, et al. Fecal microbiota transplantation in recurrent *Clostridium difficile* infections. Framework and pharmaceutical preparation aspects. [French]. *Ann Pharm Fr.* 2015;73(5 PG-323-331):323–31.
- Bauer MP, van Dissel JT. Alternative strategies for *Clostridium difficile* infection. *Int J Antimicrob Agents.* 2009;33(SUPPL. 1 PG-S51-S56):S51–6.
- Baxter M, Colville A. Adverse events in faecal microbiota transplant: a review of the literature. *J Hosp Infect.* 2016 Feb;92(2):117–27.
- Benes J, Husa P, Nyc O, Polivkova S. [Diagnosis and therapy of *Clostridium difficile* infection: Czech national guidelines]. *Klin Mikrobiol Infekc Lek.* 2014;20(2 PG-56-66):56–66.
- Benes J, Husa P, Nyc O, Polivkova S. Diagnosis and therapy of *Clostridium difficile* infection: Czech national guidelines. [Croatian]. *Klin Mikrobiol Infekc Lek.* 2014;20(2 PG-56-66):56–66.
- Benes J, Husa P, Nyc O. [Recommendations for diagnosis and therapy of colitis caused by *Clostridium difficile*]. *Klin Mikrobiol Infekc Lek.* 2012;18(5 PG-160-7):160–7.
- Benes J, Husa P, Nyc O. [Recommendations for diagnosis and therapy of colitis caused by *Clostridium difficile*]. [Czech]. *Klin Mikrobiol Infekc Lek.* 2012;18(5 PG-160-167):160–7.
- Berg AM, Farraye FA. Duodenal infusion of stool is more effective than vancomycin in patients with recurrent *Clostridium difficile*. *Evid Based Med.* 2013;18(6 PG-220-221):220–1.
- Berg D, Clemente JC, Colombel JF. Can inflammatory bowel disease be permanently treated with short-term interventions on the microbiome? *Expert Rev Gastroenterol Hepatol.* 2015;9(6 PG-781-795):781–95.
- Biehl L. Fecal microbiota transfer. *Transfusion Medicine and Hemotherapy.* 2017;44 (Supplement 1):22.
- Biltaji E, Varier R, Smith K, Roberts M, Lafleur J, Nelson RE. Cost-effectiveness analysis of treatment strategies for initial *clostridium difficile* infection. *Value Heal.* 2014;17 (3)(PG-A38):A38.
- Blackburn LM, Bales A, Caldwell M, Cordell L, Hamilton S, Kreider H. Fecal microbiota transplantation in patients with cancer undergoing treatment. *Clin J Oncol Nurs.* 2015;19(1 PG-111-4):111–4.
- Bloukh SI. *Clostridium difficile* infection: An overview of the disease and its pathogenesis, diagnosis, treatment, prevention and management. *Res J Pharm Biol Chem Sci.* 2013;4(4 PG-1219-1232):1219–32.

- Bojanova DP, Bordenstein SR. Fecal Transplants: What Is Being Transferred? PLoS Biol. 2016;14 (7) (no pagination)(e1002503 PG-).
- Bookstaver PB, Ahmed Y, Millisor VE, Siddiqui W, Albrecht H. Clostridium difficile: case report and concise review of fecal microbiota transplantation. J S C Med Assoc. 2013;109(2 PG-62-66):62–6.
- Borgia G, Maraolo AE, Foggia M, Buonomo AR, Gentile I. Fecal microbiota transplantation for Clostridium difficile infection: Back to the future. Expert Opin Biol Ther. 2015;15(7 PG-1001-1014):1001–14.
- Borody T, Fischer M, Mitchell S, Campbell J. Fecal microbiota transplantation in gastrointestinal disease: 2015 update and the road ahead. Expert Rev Gastroenterol Hepatol. 2015;9(11 PG-1379-1391):1379–91.
- Borody T, Torres M, Campbell J, Leis S, Nowak A. Reversal of inflammatory bowel disease (IBD) with recurrent faecal microbiota transplants (FMT). Am J Gastroenterol. 2011;106(PG-S366):S366.
- Borody T, Wettstein A, Campbell J, Leis S, Torres M, Finlayson S, et al. Fecal microbiota transplantation in ulcerative colitis: Review of 24 years experience. Am J Gastroenterol. 2012;107(PG-S665):S665.
- Borody TJ, Brandt LJ, Paramsothy S, Agrawal G. Fecal microbiota transplantation: A new standard treatment option for Clostridium difficile infection. Expert Rev Anti Infect Ther. 2013;11(5 PG-447-449):447–9.
- Borody TJ, Brandt LJ, Paramsothy S. Therapeutic faecal microbiota transplantation: Current status and future developments. Curr Opin Gastroenterol. 2014;30(1 PG-97-105):97–105.
- Borody TJ, Brandt LJ, Paramsothy S. Therapeutic faecal microbiota transplantation: Current status and future developments. Curr Opin Gastroenterol. 2013;19(PG-).
- Borody TJ, Campbell J. Fecal microbiota transplantation: Current status and future directions. Expert Rev Gastroenterol Hepatol. 2011;5(6 PG-653-655):653–5.
- Borody TJ, Campbell J. Fecal Microbiota Transplantation. Techniques, Applications, and Issues. Gastroenterol Clin North Am. 2012;41(4):781–803.
- Borody TJ, Campbell J. Fecal Microbiota Transplantation. Techniques, Applications, and Issues. Gastroenterol Clin North Am. 2012;41(4):781–803.
- Borody TJ, Connelly N, Mitchell SW. Fecal microbiota transplantation in gastrointestinal diseases: What practicing physicians should know. Pol Arch Med Wewn. 2015;125(11 PG-852-858):852–8.
- Borody TJ, Finlayson S, Paramsothy S. Is Crohn's disease ready for fecal Microbiota transplantation? J Clin Gastroenterol. 2014;48(7 PG-582-583):582–3.
- Borody TJ, Finlayson S. Fecal microbiota transplantation for Clostridium difficile infection: A surgeon's perspective. Semin Colon Rectal Surg. 2014;25(3 PG-163-166):163–6.
- Borody TJ, Khoruts A. Fecal microbiota transplantation and emerging applications. Nat Rev Gastroenterol Hepatol. 2012;9(2 PG-88-96):88–96.

- Borody TJ, Khoruts A. Fecal microbiota transplantation and emerging applications. *Nat Rev Gastroenterol Hepatol*. 2011;20(PG-).
- Borody TJ, Paramsothy S, Agrawal G. Fecal microbiota transplantation: Indications, methods, evidence, and future directions. *Curr Gastroenterol Rep*. 2013;15(8):1–7.
- Borody TJ, Peattie D, Kapur A. Could fecal microbiota transplantation cure all *Clostridium difficile* infections? *Future Microbiol*. 2014;9(1 PG-1-3):1–3.
- Borody TJ, Peattie D, Mitchell SW. Fecal microbiota transplantation: Expanding horizons for *Clostridium difficile* infections and beyond. *Antibiotics*. 2015;4(3 PG-254-266):254–66.
- Borody TJ, Warren EF, Leis SM, Surace R, Ashman O, Siarakas S. Bacteriotherapy using fecal flora: toying with human motions. *J Clin Gastroenterol*. 2004;38(6):475–83.
- Borody TJ, Warren EF, Leis SM, Surace R, Ashman O, Siarakas S. Bacteriotherapy using fecal flora: toying with human motions. *J Clin Gastroenterol*. 2004;38(6):475–83.
- Bourlioux P, workgroup of the French Academy of P. Faecal microbiota transplantation: Key points to consider. *Ann Pharm Fr*. 2015;73(3 PG-163-8):163–8.
- Bourlioux P. Faecal microbiota transplantation: Key points to consider. *Ann Pharm Fr*. 2015;73(3 PG-163-168):163–8.
- Bowman KA, Broussard EK, Surawicz CM. Fecal microbiota transplantation: Current clinical efficacy and future prospects. *Clin Exp Gastroenterol*. 2015;8(PG-285-291):285–91.
- Boyle ML, Ruth-Sahd LA, Zhou Z. Fecal microbiota transplant to treat recurrent *Clostridium difficile* infections. *Crit Care Nurse*. 2015;35(2 PG-51-64; quiz 65):51–64; quiz 65.
- Brandt LJ, Aroniadis OC. An overview of fecal microbiota transplantation: Techniques, indications, and outcomes. *Gastrointest Endosc*. 2013;78(2 PG-240-249):240–9.
- Brandt LJ, Borody TJ, Campbell J. Endoscopic fecal microbiota transplantation: “First-line” treatment for severe *clostridium difficile* infection? *J Clin Gastroenterol*. 2011;45(8 PG-655-657):655–7.
- Brandt LJ, Reddy SS. Fecal microbiota transplantation for recurrent *Clostridium difficile* infection. *J Clin Gastroenterol*. 2011;45(SUPPL. 3 PG-S159-S167):S159–67.
- Brandt LJ. Fecal microbiota transplant Respite, Adspice, Prospice. *J Clin Gastroenterol*. 2015;49(PG-S65-S68):S65–8.
- Brandt LJ. Fecal transplantation for the treatment of *Clostridium difficile* infection. *Gastroenterol Hepatol*. 2012;8(3 PG-191-194):191–4.
- Brandt LJ. FMT: First step in a long journey. *Am J Gastroenterol*. 2013;108(8 PG-1367-1368):1367–8.
- Brezina J, Bajer L, Spicak J, Draatich P. Faecal microbial transplantation in inflammatory bowel disease. [Czech]. *Gastroenterol a Hepatol*. 2016;70(1 PG-51-56):51–6.

- Bridges E, McNeill M, Munro N. Research in Review: Driving Critical Care Practice Change. *Am J Crit Care*. 2016;25(1 PG-76-84):76–84.
- Broecker F, Klumpp J, Moelling K. Long-term microbiota and virome in a Zurich patient after fecal transplantation against *Clostridium difficile* infection. *Ann New York Acad Sci*. 2016;(PG-).
- Brown WR. Fecal microbiota transplantation in treating *Clostridium difficile* infection. *J Dig Dis*. 2014;15(8 PG-405-408):405–8.
- Burke KE, Lamont JT. Fecal transplantation for recurrent *clostridium difficile* infection in older adults: A review. *J Am Geriatr Soc*. 2013;61(8 PG-1394-1398):1394–8.
- Burton, H. E.; Mitchell, S. A.; Watt, M. The cost effectiveness of treatments for *clostridium difficile* infection: A systematic review. *Value in Health* May 2016;19 (3):A218
- Cammarota G, Ianiro G, Bibbo S, Gasbarrini A. Fecal microbiota transplantation a new old kid on the block for the management of gut microbiota-related disease. *J Clin Gastroenterol*. 2014;48(PG-S80-S84):S80–4.
- Cammarota G, Ianiro G, Bibbo S, Gasbarrini A. Gut microbiota modulation: probiotics, antibiotics or fecal microbiota transplantation? *Intern Emerg Med*. 2014;9(4 PG-365-373):365–73.
- Cammarota, G.; Ianiro, G.; Gasbarrini, A. Fecal microbiota transplantation for the treatment of *clostridium difficile* infection: A systematic review *Journal of Clinical Gastroenterology* September 2014;48(8):693-702
- Cammarota G, Ianiro G, Tilg H, Rajilic-Stojanovic M, Kump P, Satokari R, et al. European consensus conference on faecal microbiota transplantation in clinical practice. *Gut*. 2017;66(4):569-80.
- Carlucci C, Petrof EO, Allen-Vercoe E. Fecal Microbiota-based Therapeutics for Recurrent *Clostridium difficile* Infection, Ulcerative Colitis and Obesity. *EBioMedicine*. 2016;13(PG-37-45):37–45.
- Carstensen JW, Hansen AK. [Faecal transplantation as a treatment for *Clostridium difficile* infection, ulcerative colitis and the metabolic syndrome]. *Ugeskr Laeger*. 2014;176(4 PG-17):17.
- Chapman, B. C.; Moore, H. B.; Overbey, D. M.; Morton, A. P.; Harnke, B.; Gerich, M. E.; Vogel, J. D. Fecal microbiota transplant in patients with *Clostridium difficile* infection: A systematic review *Journal of Trauma and Acute Care Surgery* 01 Oct 2016;81(4):756-764
- Chen B, Avinashi V, Dobson S. Fecal microbiota transplantation for recurrent *Clostridium difficile* infection in children. *Journal of Infection*. 2017;74:S120-S7.
- Choi HH, Cho YS. Fecal Microbiota Transplantation: Current Applications, Effectiveness, and Future Perspectives. *Clin Endosc*. 2016;49(3 PG-257-65):257–65.
- Clostridium difficile* Infection in Older Adults: Systematic Review of Efforts to Reduce Occurrence and Improve Outcomes
- Cohen NA, Ami RB, Guzner-Gur H, Santo ME, Halpern Z, Maharshak N. Fecal microbiota transplantation for *Clostridium difficile*-associated diarrhea. *Isr Med Assoc J*. 2015;17(8 PG-510-514):510–4.



- Cohen NA, Ben Ami R, Guzner-Gur H, Santo ME, Halpern Z, Maharshak N. Fecal Microbiota Transplantation for *Clostridium difficile*-Associated Diarrhea. *Isr Med Assoc J Imaj*. 2015;17(8 PG-510-4):510–4.
- Colonoscopic fecal bacteriotherapy in the treatment of recurrent *Clostridium difficile* infection--results and follow-up
- Colonoscopic versus nasogastric fecal transplantation for the treatment of *Clostridium difficile* infection: A review and pooled analysis .Consensus report: faecal microbiota transfer - clinical applications and procedures
- Consultant Pharmacist Jan 01 2017;32(1):24-41
- Cramer JP. [Infusion of donor feces in recurrent *Clostridium difficile* infection? - Infusion of donor feces: Promising intervention with several question marks]. *Dtsch Medizinische Wochenschrift*. 2013;138(12 PG-566):566.
- Cramer JP. Infusion of donor feces in recurrent *Clostridium difficile* infection? - Infusion of donor feces: Promising intervention with several question marks. [German]. *Dtsch Medizinische Wochenschrift*. 2013;138(12 PG-566):566.
- Crow JR, Davis SL, Chaykosky DM, Smith TT, Smith JM. Probiotics and fecal microbiota transplant for primary and secondary prevention of *clostridium difficile* infection. *Pharmacotherapy*. 2015;35(11 PG-1016-1025):1016–25.
- Crum-Cianflone NF, Sullivan E, Ballon-Landa G. Fecal microbiota transplantation and successful resolution of multidrug-resistant-organism colonization. *J Clin Microbiol*. 2015;53(6 PG-1986-1989):1986–9.
- Cui BT, Wang M, Ji GZ, Fan ZN, Zhang FM. Fecal microbiota transplantation: From the 4<sup>th</sup> century to 2013. [Chinese]. *World Chinese J Dig*. 2013;21(30 PG-3222-3229):3222–9.
- Dai C, Jiang M, Sun MJ. Fecal microbiota transplantation for treatment of *clostridium difficile* infection. *J Clin Gastroenterol*. 2015;49(2 PG-171-172):171–2.
- Dai T, Tang T. [Research progress of fecal microbiota transplantation]. *Zhonghua Weichang Waiké Zazhi*. 2015;18(7 PG-733-7):733–7.
- Dai T, Tang T. Research progress of fecal microbiota transplantation. [Chinese]. *Zhonghua Wei Chang Wai Ke Za Zhi*. 2015;18(7 PG-733-737):733–7.
- Dakhoul, L.; Parikh, K.; Berkelhammer, C. Fecal microbiota transplant in treatment of *clostridium difficile* colitis-pooled data analysis and a systematic review *Gastroenterology* April 2015;1()():S404
- Daloso V, Minacori R, Refolo P, Sacchini D, Craxi L, Gasbarrini A, et al. Ethical aspects of fecal microbiota transplantation (fmt). *Eur Rev Med Pharmacol Sci*. 2015;19(17 PG-3173-3180):3173–80.
- Damman CJ, Miller SI, Surawicz CM, Zisman TL. The microbiome and inflammatory bowel disease: is there a therapeutic role for fecal microbiota transplantation? *Am J Gastroenterol*. 2012;107(10):1452–9.
- Davidovics ZH, Hyams JS. Fecal transplantation: Re-discovering the value of stool. *Curr Opin Pediatr*. 2013;25(5 PG-618-623):618–23.

- Davidovics ZH, Sylvester FA. Medical stool: The future of treatment for inflammatory bowel disease? *J Pediatr Gastroenterol Nutr*. 2013;56(6 PG-583):583.
- De Vos WM. Fame and future of faecal transplantations - developing next-generation therapies with synthetic microbiomes. *Microb Biotechnol*. 2013;6(4 PG-316-325):316–25.
- Debast SB, Bauer MP, Kuijper EJ. European Society of Clinical Microbiology and Infectious Diseases: Update of the Treatment Guidance Document for *Clostridium difficile* Infection. *Clin Microbiol Infect*. 2014 Mar;20(s2):1–26.
- Debast, S. B.; Bauer, M. P.; Kuijper, E. J.; Allerberger, F.; Bouza, E.; Coia, J. E.; Cornely, O. A.; Fitzpatrick, F.; Guery, B.; Wilcox, M.; Nathwani, D.; Noren, T.; Olesen, B.; Rakoczi, E.; Welte, T.; Widmer, A. F. European society of clinical microbiology and infectious diseases: Update of the treatment guidance document for *Clostridium difficile* infection *Clinical Microbiology and Infection* 2014;20(S2):1-26
- Di Bella S, Drapeau C, Garcia-Almodovar E, Petrosillo N. Fecal microbiota transplantation: the state of the art. *Infect Dis Rep*. 2013;5(2 PG-e13):e13.
- Di Bella S, Gouliouris T, Petrosillo N. Fecal microbiota transplantation (FMT) for *Clostridium difficile* infection: Focus on immunocompromised patients. *J Infect Chemother*. 2015;21(4 PG-230-237):230–7.
- Dickinson B, Surawicz CM. Infectious Diarrhea: An Overview. *Curr Gastroenterol Rep*. 2014;16(8 PG-).
- Dickson I. Therapy: Sterile faecal transfer for *C. difficile* infection. *Nat Rev Gastroenterol Hepatol*. 2017;14(1 PG-4):4.
- Dickson I. Therapy: Sterile faecal transfer for *C. difficile* infection. *Nat Rev Gastroenterol Hepatol*. 2016;14(PG-).
- Dodin, M.; Katz, D. E.. Faecal microbiota transplantation for *Clostridium difficile* infection. *International Journal of Clinical Practice* March 2014;68(3):363-368
- Dougherty T, Taneja S, Borum ML. More than just *clostridium difficile* infection: The potential role of fecal microbiota transplantation in patients with inflammatory bowel disease. *Am J Gastroenterol*. 2016;111(PG-S833):S833.
- Drekonja, D.; Reich, J.; Gezahegn, S.; Greer, N.; Shaukat, A.; MacDonald, R.; Rutks, I.; Wilt, T. J. Fecal microbiota transplantation for *clostridium difficile* infection a systematic review *Annals of Internal Medicine* 05 May 2015;162(9):630-638
- Edelstein CA, Kassam Z, Daw J, Smith MB, Kelly CR. The regulation of fecal microbiota for transplantation: An international perspective for policy and public health. *Clin Res Regul Aff. Taylor & Francis*; 2015 Jul;32(3):99–107.
- Edmond MB. The Power of Poop: Fecal Microbiota Transplantation for *Clostridium Difficile* Infection. *Trans Am Clin Climatol Assoc*. 2016;127(PG-71-80):71–80.
- El-Matary W, Simpson R, Ricketts-Burns N. Fecal microbiota transplantation: Are we opening a can of worms? *Gastroenterology*. 2012;143(2 PG-e19):e19.



- El-Matary W. Fecal microbiota transplantation: Long-term safety issues. *Am J Gastroenterol*. 2013;108(9 PG-1537-1538):1537–8.
- Elopre L, Rodriguez M. Fecal microbiota therapy for recurrent *Clostridium difficile* infection in HIV-infected persons. *Ann Intern Med*. 2013;158(10 PG-779-780):779–80.
- Esposito S, Umbrello G, Castellazzi L, Principi N. Treatment of *Clostridium difficile* infection in pediatric patients. *Expert Rev Gastroenterol Hepatol*. 2015;9(6 PG-747-755):747–55.
- Ettinger G, Burton JP, Reid G. If microbial ecosystem therapy can change your life, what's the problem? *BioEssays*. 2013;35(6 PG-508-512):508–12.
- Evrensel A, Ceylan ME. Fecal microbiota transplantation and its usage in neuropsychiatric disorders. *Clin Psychopharmacol Neurosci*. 2016;14(3 PG-231-237):231–7.
- Faecal microbiota transplantation in recurrent *Clostridium difficile* infection: Recommendations from the French Group of Faecal microbiota Transplantation. *Dig Liver Dis*. W.B. Saunders; 2016 Mar;48(3):242–7.
- Famularo G, Trinchieri V, De Simone C. Fecal bacteriotherapy or probiotics for the treatment of intestinal diseases? *Am J Gastroenterol*. 2001;96(7 PG-2262-4):2262–4.
- Rossen NG, MacDonald JK, de Vries EM, *et al*. Fecal microbiota transplantation as novel therapy in gastroenterology: a systematic review
- Fecal microbiota transplantation via nasogastric tube for recurrent *clostridium difficile* infection in pediatric patients
- Ferre Aracil C, Aguilera Castro L, Rodriguez de Santiago E, Garcia Garcia de Paredes A, Lopez San Roman A. Fecal microbiota transplantation - something more than merely a therapeutic curiosity. *Rev Esp Enfermedades Dig*. 2015;107(PG-19):19.
- Ferre-Aracil C, Aguilera-Castro L, Rodriguez-de-Santiago E, Garcia-Garcia-de-Paredes A, Lopez-Sanroman A. Fecal microbiota transplantation - something more than merely a therapeutic curiosity. *Rev Esp Enfermedades Dig*. 2015;107(7 PG-399-401):399–401.
- Floch MH. Editorial: Fecal bacteriotherapy, fecal transplant, and the microbiome. *J Clin Gastroenterol*. 2010;44(8 PG-529-530):529–30.
- Floch MH. Fecal bacteriotherapy, fecal transplant, and the microbiome. *J Clin Gastroenterol*. 2010;44(8 PG-529-30):529–30.
- Floch MH. The power of poop: Probiotics and fecal microbial transplant. *J Clin Gastroenterol*. 2012;46(8 PG-625-626):625–6.
- Fredericks. Sages clinical guidelines for Faecal Microbiota Transplantation (FMT). *South African Gastroenterol Rev*. 2015;13(3 PG-27):27.

- Friedman-Moraco RJ, Mehta AK, Lyon GM, Kraft CS. Fecal microbiota transplantation for refractory *Clostridium difficile* colitis in solid organ transplant recipients. *Am J Transplant*. 2014;14(2 PG-477-480):477–80.
- Fu N, Wong T. *Clostridium difficile* Infection in Patients with Inflammatory Bowel Disease. *Curr Infect Dis Rep*. 2016;18 (6) (no pagination)(19 PG-).
- Fuentes S, de Vos WM. How to Manipulate the Microbiota: Fecal Microbiota Transplantation. *Adv Exp Med Biol*. 2016;902(PG-143-53):143–53.
- Fuentes S, De Vos WM. How to manipulate the microbiota: Fecal microbiota transplantation. E-mail: barbara.b.bertram@gsk.com: Springer New York LLC; 2016;(902 PG-143-153):143–53.
- Fuessl HS. Fecal microbiota transplantation helps in *Clostridium difficile* colitis: Commentary. [German]. *MMW-Fortschritte der Medizin*. 2012;154(17 PG-38):38.
- Fumery M, Corcos O, Kapel N, Stefanescu C, Thomas M, Joly F. Interest and techniques of fecal transplantation. [French, English]. *J des Anti-Infectieux*. 2013;15(4 PG-187-192):187–92.
- Furuya-Kanamori L, Doi SAR, Paterson DL, Helms SK, Yakob L, McKenzie SJ, et al. Upper Versus Lower Gastrointestinal Delivery for Transplantation of Fecal Microbiota in Recurrent or Refractory *Clostridium difficile* Infection: A Collaborative Analysis of Individual Patient Data From 14 Studies. *J Clin Gastroenterol*. 2016;11(PG-).
- Gallo A, Passaro G, Gasbarrini A, Landolfi R, Montalto M. Modulation of microbiota as treatment for intestinal inflammatory disorders: An uptodate. *World J Gastroenterol*. 2016;22(32 PG-7186-7202):7186–202.
- Garcia-Garcia-de-Paredes A, Rodriguez-de-Santiago E, Aguilera-Castro L, Ferre-Aracil C, Lopez-Sanroman A. [Fecal microbiota transplantation]. *Gastroenterol Hepatol*. 2015;38(3 PG-123-34):123–34.
- Gens KD, Elshaboury RH, Holt JS. Fecal microbiota transplantation and emerging treatments for *Clostridium difficile* infection. *J Pharm Pract*. 2013;26(5 PG-498-505):498–505.
- Gianotti RJ, Moss AC. The Use and Efficacy of Fecal Microbiota Transplantation for Refractory *Clostridium difficile* in Patients with Inflammatory Bowel Disease. *Inflamm Bowel Dis*. 2016;22(11 PG-2704-2710):2704–10.
- Giordani A, Bove V, Cianchi D. Nursing skills for management of fecal microbiota transplantation in pediatric patient with *clostridium difficile* infection. *Transpl Int*. 2015;28(PG-427):427.
- Goldberg EJ, Bhalodia S, Jacob S, Patel H, Trinh K V, Varghese B, et al. *Clostridium difficile* infection: A brief update on emerging therapies. *Am J Heal Pharm*. 2015;72(12 PG-1007-1012):1007–12.
- Goldenberg SD. Faecal microbiota transplantation for recurrent *Clostridium difficile* infection and beyond: Risks and regulation. *J Hosp Infect*. 2016;92(2 PG-115-116):115–6.
- Gough E, Shaikh H, Manges AR. Systematic review of intestinal microbiota transplantation (fecal bacteriotherapy) for recurrent *Clostridium difficile* infection. *Clin Infect Dis*. 2011;53(10):994–1002.

- Grady NG, Petrof EO, Claud EC. Microbial therapeutic interventions. *Semin Fetal Neonatal Med.* 2016;21(6 PG-418-423):418–23.
- Granitto MH, Norton CK. Fecal microbiota transplantation in recurrent *C. difficile* infection. *Nurs Crit Care.* 2016;11(1 PG-25-30):25–30.
- Grinspan AM, Kelly CR. Fecal Microbiota Transplantation for Ulcerative Colitis: Not Just Yet. *Gastroenterology.* 2015 Jul;149(1):15–8.
- Groen AK, Nieuwdorp M. An evaluation of the therapeutic potential of fecal microbiota transplantation to treat infectious and metabolic diseases. *EMBO Mol Med.* 2017;9(1 PG-1-3):1–3.
- Gross M, Meyer C. [Stool transplantation for relapsing *Cl. difficile* colitis]. *Z Gastroenterol.* 2013;51(12 PG-1441-3):1441–3.
- Gross M, Meyer C. Stool transplantation for relapsing *C. difficile* colitis. [German]. *Z Gastroenterol.* 2013;51(12 PG-1441-1443):1441–3.
- Guo WT, Dong LN, Wang JP, Liu P. New advances in clinical application of fecal microbiota transplantation. [Chinese]. *World Chinese J Dig.* 2014;22(30 PG-4593-4598):4593–8.
- Guo WT, Wang JP, Liu P, Dong LN. New advances in clinical application of fecal microbiota transplantation. *J Dig Dis.* 2014;15(PG-118-119):118–9.
- Guo, B.; Harstall, C.; Louie, T.; Veldhuyzen van Zanten, S.; Dieleman, L. A. Systematic review: faecal transplantation for the treatment of *Clostridium difficile*-associated disease. *Aliment Pharmacol Ther* 2012;35(8):865-875.
- Gupta S, Allen-Vercoe E, Petrof EO. Fecal microbiota transplantation: In perspective. *Therap Adv Gastroenterol.* 2016;9(2 PG-229-239):229–39.
- Gutierrez-Delgado, E. M.; Garza-Gonzalez, E.; Martinez-Vazquez, M. A.; Gonzalez-Gonzalez, J. A.; Maldonado-Garza, H. J.; Mendoza-Olazarán, S.; Hernandez-Balboa, C. L.; Camacho-Ortiz, A. Fecal transplant for *Clostridium difficile* infection relapses using "pooled" frozen feces from non-related donors *Acta Gastro-Enterologica Belgica* 01 Jun 2016;79(2):268-270
- Hammad TA, Khan MA, Srouf K, Abdelfattah T, Alastal Y, Lee WM, et al. Efficacy and safety of oral, capsulized, frozen fecal microbiota transplantation for recurrent *clostridium difficile* infection. A systematic review and meta-analysis. *Gastroenterology.* 2017;152 (5 Supplement 1):S346.
- Han S, Shannahan S, Pellish R. Fecal microbiota transplant: Treatment options for *clostridium difficile* infection in the intensive care unit. *J Intensive Care Med.* 2016;31(9 PG-577-586):577–86.
- Hashash JG, Binion DG. Managing *Clostridium difficile* in Inflammatory Bowel Disease (IBD). *Curr Gastroenterol Rep.* 2014;16 (7) (no pagination)(393 PG-).
- Health Quality, Ontario Fecal Microbiota Therapy for *Clostridium difficile* Infection: A Health Technology Assessment

- Hebbard AI, Slavin MA, Reed C, Teh BW, Thursky KA, Trubiano JA, et al. The epidemiology of *Clostridium difficile* infection in patients with cancer. *Expert Rev Antiinfective Ther*. 2016;14(11 PG-1077-1085):1077–85.
- Hebbard AIT, Slavin MA, Reed C, Teh BW, Thursky KA, Trubiano JA, et al. The epidemiology of *Clostridium difficile* infection in patients with cancer. *Expert Rev Anti Infect Ther*. 2016;14(11 PG-1077-1085):1077–85.
- Holmes E, Wijeyesekera A, Taylor-Robinson SD, Nicholson JK. The promise of metabolic phenotyping in gastroenterology and hepatology. *Nat Rev Gastroenterol Hepatol*. 2015;12(8 PG-458-471):458–71.
- Honda H, Dubberke ER. *Clostridium difficile* infection in solid organ transplant recipients. *Curr Opin Infect Dis*. 2014;27(4 PG-336-341):336–41.
- Honda H, Dubberke ER. The changing epidemiology of *Clostridium difficile* infection. *Curr Opin Gastroenterol*. 2014;30(1):54–62.
- Hookman P, Barkin JS. *Clostridium difficile* associated infection, diarrhea and colitis. *World J Gastroenterol*. 2009;15(13 PG-1554-1580):1554–80.
- Hourigan SK, Oliva-Hemker M. Fecal microbiota transplantation in children: A brief review. *Pediatr Res*. 2016;80(1 PG-2-6):2–6.
- Hryckowian AJ, Pruss KM, Sonnenburg JL. The emerging metabolic view of *Clostridium difficile* pathogenesis. *Curr Opin Microbiol*. 2017;35(PG-42-47):42–7.
- Hryckowian AJ, Pruss KM, Sonnenburg JL. The emerging metabolic view of *Clostridium difficile* pathogenesis. *Curr Opin Microbiol*. 2016;35(PG-42-47):42–7.
- Hudson LE, Anderson SE, Corbett AH, Lamb TJ. Gleaning Insights from Fecal Microbiota Transplantation and Probiotic Studies for the Rational Design of Combination Microbial Therapies. *Clin Microbiol Rev*. 2017;30(1):191-231.
- Hudson LE, Anderson SE, Corbett AH, Lamb TJ. Gleaning insights from fecal microbiota transplantation and probiotic studies for the rational design of combination microbial therapies. *Clin Microbiol Rev*. 2017;30(1 PG-191-231):191–231.
- Huebner ES, Surawicz CM. Treatment of recurrent *Clostridium difficile* diarrhea. *Gastroenterol Hepatol*. 2006;2(3 PG-203-208):203–8.
- Hur KY, Lee MS. Gut microbiota and metabolic disorders. *Diabetes Metab J*. 2015;39(3 PG-198-203):198–203.
- Hussack G, Tanha J. An update on antibody-based immunotherapies for *Clostridium difficile* infection. *Clin Exp Gastroenterol*. 2016;9(PG-209-224):209–24.
- i, Y. T.; Cai, H. F.; Wang, Z. H.; Xu, J.; Fang, J. Y.
- Ianiro G, Bibbò S, Gasbarrini A, Cammarota G. Therapeutic Modulation of Gut Microbiota: Current Clinical Applications and Future Perspectives. *Curr Drug Targets*. 2014 Jul 31;15(8):762–70.

- Ianiro G, Bibbò S, Scaldaferri F, Gasbarrini A, Cammarota G. Fecal microbiota transplantation in inflammatory bowel disease: beyond the excitement. *Medicine (Baltimore)*. 2014;93(19):e97.
- Ianiro G, Bibbò S, Scaldaferri F, Gasbarrini A, Cammarota G. Fecal microbiota transplantation in inflammatory bowel disease: beyond the excitement. *Medicine (Baltimore)*. 2014;93(19):e97.
- Ianiro G, Bibbò S, Scaldaferri F, Gasbarrini A, Cammarota G. Fecal microbiota transplantation in inflammatory bowel disease: beyond the excitement. *Medicine (Baltimore)*. 2014;93(19):e97.
- Ince MN, Blazar BR, Edmond MB, Tricot G, Wannemuehler MJ. Understanding Luminal Microorganisms and Their Potential Effectiveness in Treating Intestinal Inflammation. *Inflamm Bowel Dis*. 2016;22(1 PG-194-201):194–201.
- Infection December 2012;40(6):643-648  
Infection February 2014;42(1):43-59
- Iv EC, Iii EC, Johnson DA. Clinical update for the diagnosis and treatment of *Clostridium difficile* infection. *World J Gastrointest Pharmacol Ther*. 2014;5(1 PG-1-26):1–26.
- Jarrad AM, Karoli T, Blaskovich MA, Lyras D, Cooper MA. *Clostridium difficile* drug pipeline: challenges in discovery and development of new agents. *J Med Chem*. 2015;58(13 PG-5164-85):5164–85.
- Jarrad AM, Karoli T, Blaskovich MAT, Lyras D, Cooper MA. *Clostridium difficile* Drug Pipeline: Challenges in Discovery and Development of New Agents. *J Med Chem*. 2015;58(13 PG-5164-5185):5164–85.
- Jaworski A, Borody TJ, Leis S, Gadalla S, Dawson V. Treatment of first-time *clostridium difficile* infection with fecal microbiota transplantation. *Am J Gastroenterol*. 2015;110(PG-S587):S587.
- Jaworski A, Mitchell SW, Wong C, Chapman B, Bull M, Gadalla S, et al. FMT; how do alternate formats compare? *Am J Gastroenterol*. 2016;111(PG-S438):S438.
- Jaworski A, Mitchell SW, Wong C, Gadalla S, Borody TJ. Patient with relapsing *C. difficile* successfully treated with lyophilised encapsulated faecal microbiota transplant product. *J Gastroenterol Hepatol*. 2016;31(PG-161):161.
- Jehangir A, Bennett K, Fareedy SB, Rettew A, Shaikh B, Qureshi A, et al. Recurrent *C. difficile* in a Patient with IgG Deficiency. *Case Rep Gastrointest Med*. 2015;2015(PG-356293):356293.
- Jeon YD, Hong N, Kim JH, Park SH, Kim SB, Song IJ, et al. Fecal Transplantation using a Nasoenteric Tube during an Initial Episode of Severe *Clostridium difficile* Infection. *Infect Chemother*. 2016;48(1 PG-31-5):31–5.
- Jeon YD, Hong N, Kim JH, Park SH, Song IJ, Ann HW, et al. Fecal transplantation using a nasoenteric tube during an initial episode of severe *clostridium difficile* infection. *Infect Chemother*. 2016;48(1 PG-31-35):31–5.
- Jia N. A Misleading Reference for Fecal Microbiota Transplant. *Am J Gastroenterol*. 2015;110(12 PG-1731):1731.

- Jiang ZD, Ajami N, Lasco T, Petrosino J, Hochman F, Ankoma-Sey V, et al. Fresh, frozen, or lyophilized fecal microbiota transplantation (FMT) for multiple recurrent *C. Difficile* Infection (CDI). *Am J Gastroenterol*. 2014;109(PG-S213):S213.
- Jiang ZD, Dupont H, Ajami N, Lasco T, Ke S, Petrosino J, et al. Donor species richness determines fecal microbiota transplantation success in patients with recurrent *clostridium difficile* infection. *Gastroenterology*. 2016;1(PG-S895):S895.
- Jiang ZD, DuPont H, Ke S. A mouse model of *clostridium difficile* infection (CDI) suitable for study of human fecal microbiota transplantation (FMT). *Am J Gastroenterol*. 2015;110(PG-S577):S577.
- Jiang ZD, Hoang LN, Lasco TM, Garey KW, DuPont HL. Physician attitudes toward the use of fecal transplantation for recurrent *Clostridium difficile* infection in a metropolitan area. *Clin Infect Dis*. 2013;56(7 PG-1059-1060):1059–60.
- Johnson S. Recurrent *Clostridium difficile* infection: A review of risk factors, treatments, and outcomes. *J Infect*. 2009;58(6 PG-403-410):403–10.
- Jones JD, Murphy DW. Rescue fecal microbiota transplantation in refractory severe and complicated *clostridium difficile* infection using frozen stool specimens. *Gastroenterology*. 2015;1(PG-S641):S641.
- Jones L, Jones C. Does the donor matter? Donor vs. patient effects in the outcome of next-generation fecal transplant for recurrent *clostridium difficile* infection. *Gastroenterology*. 2015;1(PG-S328-S329):S328–9.
- Joseph J, Singhal S, Patel GM, Anand S. *Clostridium difficile* colitis: Review of the therapeutic approach. *Am J Ther*. 2014;21(5 PG-385-394):385–94.
- Joseph J, Singhal S, Patel GM, Anand S. *Clostridium difficile* colitis: Review of the therapeutic approach. *Am J Ther*. 2012;17(PG-).
- Journal of Digestive Diseases September 2016;17():43
- Journal of Gastroenterology and Hepatology (Australia) November 2016;31():155
- Journal of Pediatric Gastroenterology and Nutrition 13 Jan 2015;60(1):23-26
- Jung Lee W, Lattimer LD, Stephen S, Borum ML, Doman DB. Fecal Microbiota Transplantation: A Review of Emerging Indications Beyond Relapsing *Clostridium difficile* Toxin Colitis. *Gastroenterol Hepatol (N Y)*. 2015;11(1 PG-24-32):24–32.
- Kahn SA, Goepfinger SR, Rubin DT. Fecal microbiota transplantation: an interest in IBD? *Nestle Nutr Inst Workshop Ser*. 2014;79(PG-101-114):101–14.
- Kahn SA, Goepfinger SR, Rubin DT. Fecal microbiota transplantation: A new therapy for IBD? *Proceeding with caution. Inflamm Bowel Dis Monit*. 2013;13(4 PG-127-134):127–34.
- Kahn SA, Young S, Rubin DT. Colonoscopic fecal microbiota transplant for recurrent *clostridium difficile* infection in a child. *Am J Gastroenterol*. 2012;107(12 PG-1930-1931):1930–1.
- Kaiser AM, Hogen R, Bordeianou L, Alavi K, Wise PE, Sudan R, et al. *Clostridium Difficile* Infection from a Surgical Perspective. *J Gastrointest Surg*. 2015;19(7 PG-1363-77):1363–77.



- Kang DJ, Hylemon PB, Bajaj JS. Fecal transplant to mitigate hyperammonemia and hepatic encephalopathy in animal models. *Ann Hepatol*. 2015;14(5 PG-762-763):762–3.
- Kapel N, Thomas M, Corcos O, Mayeur C, Barbot-Trystram L, Bouhnik Y, et al. Practical implementation of faecal transplantation. *Clin Microbiol Infect*. 2014;20(11 PG-1098-1105):1098–105.
- Karadsheh Z, Sule S. Fecal transplantation for the treatment of recurrent *Clostridium difficile* infection. *N Am J Med Sci*. 2013;5(6 PG-339-343):339–43.
- Kassam Z, Blackler D, Osman M, Dubois N, Ling K, Burns L, et al. Novel safety features in fecal microbiota transplantation for recurrent *clostridium difficile* infection: Quality assurance and adverse events workflow. *Am J Gastroenterol*. 2015;110(PG-S589):S589.
- Kassam Z, Lee CH, Hunt RH. Review of the emerging treatment of *clostridium difficile* infection with fecal microbiota transplantation and insights into future challenges. *Clin Lab Med*. 2014;34(4 PG-787-798):787–98.
- Kassam, Z.; Lee, C. H.; Yuan, Y.; Hunt, R. H. Fecal microbiota transplantation for *clostridium difficile* infection: Systematic review and meta-analysis *American Journal of Gastroenterology* April 2013;108(4):500-508
- Kee VR. *Clostridium difficile* infection in older adults: A review and update on its management. *Am J Geriatr Pharmacother*. 2012;10(1 PG-14-24):14–24.
- Keller JJ, Kuijper EJ. Treatment of recurrent and severe *clostridium difficile* infection. 4139 El Camino Way, P.O. Box 10139, Palo Alto CA 94306, United States: Annual Reviews Inc.; 2015;(66 PG-373-386):373–86.
- Keller JJ, Kuijper EJ. Treatment of recurrent and severe *Clostridium difficile* infection. *Annu Rev Med*. 2015;66(PG-373-86):373–86.
- Keller PM, Weber MH. Rational therapy of *Clostridium difficile* infections. *Viz Gastrointest Med Surg*. 2014;30(5 PG-304-309):304–9.
- Kellermayer R. Prospects and challenges for intestinal microbiome therapy in pediatric gastrointestinal disorders. *World J Gastrointest Pathophysiol*. 2013;4(4 PG-91-3):91–3.
- Kelly CP. Fecal microbiota transplantation - An old therapy comes of age. *N Engl J Med*. 2013;368(5 PG-474-475):474–5.
- Kelly CP. Fecal microbiota transplantation--an old therapy comes of age. *N Engl J Med*. 2013;368(5 PG-474-5):474–5.
- Kelly CR, Kahn S, Kashyap P, Laine L, Rubin D, Atreja A, et al. Update on Fecal Microbiota Transplantation 2015: Indications, Methodologies, Mechanisms, and Outlook. *Gastroenterology*. Elsevier; 2015 Nov 19;149(1):223–37.
- Kelly P. Infectious diarrhoea. *Med (United Kingdom)*. 2015;43(5 PG-253-258):253–8.
- Kerman DH. Endoscopic Delivery of Fecal Biotherapy in Inflammatory Bowel Disease. *Gastrointest Endosc Clin N Am*. 2016;26(4 PG-707-717):707–17.

- Khan, M. A.; Sofi, A. A.; Ahmad, U.; Alaradi, O.; Khan, A. R.; Hammad, T.; Pratt, J.; Sodeman, T.; Sodeman, W.; Kamal, S.; Nawras, A. Efficacy and safety of, and patient satisfaction with, colonoscopic-administered fecal microbiota transplantation in relapsing and refractory community- and hospital-acquired *Clostridium difficile* infection Canadian Journal of Gastroenterology and Hepatology 01 Sep 2014;28(8):434-438
- Khanna S, Pardi DS. *Clostridium difficile* infection: Management strategies for a difficult disease. Therap Adv Gastroenterol. 2014;7(2 PG-72-86):72–86.
- Khanna S, Pardi DS. *Clostridium difficile* infection: New insights into management. Mayo Clin Proc. 2012;87(11 PG-1106-1117):1106–17.
- Khanna S, Tosh PK. A clinician's primer on the role of the microbiome in human health and disease. Mayo Clin Proc. 2014;89(1 PG-107-14):107–14.
- Khanna, S.; Pardi, D. S.; Kelly, C. R.; Kraft, C. S.; Dhere, T.; Henn, M. R.; Lombardo, M. J.; Vulic, M.; Ohsumi, T.; Winkler, J.; Pindar, C.; McGovern, B. H.; Pomerantz, R. J.; Aunins, J. G.; Cook, D. N.; Hohmann, E. L. A Novel Microbiome Therapeutic Increases Gut Microbial Diversity and Prevents Recurrent *Clostridium difficile* Infection Journal of Infectious Diseases 15 Jul 2016;214(2):173-181
- Khoruts A, Sadowsky MJ, Hamilton MJ. Development of Fecal Microbiota Transplantation Suitable for Mainstream Medicine. Clin Gastroenterol Hepatol. 2015;13(2 PG-246-250):246–50.
- Khoruts A, Sadowsky MJ. Therapeutic transplantation of the distal gut microbiota. Mucosal Immunol. 2011;4(1 PG-4-7):4–7.
- Khoruts A, Sadowsky MJ. Understanding the mechanisms of faecal microbiota transplantation. Nat Rev Gastroenterol Hepatol. 2016 Jun;13(9):508–16.
- Khoruts A, Weingarden AR. Emergence of fecal microbiota transplantation as an approach to repair disrupted microbial gut ecology. Immunol Lett. 2014;162(2 Pt A PG-77-81):77–81.
- Khoruts A. Faecal Microbiota transplantation in 2013: Developing human gut Microbiota as a class of therapeutics. Nat Rev Gastroenterol Hepatol. 2014;11(2 PG-79-80):79–80.
- Khoruts A. Fecal microbiota transplantation-early steps on a long journey ahead. Gut Microbes. 2017;8(3):199-204.
- Kim S, Lee Y, Kim SH. Safety and effectiveness of fecal microbiota transplantation: A systematic review. [Korean]. Journal of the Korean Medical Association. 2017;60(9):761-8.
- Kim YG, Jang BI. Current advances related to *Clostridium difficile* infection. Indian J Med Res. 2015;141(2 PG-172-174):172–4.
- Martins FS. Fecal Microbiota Transplantation for Ulcerative Colitis: FoMenTing Change? Dig Dis Sci. 2016;61(8 PG-2154-2155):2154–5.
- Knight CL, Surawicz CM. *Clostridium difficile* Infection. Med Clin North Am. 2013;97(4 PG-523-536):523–36.



- Kociolek LK, Gerding DN. Breakthroughs in the treatment and prevention of *Clostridium difficile* infection. *Nat Rev Gastroenterol Hepatol*. 2016;13(3 PG-150-160):150–60.
- Koenigsnecht MJ, Young VB. Faecal microbiota transplantation for the treatment of recurrent *Clostridium difficile* infection: Current promise and future needs. *Curr Opin Gastroenterol*. 2013;29(6 PG-628-632):628–32.
- Konig J, Brummer RJ. Alteration of the intestinal microbiota as a cause of and a potential therapeutic option in irritable bowel syndrome. *Benef Microbes*. 2014;5(3 PG-247-261):247–61.
- Konig J, Siebenhaar A, Hogenauer C, Arkkila P, Nieuwdorp M, Noren T, et al. Consensus report: faecal microbiota transfer - clinical applications and procedures. *Aliment Pharmacol Ther*. 2017;45(2):222-39.
- Konturek PC, Haziri D, Brzozowski T, Hess T, Heyman S, Kwiecien S, et al. Emerging role of fecal microbiota therapy in the treatment of gastrointestinal and extra-gastrointestinal diseases. *J Physiol Pharmacol*. 2015;66(4 PG-483-491):483–91.
- Konturek PC, Hess T. Stool transplantation - Gut bacteria as a novel therapeutic option. *MMW-Fortschritte der Medizin*. 2015;157(3 PG-61-63):61–3.
- Korman TM. Diagnosis and Management of *Clostridium difficile* Infection. *Semin Respir Crit Care Med*. 2015;36(1 PG-31-43):31–43.
- Kump PK, Krause R, Allerberger F, Hogenauer C. Faecal microbiota transplantation--the Austrian approach. *Clin Microbiol Infect*. 2014;20(11 PG-1106-11):1106–11.
- Kump PK, Krause R, Allerberger F, Hogenauer C. Faecal microbiota transplantation-the Austrian approach. *Clin Microbiol Infect*. 2014;20(11 PG-1106-1111):1106–11.
- Lagier JC. Gut microbiota and *Clostridium difficile* infections. *Hum Microbiome J*. 2016;2(PG-10-14):10–4.
- Landy J, Al-Hassi HO, McLaughlin SD, Walker AW, Ciclitira PJ, Nicholls RJ, et al. Review article: Faecal transplantation therapy for gastrointestinal disease. *Aliment Pharmacol Ther*. 2011;34(4 PG-409-415):409–15.
- Langdon A, Crook N, Dantas G. The effects of antibiotics on the microbiome throughout development and alternative approaches for therapeutic modulation. *Genome Med. BioMed Central*; 2016 Apr 13;8(1):39.
- Lankelma JM, Nieuwdorp M, de Vos WM, Wiersinga WJ. The gut microbiota in internal medicine: Implications for health and disease. *Neth J Med*. 2015;73(2 PG-61-68):61–8.
- Le Monnier A, Zahar JR, Barbut F. Update on *clostridium difficile* infections. *Med Mal Infect*. 2014;44(8 PG-354-365):354–65.
- Leffler DA, Lamont JT. *Clostridium difficile* infection. *N Engl J Med*. 2015;372(16 PG-1539-1548):1539–48.

- Lemon KP, Armitage GC, Relman DA, Fischbach MA. Microbiota-targeted therapies: An ecological perspective. *Sci Transl Med*. 2012;4 (137) (no pagination)(137rv5 PG-).
- Leszczyszyn JJ, Radomski M, Leszczyszyn AM. Intestinal microbiota transplant - Current state of knowledge. *Reumatologia*. 2016;54(1 PG-24-28):24–8.
- Lewis SS, Anderson DJ. Treatment of *Clostridium difficile* infection: Recent trial results. *Clin Investig (Lond)*. 2013;3(9 PG-875-886):875–86.
- Liubakka A, Vaughn BP. *Clostridium difficile* Infection and Fecal Microbiota Transplant. *AACN Adv Crit Care*. 2016;27(3 PG-324-337):324–37.
- Lo Vecchio A, Cohen MB. Fecal microbiota transplantation for *Clostridium difficile* infection: Benefits and barriers. *Curr Opin Gastroenterol*. 2014;30(1 PG-47-53):47–53.
- Lo Vecchio A, Zacur GM. *Clostridium difficile* infection: An update on epidemiology, risk factors, and therapeutic options. *Curr Opin Gastroenterol*. 2012;28(1 PG-1-9):1–9.
- Lopez J, Grinspa A. Fecal microbiota transplantation for inflammatory bowel disease. *Gastroenterol Hepatol*. 2016;12(6 PG-374-379):374–9.
- Lubbert C, John E, von Muller L. *Clostridium difficile* infection: guideline-based diagnosis and treatment. *Dtsch Arztebl Int*. 2014;111(43 PG-723-731):723–31.
- Lubbert C. Antimicrobial therapy of acute diarrhoea: A clinical review. *Expert Rev Anti Infect Ther*. 2016;14(2 PG-193-206):193–206.
- Malikowski T, Khanna S, Pardi DS. Fecal microbiota transplantation for gastrointestinal disorders. *Curr Opin Gastroenterol*. 2017;33(1 PG-8-13):8–13.
- Malnick S, Melzer E. Human microbiome: From the bathroom to the bedside. *World J Gastrointest Pathophysiol*. 2015;6(3 PG-79-85):79–85.
- Manges AR, Steiner TS, Wright AJ. Fecal microbiota transplantation for the intestinal decolonization of extensively antimicrobial-resistant opportunistic pathogens: a review. *Infect Dis (Auckl)*. 2016;48(8 PG-587-92):587–92.
- Marra F, Ng K. Controversies Around Epidemiology, Diagnosis and Treatment of *Clostridium difficile* Infection. *Drugs*. 2015;75(10 PG-1095-1118):1095–118.
- Martin J, Mawer D, Wilcox MH. *Clostridium difficile*: Biological therapies. *Curr Opin Infect Dis*. 2013;26(5 PG-454-460):454–60.
- Martin J, Wilcox M. New and emerging therapies for *Clostridium difficile* infection. *Curr Opin Infect Dis*. 2016 Dec;29(6):546–54.
- Mathur H, Rea MC, Cotter PD, Paul Ross R, Hill C. The potential for emerging therapeutic options for *Clostridium difficile* infection. *Gut Microbes*. 2015;5(6 PG-696-710):696–710.

- Mathur H, Rea MC, Cotter PD, Ross RP, Hill C. The potential for emerging therapeutic options for *Clostridium difficile* infection. *Gut Microbes*. 2014;5(6 PG-696-710):696–710.
- Matsuoka K, Mizuno S, Hayashi A, Hisamatsu T, Naganuma M, Kanai T. Fecal microbiota transplantation for gastrointestinal diseases. *Keio J Med*. 2014;63(4 PG-69-74):69–74.
- Mattner J, Schmidt F, Siegmund B. Faecal microbiota transplantation-A clinical view. *Int J Med Microbiol*. 2016;306(5 PG-310-315):310–5.
- McCune VL, Struthers JK, Hawkey PM. Faecal transplantation for the treatment of *Clostridium difficile* infection: A review. *Int J Antimicrob Agents*. 2014;43(3 PG-201-206):201–6.
- Merenstein D, El-Nachef N, Lynch S V. Fecal microbial therapy: Promises and pitfalls. *J Pediatr Gastroenterol Nutr*. 2014;59(2 PG-157-161):157–61.
- Mergenhagen KA, Wojciechowski AL, Paladino JA. A review of the economics of treating *Clostridium difficile* infection. *Pharmacoeconomics*. 2014;32(7 PG-639-650):639–50.
- Mizusawa M, Doron S, Gorbach S. *Clostridium difficile* Diarrhea in the Elderly: Current Issues and Management Options. *Drugs and Aging*. 2015;32(8 PG-639-647):639–47.
- Moayyedi P, Marshall J, Yuan Y, Hunt R. Canadian Association of Gastroenterology position statement: Fecal microbiota transplant therapy. *Can J Gastroenterol Hepatol*. 2014;28(2 PG-66-68):66–8.
- Moayyedi P. Fecal transplantation: Any real hope for inflammatory bowel disease? *Curr Opin Gastroenterol*. 2016;32(4 PG-282-286):282–6.
- Monsour Jr. HP, Quigley EM. The Microbiome: What Will the Future Hold? *Semin Liver Dis*. 2016;36(4 PG-354-359):354–9.
- Monsour HP, Quigley EMM. The Microbiome: What Will the Future Hold? *Semin Liver Dis*. 2016;36(4 PG-354-359):354–9.
- Moore T, Rodriguez A, Bakken JS. Fecal microbiota transplantation: A practical update for the infectious disease specialist. *Clin Infect Dis*. 2014;58(4 PG-541-545):541–5.
- Mosby D, Lopresto BI, Bacon A, Levy S. Systematic review: Fecal microbiota transplantation in the treatment of pediatric gastrointestinal diseases. *Inflamm Bowel Dis*. 2016;22(PG-S73):S73.
- Moayyedi P, Yuan Y, Baharath H, Ford AC. Faecal microbiota transplantation for *Clostridium difficile*-associated diarrhoea: a systematic review of randomised controlled trials. *Med J Aust*. 2017;207(4):166–72.
- Mulcahy-O’Grady H, Workentine ML. The challenge and potential of metagenomics in the clinic. *Front Immunol*. 2016;7 (FEB) (no pagination)(29 PG-).
- Mullish BH, Marchesi JR, Thursz MR, Williams HR. Microbiome manipulation with faecal microbiome transplantation as a therapeutic strategy in *Clostridium difficile* infection. *Qjm*. 2015;108(5 PG-355-9):355–9.

- Mullish BH, Marchesi JR, Thursz MR, Williams HRT. Microbiome manipulation with faecal microbiome transplantation as a therapeutic strategy in *Clostridium difficile* infection. *Qjm*. 2015;108(5 PG-355-359):355–9.
- Mullish BH, McDonald JAK, Pechlivanis A, Rees DN, Williams HRT, Marchesi JR, et al. Understanding the efficacy of faecal microbiota transplantation in *clostridium difficile* infection: Re-Establishment of gut microbiota with the ability to degrade bile? *Gut*. 2016;65(PG-A184):A184.
- Mullish BH, Williams HR. Obstacles to establishing an NHS faecal transplant programme. *BMJ*. 2015;351(PG-h6043):h6043.
- Mullish BH, Williams HRT. Obstacles to establishing an NHS faecal transplant programme. *BMJ*. 2015;351 (no pagination)(3496 PG-).
- Myers F. Beyond mainstream: making the case for fecal bacteriotherapy. *Nursing (Lond)*. 2011;41(12 PG-50-53):50–3.
- Navarro F, Liu Y, Rhoads JM. Can probiotics benefit children with autism spectrum disorders? *World J Gastroenterol*. 2016;22(46 PG-10093-10102):10093–102.
- Nicholson MR, Osgood CL, Acra SA, Edwards KM. *Clostridium difficile* infection in the pediatric transplant patient. *Pediatr Transplantation*. 2015;(PG-).
- Nitzan O, Elias M, Chazan B, Raz R, Saliba W. *Clostridium difficile* and inflammatory bowel disease: role in pathogenesis and implications in treatment. *World J Gastroenterol*. China; 2013;19(43):7577–85.
- Obi O, Hampton D, Anderson T, Leung P, Abdul MKM, Chandra G, et al. Fecal microbiota transplant for treatment of resistant *C. Difficile* infection using a standardized protocol: A community hospital experience. *Am J Gastroenterol*. 2014;109(PG-S629):S629.
- Ofosu A. *Clostridium difficile* infection: A review of current and emerging therapies. *Ann Gastroenterol*. 2016;29(2 PG-147-154):147–54.
- Orenstein R, Griesbach CL, Dibaise JK. Moving fecal microbiota transplantation into the mainstream. *Nutr Clin Pract*. 2013;28(5 PG-589-598):589–98.
- P. K, Kump P, Hogenauer C. Any Future for Fecal Microbiota Transplantation as Treatment Strategy for Inflammatory Bowel Diseases?. *Dig Dis*. Switzerland: S. Karger AG; 2016;34(1 Supplement 1):74–81.
- Paasche S. Fecal microbiota transplantation: An innovative approach to treating *Clostridium difficile* disease. *J Am Acad Physician Assist*. 2013;26(8 PG-46-49):46–9.
- Pacheco SM, Johnson S. Important clinical advances in the understanding of *Clostridium difficile* infection. *Curr Opin Gastroenterol*. 2013;29(1 PG-42-48):42–8.
- Padua D, Pothoulakis C. Novel approaches to treating *Clostridium difficile*-associated colitis. *Expert Rev Gastroenterol Hepatol*. 2016;10(2 PG-193-204):193–204.

- Pant C, Sferra TJ, Deshpande A, Minocha A. Clinical approach to severe *Clostridium difficile* infection: Update for the hospital practitioner. *Eur J Intern Med*. 2011;22(6 PG-561-568):561–8.
- Pathak R, Enuh HA, Patel A, Wickremesinghe P. Treatment of relapsing *clostridium difficile* infection using fecal microbiota transplantation. *Clin Exp Gastroenterol*. 2014;7(1 PG-).
- Petrof EO, Khoruts A. From stool transplants to next-generation microbiota therapeutics. *Gastroenterology*. 2014;146(6 PG-1573-1582):1573–82.
- Quraishi MN, Widlak M, Bhala N, Moore D, Price M, Sharma N, et al. Systematic review with meta-analysis: the efficacy of faecal microbiota transplantation for the treatment of recurrent and refractory *Clostridium difficile* infection. *Aliment Pharmacol Ther*. 2017;46(5):479-93.
- Rabe SM. Treatment of recurrent *Clostridium difficile* infection with fecal transplantation. *Gastroenterol Nurs*. 2014;37(2 PG-156-63; quiz 164-5):156–63; quiz 164.
- Raghunath V, Levy M, Koo K, Foo H, Borody TJ, Wong J. Recurrent *clostridium difficile* infection in a renal transplant recipient successfully treated with fecal microbiota transplantation. *Nephrology*. 2014;19(PG-98):98.
- Rao K, Higgins PD. Epidemiology, Diagnosis, and Management of *Clostridium difficile* Infection in Patients with Inflammatory Bowel Disease. *Inflamm Bowel Dis*. 2016;22(7 PG-1744-54):1744–54.
- Rao K, Higgins PDR. Epidemiology, Diagnosis, and Management of *Clostridium difficile* Infection in Patients with Inflammatory Bowel Disease. *Inflamm Bowel Dis*. 2016;22(7 PG-1744-1754):1744–54.
- Rao K, Safdar N. Fecal microbiota transplantation for the treatment of *Clostridium difficile* infection. *J Hosp Med*. 2016;11(1 PG-56-61):56–61.
- Rao K, Young VB, Aronoff DM. Fecal microbiota therapy: Ready for prime time? *Infect Control Hosp Epidemiol*. 2014;35(1 PG-28-30):28–30.
- Rao K, Young VB. Fecal Microbiota Transplantation for the Management of *Clostridium difficile* Infection. *Infect Dis Clin North Am*. 2015;29(1 PG-109-122):109–22.
- Rao K, Young VB, Malani PN. Capsules for Fecal Microbiota Transplantation in Recurrent *Clostridium difficile* Infection: The New Way Forward or a Tough Pill to Swallow? *Jama*. 2017;318(20):1979-80.
- Raoult D. Faecal transplantation and infectious diseases practitioners. *Clin Microbiol Infect*. 2014;20(11 PG-1097):1097.
- Raoult D. The return of microbes. *Clin Microbiol Infect*. 2016;22(10 PG-822-823):822–3.
- Ray K. Infection : Microbiota reconstitution for resistance to *Clostridium difficile* infection-fight fire with fire? *Nat Rev Gastroenterol Hepatol*. 2015;12(1 PG-4):4.

- Ray K. Infection: microbiota reconstitution for resistance to *Clostridium difficile* infection--fight fire with fire? *Nat Rev Gastroenterol Hepatol*. 2015;12(1 PG-4):4.
- Rineh A, Kelso MJ, Vatansever F, Tegos GP, Hamblin MR. *Clostridium difficile* infection: Molecular pathogenesis and novel therapeutics. *Expert Rev Anti Infect Ther*. 2014;12(1 PG-131-150):131–50.
- Rodriguez C, Taminiau B, Van Broeck J, Delmee M, Daube G. *Clostridium difficile* infection and intestinal microbiota interactions. *Microb Pathog*. 2015;89(PG-201-209):201–9.
- Rogers GB, Bruce KD. Challenges and opportunities for faecal microbiota transplantation therapy. *Epidemiol Infect*. 2013;141(11 PG-2235-2242):2235–42.
- Rohlke F, Stollman N. Fecal microbiota transplantation in relapsing *Clostridium difficile* infection. *Therap Adv Gastroenterol*. 2012;5(6):403–20.
- Rubin DT, Kirsner JB. Fecal microbiota transplantation for the treatment of inflammatory bowel disease. *Gastroenterol Hepatol*. 2015;11(9 PG-618-620):618–20.
- Rubin DT. Curbing our enthusiasm for fecal transplantation in ulcerative colitis. *Am J Gastroenterol*. 2013;108(10 PG-1631-1633):1631–3.
- Rubin TA, Gessert CE, Aas J. Stool transplantation for older patients with *clostridium difficile* infection. *J Am Geriatr Soc*. 2009;57(12 PG-2386-2387):2386–7.
- Russell RK, Protheroe A, Roughton M, Croft N, Murphy MS, Spray C, et al. Inpatient paediatric UC care in the UK in 2011 is characterised by increasing rates of rescue therapy and stool cultures but low use of pucai scores. *Gut*. 2012;61(PG-A22):A22.
- Sageer M, Barto A. Recurrent *Clostridium difficile* infection: The scope of the problem and management decisions. *Semin Colon Rectal Surg*. 2014;25(3 PG-158-162):158–62.
- Salman S, Vardatsikos G, Avar D, Palmour N, Dewar K, Zawati MH. FMT Happens: Regulating Fecal Microbiota Therapy in Canada; What You Need to Know. *World Med Heal Policy*. 2016;8(1 PG-95-106):95–106.
- Sampath K, Levy LC, Gardner TB. Fecal transplantation: Beyond the aesthetic. *Gastroenterology*. 2013;145(5 PG-1151-1153):1151–3.
- Satokari R, Mattila E, Kainulainen V, Arkkila PE. Editorial: a simple faecal preparation for faecal microbiota transplantation--authors' reply. *Aliment Pharmacol Ther*. 2015;41(3 PG-321):321.
- Satokari R, Mattila E, Kainulainen V, Arkkila PE. Editorial: A simple faecal preparation for faecal microbiota transplantation - Authors' reply. *Aliment Pharmacol Ther*. 2015;41(3 PG-321):321.
- Scaldaferri F, Pecere S, Petito V, Zambrano D, Fiore L, Lopetuso LR, et al. Efficacy and Mechanisms of Action of Fecal Microbiota Transplantation in Ulcerative Colitis: Pitfalls and Promises from a First Meta-Analysis. *Transplant Proc*. 2016;48(2 PG-402-407):402–7.



- Schenck LP, Beck PL, MacDonald JA. Gastrointestinal dysbiosis and the use of fecal microbial transplantation in *Clostridium difficile* infection. *World J Gastrointest Pathophysiol*. 2015;6(4 PG-169-80):169–80.
- Scott KP, Antoine JM, Midtvedt T, van Hemert S. Manipulating the gut microbiota to maintain health and treat disease. *Microb Ecol Heal Dis*. 2015;26(PG-25877):25877.
- Shin JH, Chaves-Olarte E, Warren CA. *Clostridium difficile* Infection. *Microbiol Spectr*. 2016;4(3 PG-).
- Siebenhaar A, Rosien U. Fecal microbiome transfer. *Internist Prax*. 2016;56(2 PG-269-277):269–77.
- Singh B, Qin N, Reid G. Microbiome regulation of autoimmune, gut and liver associated diseases. *Inflamm Allergy - Drug Targets*. 2015;14(2 PG-84-93):84–93.
- Singh N, Suskind D, Wahbeh G. Fecal bacteriotherapy in a 6 year old patient with ulcerative colitis and *clostridium difficile*. *Inflamm Bowel Dis*. 2012;18(PG-S69):S69.
- Singh R, Nieuwdorp M, ten Berge IJ, Bemelman FJ, Geerlings SE. The potential beneficial role of faecal microbiota transplantation in diseases other than *Clostridium difficile* infection. *Clin Microbiol Infect*. 2014;20(11 PG-1119-25):1119–25.
- Singh R, Nieuwdorp M, ten Berge IJM, Bemelman FJ, Geerlings SE. The potential beneficial role of faecal microbiota transplantation in diseases other than *Clostridium difficile* infection. *Clin Microbiol Infect*. 2014;20(11 PG-1119-1125):1119–25.
- Sirisinha S. The potential impact of gut microbiota on your health: Current status and future challenges. *Asian Pacific J Allergy Immunol*. 2016;34(4 PG-249-264):249–64.
- Smits LP, Bouter KE, de Vos WM, Borody TJ, Nieuwdorp M. Therapeutic potential of fecal microbiota transplantation. *Gastroenterology*. 2013;145(5):946–53.
- Smits WK, Lyras D, Lacy DB, Wilcox MH, Kuijper EJ. *Clostridium difficile* infection. *Nat Rev Dis Prim*. 2016;2(PG-1-20):1–20.
- Steinert A, Radulovic K, Niess J. Gastro-intestinal tract: The leading role of mucosal immunity. *Swiss Med Wkly*. 2016;146(PG-w14293):w14293.
- Stuntz M, Des Vignes F. Treating *Clostridium difficile* infections: Should fecal microbiota transplantation be reclassified from investigational drug to human tissue? *Contemp Clin Trials Commun*. 2015;1(PG-39-41):39–41.
- Surawicz CM. Fecal microbiota transplantation: What we know and what we need to know. *Ann Intern Med*. 2015;162(9 PG-662-663):662–3.
- Suwantarat N, Bobak DA. Fecal bacteriotherapy for recurrent *clostridium difficile* infection: What's old is new again? *Curr Infect Dis Rep*. 2013;15(2 PG-101-103):101–3.
- Tamma PD, Sandora TJ. *Clostridium difficile* infection in children: Current state and unanswered questions. *J Pediatric Infect Dis Soc*. 2012;1(3 PG-230-243):230–43.

- 1  
2  
3 Tang G, Yin W, Liu W. Is frozen fecal microbiota transplantation as effective as fresh fecal microbiota  
4 transplantation in patients with recurrent or refractory *Clostridium difficile* infection: A meta-analysis?  
5 *Diagnostic Microbiology and Infectious Disease*. 2017;88(4):322-9.  
6  
7 Tariq R, Khanna S. *Clostridium difficile* infection: Updates in management. *Indian J Gastroenterol*. 2016;(PG-  
8 1-8):1-8.  
9  
10 Taur Y, Pamer EG. Harnessing microbiota to kill a pathogen: Fixing the microbiota to treat *Clostridium difficile*  
11 infections. *Nat Med*. 2014;20(3 PG-246-247):246-7.  
12  
13 Taur Y, Pamer EG. The intestinal microbiota and susceptibility to infection in immunocompromised patients.  
14 *Curr Opin Infect Dis*. 2013;26(4 PG-332-337):332-7.  
15  
16 Thomas L V, Suzuki K, Zhao J. Probiotics: a proactive approach to health. A symposium report. *Br J Nutr*.  
17 2015;114 Suppl 1(PG-S1-15):S1-15.  
18  
19 Too early to determine whether fecal microbiota transplant has therapeutic promise for ulcerative colitis?  
20 *Journal of Pediatric Gastroenterology and Nutrition*. G.H. Russell, Division of Gastroenterology and Nutrition,  
21 Boston Children's Hospital, 300 Longwood Ave, Boston, MA 02115, United States: Lippincott Williams and  
22 Wilkins; 2015. p. 3.  
23  
24 Tran MC, Claros MC, Goldstein EJ. Therapy of *Clostridium difficile* infection: perspectives on a changing  
25 paradigm. *Expert Opin Pharmacother*. 2013;14(17 PG-2375-86):2375-86.  
26  
27 Tran MCN, Claros MC, Goldstein EJC. Therapy of *Clostridium difficile* infection: Perspectives on a changing  
28 paradigm. *Expert Opin Pharmacother*. 2013;14(17 PG-2375-2386):2375-86.  
29  
30 Treating *Clostridium difficile* Infection With Fecal Microbiota Transplantation. *Clin Gastroenterol Hepatol*.  
31 W.B. Saunders; 2011 Dec;9(12):1044-9.  
32  
33 Trifan A, Stoica O, Stanciu C, Cojocariu C, Singeap AM, Girleanu I, et al. *Clostridium difficile* infection in  
34 patients with liver disease: a review. *Eur J Clin Microbiol Infect Dis*. 2015;34(12 PG-2313-2324):2313-24.  
35  
36 Trubiano JA, Cheng AC, Korman TM, Roder C, Campbell A, May ML, et al. Australasian Society of Infectious  
37 Diseases updated guidelines for the management of *Clostridium difficile* infection in adults and children in  
38 Australia and New Zealand. *Intern Med J*. 2016;46(4 PG-479-93):479-93.  
39  
40 Turner JD. Evaluation of *clostridium difficile* treatment with fecal microbiota transplantation. *Am J Trop Med*  
41 *Hyg*. 2013;1(PG-217):217.  
42  
43 Unal CM, Steinert M. Novel therapeutic strategies for *Clostridium difficile* infections. *Expert Opin Ther*  
44 *Targets*. 2016;20(3 PG-269-285):269-85.  
45  
46 Vaishnavi C. Fecal microbiota transplantation for management of *Clostridium difficile* infection. *Indian J*  
47 *Gastroenterol*. 2014;33(4 PG-301-307):301-7.  
48  
49 Van Dissel JT. Alternative strategies for *Clostridium difficile* infection. *Antonie van Leeuwenhoek, Int J Gen*  
50 *Mol Microbiol*. 2009;95(PG-41):41.  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60



- van Nispen tot Pannerden CM, Verbon A, Kuipers EJ. Recurrent *Clostridium difficile* infection: what are the treatment options? *Drugs*. 2011;71(7 PG-853-68):853–68.
- Van Nispen Tot Pannerden CMF, Verbon A, Kuipers EJ. Recurrent *clostridium difficile* infection: What are the treatment options? *Drugs*. 2011;71(7 PG-853-868):853–68.
- van Nood E, Speelman P, Kuijper EJ, Keller JJ. Struggling with recurrent *Clostridium difficile* infections: is donor faeces the solution? *Euro Surveill*. 2009;14(34 PG-):.
- Van Nood E, Speelman P, Nieuwdorp M, Keller J. Fecal microbiota transplantation: Facts and controversies. *Curr Opin Gastroenterol*. 2014;30(1 PG-34-39):34–9.
- Vandenplas Y, Pierard D, De Greef E. Fecal Microbiota Transplantation: Just a Fancy Trend? *J Pediatr Gastroenterol Nutr*. 2015;61(1 PG-4-7):4–7.
- Vemuri RC, Gundamaraju R, Shinde T, Eri R. Therapeutic interventions for gut dysbiosis and related disorders in the elderly: antibiotics, probiotics or faecal microbiota transplantation? *Benef Microbes*. 2016;(PG-1-15):1–15.
- Venuto C, Butler M, Ashley ED, Brown J. Alternative therapies for *Clostridium difficile* infections. *Pharmacother J Hum Pharmacol Drug Ther*. 2010;30(12 PG-1266-78):1266–78.
- Venuto C, Butler M, Dodds Ashley E, Brown J. Alternative therapies for *Clostridium difficile* infections. *Pharmacotherapy*. 2010;30(12 PG-1266-1278):1266–78.
- Verbeke KA, Boesmans L, Boets E. Modulating the microbiota in inflammatory bowel diseases: prebiotics, probiotics or faecal transplantation?. *Proc Nutr Soc. England*; 2014;73(4):490–7.
- Verspohl E. Therapy of *Clostridium difficile*-associated diarrhea. Fecal microbiota transplants: History and clinical trials. *Med Monatsschr Pharm*. 2016;39(12 PG-539-542):539–42.
- Vestal R. Fecal Microbiota Transplant. *Hosp Med Clin*. 2016;5(1 PG-58-70):58–70.
- Vincent C, Manges AR. Antimicrobial use, human gut microbiota and *Clostridium difficile* colonization and infection. *Antibiotics*. 2015;4(3 PG-230-253):230–53.
- Vincent Y, Manji A, Gregory-Miller K, Lee C. A review of management of *Clostridium difficile* infection: Primary and recurrence. *Antibiotics*. 2015;4(4 PG-411-423):411–23.
- Vindigni SM, Broussard EK, Surawicz CM. Alteration of the intestinal microbiome: Fecal microbiota transplant and probiotics for *Clostridium difficile* and beyond. *Expert Rev Gastroenterol Hepatol*. 2013;7(7 PG-615-628):615–28.
- Vindigni SM, Surawicz CM. The gut microbiome: A clinically significant player in transplantation? *Expert Rev Clin Immunol*. 2015;11(7 PG-781-783):781–3.
- von Ameln-Mayerhofer A. [*Clostridium difficile* infection - an update]. *Med Monatsschr Pharm*. 2015;38(6 PG-211-8; quiz 219-20):211–8; quiz 219.

- Vyas D, L'Esperance H E, Vyas A. Stool therapy may become a preferred treatment of recurrent *Clostridium difficile*? *World J Gastroenterol*. 2013;19(29 PG-4635-7):4635–7.
- Vyas D, L'Esperance HE, Vyas A. Stool therapy may become a preferred treatment of recurrent *Clostridium difficile*? *World J Gastroenterol*. 2013;19(29 PG-4635-4637):4635–7.
- Walia R, Kunde S, Mahajan L. Fecal microbiota transplantation in the treatment of refractory *Clostridium difficile* infection in children: An update. *Curr Opin Pediatr*. 2014;26(5 PG-573-578):573–8.
- Walker AW, Lawley TD. Therapeutic modulation of intestinal dysbiosis. *Pharmacol Res*. 2013;69(1 PG-75-86):75–86.
- Walsh CJ, Guinane CM, O'Toole PW, Cotter PD. Beneficial modulation of the gut microbiota. *FEBS Lett*. 2014;588(22 PG-4120-4130):4120–30.
- Waltz P, Zuckerbraun B. Novel therapies for severe *Clostridium difficile* colitis. *Curr Opin Crit Care*. 2016;22(2 PG-167-73):167–73.
- Wang AY, Popov J, Pai N. Fecal microbial transplant for the treatment of pediatric inflammatory bowel disease. *World J Gastroenterol*. 2016;22(47 PG-10304-10315):10304–15.
- Wang SN, Cao HL, Wang BM. Systematic review: Adverse events of fecal microbiota transplantation. *J Dig Dis*. 2016;17(PG-59):59.
- Wang ZK, Yang YS, Chen Y, Yuan J, Sun G, Peng LH. Intestinal microbiota pathogenesis and fecal microbiota transplantation for inflammatory bowel disease. *World J Gastroenterol*. 2014;20(40 PG-14805-14820):14805–20.
- Weissman JS, Coyle W. Stool transplants: Ready for prime time? *Curr Gastroenterol Rep*. 2012;14(4 PG-313-316):313–6.
- West CE, Renz H, Jenmalm MC, Kozyrskyj AL, Allen KJ, Vuillermin P, et al. The gut microbiota and inflammatory noncommunicable diseases: associations and potentials for gut microbiota therapies. *J Allergy Clin Immunol*. 2015;135(1 PG-3-13; quiz 14):3–13; quiz 14.
- West CE, Renz H, Jenmalm MC, Kozyrskyj AL, Allen KJ, Vuillermin P, et al. The gut microbiota and inflammatory noncommunicable diseases: Associations and potentials for gut microbiota therapies. *J Allergy Clin Immunol*. 2015;135(1 PG-3-13):3–13.
- Wiedel N, Gilbert J, Baloun B, Nelson C. *Clostridium difficile* Associated Diarrhea. *S D Med*. 2016;69(3 PG-124-127):124–7.
- Wischmeyer PE, McDonald D, Knight R. Role of the microbiome, probiotics, and “dysbiosis therapy” in critical illness. *Curr Opin Crit Care*. 2016;22(4 PG-347-53):347–53.
- Wise J. Frozen faecal matter works as well as fresh for transplantation in *C difficile* patients. *BMJ*. 2016;352(PG-i138):i138.
- Woodworth MH, Carpentieri C, Sitchenko KL, Kraft CS. Challenges in fecal donor selection and screening for

fecal microbiota transplantation: A review. *Gut Microbes*. 2017;8(3):225-37.

Woodworth MH, Neish EM, Miller NS, Dhere T, Burd EM, Carpentieri C, et al. Laboratory Testing of Donors and Stool Samples for Fecal Microbiota Transplantation for Recurrent *Clostridium difficile* Infection. *J Clin Microbiol*. 2017;55(4):1002-10.

Woodworth MH, Neish EM, Miller NS, Dhere T, Burd EM, Carpentieri C, et al. Laboratory Testing of Donors and Stool for Fecal Microbiota Transplantation for Recurrent *C. difficile* Infection. *J Clin Microbiol*. 2017;11(PG-11):11.

Xu MQ, Cao HL, Wang WQ, Wang S, Cao XC, Yan F, et al. Fecal microbiota transplantation broadening its application beyond intestinal disorders. *World J Gastroenterol*. 2015;21(1 PG-102-111):102-11.

Yang X, Wu M. Fecal microbiota transplantation and inflammatory bowel disease. [Chinese]. *Chinese J Gastroenterol*. 2016;21(8 PG-491-493):491-3.

Ye L. Fecal microbiota transplantation: A potential therapy for inflammatory bowel disease? *J Dig Dis*. 2015;16(PG-81):81.

Young VB. Therapeutic manipulation of the microbiota: past, present, and considerations for the future. *Clin Microbiol Infect*. 2016;22(11 PG-905-909):905-9.

Youngster I, Gerding DN. Editorial: Making Fecal Microbiota Transplantation Easier to Swallow: Freeze-Dried Preparation for Recurrent *Clostridium difficile* Infections. *Am J Gastroenterol*. 2017;112(6):948-50.

Youngster I. Fecal microbiota transplants - The clinical perspective. *Int J Infect Dis*. 2016;45(PG-37):37.

Yuille S, Mackay WG, Morrison DJ, Tedford MC. Optimising gut colonisation resistance against *Clostridium difficile* infection. *Eur J Clin Microbiol Infect Dis*. 2015;34(11 PG-2161-2166):2161-6.

Zagaria MAE. Fecal transplantation for recurrent *clostridium difficile* infection. *US*. 2014;Pharmacist. 39(12 PG-20-22):20-2.

Zainah H, Hassan M, Shiekh-Sroujeh L, Hassan S, Alangaden G, Ramesh M. Intestinal Microbiota Transplantation, a Simple and Effective Treatment for Severe and Refractory *Clostridium Difficile* Infection. *Dig Dis Sci*. 2015 Jan 23;60(1):181-5.

Zalig S, Rupnik M. *Clostridium difficile* infection and gut microbiota. *Semin Colon Rectal Surg*. 2014;25(3 PG-124-127):124-7.

Zanella Terrier MC, Frossard JL, Simonet ML. [Recurrent *Clostridium difficile* infections: the importance of the intestinal microbiota]. *Rev Med Suisse*. 2013;9(402 PG-1898, 1900-4):1898,1900-1904.

Zanella Terrier MC, Simonet ML, Bichard P, Frossard JL. Recurrent *Clostridium difficile* infections: The importance of the intestinal microbiota. *World J Gastroenterol*. 2014;20(23 PG-7416-7423):7416-23.

#### D.1.8. Commentary/ editorials/ opinion:

- Brandt LJ. Editorial commentary: Fecal microbiota transplantation: Patient and physician attitudes. *Clin Infect Dis*. 2012;55(12 PG-1659-1660):1659–60.
- Glauser W. Risk and rewards of fecal transplants. *CMAJ*. 2011;183(5 PG-541-542):541–2.
- Goyal H, Singla U. Infectious diseases society of america or american college of gastroenterology guidelines for treatment of clostridium difficile infection: Which one to follow? *Am J Med*. 2015;128(4 PG-).
- Hecht GA, Blaser MJ, Gordon J, Kaplan LM, Knight R, Laine L, et al. What is the value of a food and drug administration investigational new drug application for fecal microbiota transplantation to treat clostridium difficile infection? *Clin Gastroenterol Hepatol*. 2014;12(2 PG-289-291):289–91.
- Hellems R, Naegels S, Holvoet J. Fecal transplantation for recurrent Clostridium difficile colitis, an underused treatment modality. *Acta Gastroenterol Belg*. 2009;72(2 PG-269-270):269–70.
- Heuer AH. Fecal transplantation with side effect: Bearer of hope for Clostridium difficile patients is still insufficiently researched. *Dtsch Apotheker Zeitung*. 2016;156(46 PG-).
- Johnson S, Gerding DN. Fecal Fixation: Fecal Microbiota Transplantation for Clostridium difficile Infection. *Clin Infect Dis*. 2016;9(PG-09):9.
- Karakan T. Fecal microbiota transplant in immunocompromised patients: Encouraging results in a vulnerable population. *Turkish J Gastroenterol*. 2014;25(3 PG-346):346.
- Keller JJ, Van Nood E, Speelman P, Kuijper EJ. Application of feces transplantation for treatment of relapsing Clostridium difficile infection. *Antonie van Leeuwenhoek, Int J Gen Mol Microbiol*. 2009;95(PG-40-41):40–1.
- Kellermayer R. Burdening questions about clostridium difficile in pediatric inflammatory bowel diseases. *J Pediatr Gastroenterol Nutr*. 2015;60(4 PG-421-422):421–2.
- Kelly CR. Editorial: A simple faecal preparation protocol for faecal microbiota transplantation. *Aliment Pharmacol Ther*. 2015;41(3 PG-320):320.
- Kuperman AA, Koren O. The era of fecal microbiota transplantation. *Isr Med Assoc J*. 2015;17(8 PG-515-516):515–6.
- Laffin M, Millan B, Madsen KL. Fecal microbial transplantation as a therapeutic option in patients colonized with antibiotic resistant organisms. *Gut Microbes*. 2017;(PG-0):0.
- Louie T, Adams PC. Nature's therapy for recurrent Clostridium difficile diarrhea. *Can J Gastroenterol*. 2008;22(5 PG-455-456):455–6.
- Lynch S V. Fecal microbiota transplantation for recurrent clostridium difficile infection in pediatric patients: Encouragement wrapped in caution. *J Pediatr Gastroenterol Nutr*. 2015;60(1 PG-1-3):1–3.
- Malani PN, Rao K. Expanded evidence for frozen fecal microbiota transplantation for clostridium difficileinfection a fresh take. *JAMA - J Am Med Assoc*. 2016;315(2 PG-137-138):137–8.

- Mardani M. Intestinal microbiota transplantation for recurrent *Clostridium difficile* infection. *Iran J Clin Infect Dis*. 2011;6(3 PG-103):103.
- Marsh JW, Curry SR. Therapeutic approaches for *clostridium difficile* infections. *Curr Protoc Microbiol*. 2013;(SUPPL.30) (no pagination)(9A.3 PG-).
- Mayor S. Donor faecal transplantation is highly curative in recurrent *C difficile* infection, trial finds. *BMJ*. 2016;354 (no pagination)(i4638 PG-).
- McFarland L V. The role of compassion in the practice of evidence-based medicine. *Am J Gastroenterol*. 2012;107(5 PG-768-769):768–9.
- McKinney M. Despite “eww” factor... fecal transplants gain ground against *C. diff*. *Mod Healthc*. 2013;43(4 PG-12-13):12–3.
- McKinney M. FDA slaps regs on fecal transplants. Increased steps for *C. diff* treatment draw mixed reactions from providers. *Mod Healthc*. 2013;43(21 PG-10):10.
- Nieuwdorp M. Faecal microbiota transplantation. *Br J Surg*. 2014;101(8 PG-887-8):887–8.
- Olson DC, Scobey MW. The Challenge of *Clostridium difficile* Infection. *N C Med J*. 2016;77(3 PG-206-10):206–10.
- Owens C, Broussard E, Surawicz C. Fecal microbiota transplantation and donor standardization. *Trends Microbiol*. 2013;21(9 PG-443-445):443–5.
- Pakyz AL, Moczygemba LR, Vanderwielen LM, Edmond MB. Fecal microbiota transplantation for recurrent *Clostridium difficile* infection: The patient experience. *Am J Infect Control*. 2016;44(5 Supplement PG-554-559):554–9.
- Pamer EG. Fecal microbiota transplantation: Effectiveness, complexities, and lingering concerns. *Mucosal Immunol*. 2014;7(2 PG-210-214):210–4.
- Patel K V, Digby-Bell JL, Goel RM, Henry N, Sanderson JD, Irving PM, et al. Faecal microbiota transplantation: Implementing a new treatment for recurrent/refractory *clostridium difficile* infection using banked stool in a tertiary UK centre. *Gut*. 2015;64(PG-A23):A23.
- Patel K, Spector TD. Estimating the risks of faecal transplants. *J Hosp Infect*. 2016;92(2 PG-128-129):128–9.
- Rutecki GW. An “aesthetically unappealing” transplant: Fecal microbiota. *Consultant*. 2013;53(5 PG-338):338.
- Sachs RE, Edelstein CA. Ensuring the safe and effective FDA regulation of fecal microbiota transplantation. *J Law Biosci*. 2015;2(2 PG-396-415):396–415.
- Senior K. Faecal transplantation for recurrent *C difficile* diarrhoea. *Lancet Infect Dis*. 2013;13(3 PG-200-201):200–1.

Sokol H. Toward Rational Donor Selection in Faecal Microbiota Transplantation for IBD. *J Crohns Colitis*. 2016;10(4 PG-375-6):375–6.

#### D.1.9. Letters

Anonymous. Fecal Transplant for *Clostridium difficile*-Reply. *Arch Intern Med*. 2012;172(10 PG-825-6):825–6.

Green MR, Acharya UH, Yeager AM. Is fidaxomicin the drug of choice for treating *clostridium difficile* - Associated diarrhea in patients with cancer? *J Clin Oncol*. 2013;31(34 PG-4376-4378):4376–8.

Gundacker ND, Morrow CD, Rodriguez M. Letter: a simple out-patient faecal microbiota transplant technique. *Aliment Pharmacol Ther*. 2016;44(1 PG-101):101.

Gutman J, Kurchin A. Split donation fecal microbiota transplantation. *Am J Gastroenterol*. 2013;108(10 PG-1659-1660):1659–60.

Ho N, Prasad V. *Clostridium difficile* diarrhea and fecal transplantation. *J Clin Gastroenterol*. 2011;45(8 PG-742-743):742–3.

Ho N, Prasad V. Lacking the incentive to cure? Recurring *clostridium difficile* diarrhea and our reluctance to use fecal transplantation. *J Clin Gastroenterol*. 2011;45(4 PG-379-380):379–80.

Hodes RM. Fecal flora reconstitution for amyotrophic lateral sclerosis. *J Clin Gastroenterol*. 2013;47(7 PG-655):655.

Hogenauer C, Kump PK, Krause R. Tempered enthusiasm for fecal transplantation? *Clin Infect Dis*. 2014;59(9 PG-1348-1349):1348–9.

Ianiro G, Gasbarrini A, Cammarota G. Letter: Faecal microbiota transplantation - Not a one-size-fits-all approach. *Aliment Pharmacol Ther*. 2014;40(1 PG-119):119.

Jouhten H, Mattila E, Arkkila P, Satokari R. Reduction of Antibiotic Resistance Genes in Intestinal Microbiota of Patients With Recurrent *Clostridium difficile* Infection After Fecal Microbiota Transplantation. *Clin Infect Dis*. 2016;63(5 PG-710-711):710–1.

Kassam Z, Lee CH, Yuan Y, Hunt RH. Navigating long-term safety in fecal microbiota transplantation. *Am J Gastroenterol*. 2013;108(9):1538.

Konstantinov SR, Peppelenbosch MP. Fecal microbiota transfer may increase irritable bowel syndrome and inflammatory bowel diseases-associated bacteria. *Gastroenterology*. 2013;144(4 PG-e19-e20):e19–20.

Malnick SD, Oppenheim A, Melzer E. Immune Thrombocytopenia Caused by Fecal Microbial Transplantation in a Patient With Severe Recurrent *Clostridium difficile* Infection. *J Clin Gastroenterol*. 2015;49(10 PG-888-9):888–9.



- Malnick SDH, Oppenheim A, Melzer E. Immune thrombocytopenia caused by fecal microbial transplantation in a patient with severe recurrent clostridium difficile infection. *J Clin Gastroenterol*. 2015;49(10 PG-888-889):888–9.
- Martin L. Modified fecal transplantation. *J Clin Gastroenterol*. 2011;45(8 PG-742):742.
- Matuchansky C. Fecal microbiota transplantation: the case of immunocompromised patients. *Am J Med*. 2015;128(3 PG-e21):e21.
- Mawer DP, Wilcox MH. Clarifying the management of Clostridium difficile infection. *BMJ*. 2015;351(PG-h6130):h6130.
- Mittal C, Miller N, Meighani A, Hart BR, John A, Ramesh M. Fecal microbiota transplant for recurrent Clostridium difficile infection after peripheral autologous stem cell transplant for diffuse large B-cell lymphoma. *Bone Marrow Transplant*. 2015;50(7 PG-1010):1010.
- Pecere S, Sabatelli M, Fantoni M, Ianiro G, Gasbarrini A, Cammarota G. Letter: Faecal microbiota transplantation in combination with fidaxomicin to treat severe complicated recurrent Clostridium difficile infection. *Aliment Pharmacol Ther*. 2015;42(8 PG-1030):1030.
- Schwartz M, Gluck M, Koon S. Norovirus gastroenteritis after fecal microbiota transplantation for treatment of clostridium difficile infection despite asymptomatic donors and lack of sick contacts. *Am J Gastroenterol*. 2013;108(8 PG-1367):1367.
- Sha S, Wu K. Letter: Faecal microbiota transplantation - Not a one-size-fits-all approach; Authors' reply. *Aliment Pharmacol Ther*. 2014;40(1 PG-119-120):119–20.
- Sohn KM, Cheon S, Kim YS. Can Fecal Microbiota Transplantation (FMT) Eradicate Fecal Colonization with Vancomycin-Resistant Enterococci (VRE)? *Infect Control Hosp Epidemiol*. 2016;37(12 PG-1519-1521):1519–21.
- Solari PR, Fairchild PG, Noa LJ, Wallace MR. Tempered enthusiasm for fecal transplant. *Clin Infect Dis*. 2014;59(2 PG-319):319.
- Solari PR, Wallace MR. Reply to Krause et al. *Clin Infect Dis*. 2014;59(9 PG-1349):1349.
- Spector T, Knight R. Authors' reply to Mawer and Wilcox and Mullish and Williams. *BMJ*. 2015;351(PG-h6132):h6132.
- Spector T, Knight R. Faecal transplants. *BMJ*. 2015;351 (no pagination)(h5149 PG-).
- Tacke D, Wisplinghoff H, Kretzschmar A, Farowski F, Koehler P, Herweg J, et al. First implementation of frozen, capsulized faecal microbiota transplantation for recurrent Clostridium difficile infection into clinical practice in Europe. *Clin Microbiol Infect*. 2015;21(11 PG-e82-e84):e82–4.
- Tacke D, Wisplinghoff H, Kretzschmar A, Farowski F, Koehler P, Herweg J, et al. First implementation of frozen, capsulized faecal microbiota transplantation for recurrent Clostridium difficile infection into clinical practice in Europe. *Clin Microbiol Infect*. 2015;21(11 PG-e82-4):e82-4.

Wang XJ, Kraft CS, Dhere T. Use of standard donors in fecal microbiotal transplants. *South Med J*. 2015;108(1 PG-68-69):68–9.

Youngster I, Hohmann EL. Fecal microbiota transplantation for *Clostridium difficile* infection--reply. *JAMA*. 2015;313(7 PG-726):726.

#### **D.1.10. Not relevant, miscellaneous**

A.M. B, C.P. K, Berg AM, Kelly CP, Farraye FA. *Clostridium difficile* infection in the inflammatory bowel disease patient. *Inflamm Bowel Dis*. A.M. Berg, Section of Gastroenterology, Boston Medical Center, 85 East Concord St., Suite 7720, Boston, MA 02118-2338, United States. E-mail: adam.berg@bmc.org; Lippincott Williams and Wilkins (530 Walnut Street, P O Box 327, Philadelphia PA 19106-3621, United States); 2013;19(1):194–204.

Ahir HB, Marcella S, Jiang Y, Mayes A, Burnett H, Rajpura J. Systematic literature review of economic evaluations and healthcare resource utilisation studies in the treatment of *clostridium difficile* infection. *Value in Health*. 2017;20 (9):A791

Allegretti J, Eysenbach LM, El-Nachef N, Fischer M, Kelly C, Kassam Z. The Current Landscape and Lessons from Fecal Microbiota Transplantation for Inflammatory Bowel Disease: Past, Present, and Future. *Inflammatory Bowel Diseases*. 2017;23(10):1710-7.

Allegretti JR, Bry L, Gerber G, Clish C, Korzenik JR. Investigation of dysbiosis and bile salt composition associated with *c. Difficile* infection. *Am J Gastroenterol*. 2015;110(PG-S573-S574):S573–4.

Allegretti JR, Kao DH, Sitko J, Fischer M, Kassam Z. Prevalence of early antibiotic use post-fecal microbiota transplantation and corresponding risk of FMT failure. *Gastroenterology*. 2017;152 (5 Supplement 1):S342-S3.

Allegretti JR, Kassam Z, Chan WW. Small Intestinal Bacterial Overgrowth: Should Screening Be Included in the Pre-fecal Microbiota Transplantation Evaluation? *Digestive Diseases and Sciences*. 2017:1-5.

Allegretti JR, Kassam Z, Smith M, Korzenik JR, Chan WW. Irregular bowel movements following fecal microbiota transplantation (FMT) are associated with pre-existing irritable bowel syndrome but not FMT-related factors. *Gastroenterology*. 2016;1(PG-S742):S742.

Alonso CD, Braun DA, Patel I, Akbari M, Oh DJ, Jun T, et al. A multicenter, retrospective, case-cohort study of the epidemiology and risk factors for *Clostridium difficile* infection among cord blood transplant recipients. *Transpl Infect Dis*. 2017;19 (4) (no pagination)(e12728).

American Journal of Gastroenterology Lecture: Intestinal microbiota and the role of fecal microbiota transplant (FMT) in treatment of *C. difficile* infection. *American Journal of Gastroenterology*. L.J. Brandt, Montefiore Medical Center, 111 East 210th Street, Bronx, NY 10467, United States. E-mail: lbrandt@montefiore.org; Nature Publishing Group (Houndmills, Basingstoke, Hampshire RG21 6XS, United Kingdom); 2013. p. 177–85.

Anand R, Song Y, Garg S, Girotra M, Sinha A, Sivaraman A, et al. Effect of Aging on the Composition of Fecal



- Microbiota in Donors for FMT and Its Impact on Clinical Outcomes. *Dig Dis Sci*. 2017;62(4):1002-8.
- Andermann TM, Rezvani A, Bhatt AS. Microbiota Manipulation With Prebiotics and Probiotics in Patients Undergoing Stem Cell Transplantation. *Curr Hematol Malig Rep*. 2016;11(1 PG-19-28):19-28.
- Anonymous. ALS Untangled No. 21: Fecal transplants. *Amyotroph Lateral Scler Front Degener*. 2013;14(5-6-482-485):482-5.
- Anonymous. AMMI Canada-CACMID 2012 Annual Conference Abstracts. *Can J Infect Dis Med Microbiol Conf AMMI Canada CACMID*. 2012;23(no pagination PG-).
- Anonymous. Erratum: Oral, capsulized frozen fecal microbiota transplantation for relapsing clostridium difficile infection (*JAMA - Journal of the American Medical Association* (2014) 312:17 (1772-1778) DOI: 10.1001/jama.2014.13875). *JAMA - J Am Med Assoc*. 2015;313(7 PG-729):729.
- Anonymous. Faecal microbiota transplantation for ulcerative colitis. *Drug and Therapeutics Bulletin*. 2017;55(5):51.
- Arkkila P, Hillamaa A, Jalanka J, Mattila E, Anttila V, Satokari R. Long-term safety and effect on gastrointestinal symptoms of fecal microbiota transplantation. *United European Gastroenterology Journal*. 2017;5 (5 Supplement 1):A757.
- Armstrong MJ, Pathmakanthan S, Iqbal TH. Fecal microbiota transplantation for *Clostridium difficile* infection. *JAMA - J Am Med Assoc*. 2015;313(7 PG-725-726):725-6.
- Assar S, Nakhle A, Lazar M. Eat your heart out: Right heart collapse from bowel obstruction. *American Journal of Respiratory and Critical Care Medicine Conference: American Thoracic Society International Conference, ATS*. 2017;195(no pagination).
- Bagdasarian N, Rao K, Malani PN. Changing Epidemiology and Control of *Clostridium difficile* in Older Adults. *Curr Transl Geriatr Gerontol Reports*. 2013;2(3 PG-143-150):143-50.
- Bajaj JS, Fagan A, Sikaroodi M, White MB, Sterling RK, Gilles H, et al. Liver transplant modulates gut microbial dysbiosis and cognitive function in cirrhosis. *Liver Transplantation*. 2017;23(7):907-14.
- Barketi-Klai A, Hoys S, Lambert-Bordes S, Collignon A, Kansau I. Role of fibronectin-binding protein A in *Clostridium difficile* intestinal colonization. *J Med Microbiol*. 2011;60(8 PG-1155-1161):1155-61.
- Barnes D, Park KT, Smith M, Kassam Z. Feasibility of a competitively selected universal donor fecal microbiota transplantation protocol and characterization of post-transplant microbiota modification. *J Pediatr Gastroenterol Nutr*. 2016;63(PG-S142-S143):S142-3.
- Barnes DM, Park KT, Kassam Z, Smith MB. Optimizing fecal microbiota transplant: An innovative method to competitively select a universal donor. *Am J Gastroenterol*. 2015;110(PG-S985):S985.
- Baro E, Galperine T, Denies F, Lannoy D, Lenne X, Odou P, et al. Cost-Effectiveness Analysis of Five Competing Strategies for the Management of Multiple Recurrent Community-Onset *Clostridium difficile* Infection in France. *PLoS ONE*. 2017;12(1):e0170258.

- 1  
2 Bashan A, Gibson TE, Friedman J, Carey VJ, Weiss ST, Hohmann EL, et al. Universality of human microbial  
3 dynamics. *Nature*. 2016;534(7606 PG-259-262):259–62.  
4  
5  
6 Battaglioli E, Hale V, Chen J, Jeraldo P, Rekdal VM, Huq L, et al. Prophylactic fecal microbial transplant  
7 restores *clostridium difficile* colonization resistance in a dysbiotic subset of diarrhea associated human  
8 microbial communities modeled in germ free mice. *Gastroenterology*. 2017;152 (5 Supplement 1):S348.  
9  
10  
11 Baty V, Mouglin B. What about public perception of antibiotics in the era of the fecal microbiota  
12 transplantation? between the devil and the deep blue sea. *Am J Gastroenterol*. 2013;108(9 PG-1540):1540.  
13  
14 Baumgart DC. [The human microbiome]. *Dtsch Medizinische Wochenschrift*. 2015;140(19 PG-1451-6):1451–  
15 6.  
16  
17 Baumgart DC. The human microbiome. [German]. *Dtsch Medizinische Wochenschrift*. 2015;149(19 PG-1451-  
18 1456):1451–6.  
19  
20  
21 Baxter M, Ahmad T, Colville A, Sheridan R. Fatal aspiration pneumonia as a complication of fecal microbiota  
22 transplant. *Clin Infect Dis*. 2015;61(1 PG-136-137):136–7.  
23  
24  
25 Bella CJ, Coulson S, Vitetta L. Is co-prescribing a multi-strain probiotic the solution for treating and preventing  
26 proton pump inhibitor (PPIs) induced *Clostridium difficile* associated diarrhoea (CDAD) while maintaining  
27 evidence based pharmacotherapy? *Adv Integr Med*. 2014;1(1 PG-52-54):52–4.  
28  
29  
30 Benes J, Polivkova S. [Antibiotic treatment of clostridial colitis]. *Epidemiol Mikrobiol Imunol*. 2016;65(1 PG-  
31 15-24):15–24.  
32  
33  
34 Beus A. Recurrent *Clostridium difficile* infections: Significance and treatment. *Infektoloski Glas*. 2011;31(3  
35 PG-155-161):155–61.  
36  
37  
38 Bhanvadia A, Zhu R, Amarnani A, Yang J, Smith M, Grossman EB, et al. Gut microbiota profiling in patients  
39 with *clostridium difficile* infections at urban safety net hospitals: A comparison to the human microbiome  
40 project. *Gastroenterology*. 2016;1)(PG-S895-S896):S895–6.  
41  
42 Bi Z, Lu Y, Weigarden AR, Yao D, Wang L, Khoruts A, et al. Identification of p-cresol sulfate and secondary bile  
43 salts in human urine as sensitive biomarkers of fecal microbiota transplantation in R-CDI patients. *FASEB*  
44 *Journal Conference: Experimental Biology*. 2017;31(1 Supplement 1).  
45  
46  
47 Biedermann L, Rogler G. *Clostridium difficile* colitis. [German]. *Gastroenterologe*. 2014;9(4 PG-350-  
48 359):350–9.  
49  
50  
51 Biedermann L, Rogler G. *Clostridium difficile* infection. [German]. *Gastroenterologe*. 2017;12(3):237-52.  
52  
53  
54 Biedermann L, Rogler G. The intestinal microbiota: its role in health and disease. *Eur J Pediatr*. 2015;174(2  
55 PG-151-167):151–67.  
56  
57  
58 Bilal M, Khehra R, Strahotin C, Mitre R. Long-term follow-up of fecal microbiota transplantation for treatment  
59 of recurrent *clostridium difficile* infection in a dual solid organ transplant recipient. *Case Rep Gastroenterol*.  
60 2015;9(PG-156-159):156–9.

- Blosser R. Probiotic infusion during colonoscopy is an effective therapeutic alternative for refractory or recurrent *C. Difficile* colitis. *Am J Gastroenterol*. 2013;108(PG-S181):S181.
- Breaux JL, Ray A, Michael S. Characteristics of patients undergoing fecal microbiota transplantation for *clostridium difficile* infection: One institution's story. *Gastroenterology*. 2017;152 (5 Supplement 1):S633.
- Broecker F, Klumpp J, Moelling K. Long-term microbiota and virome in a Zurich patient after fecal transplantation against *Clostridium difficile* infection. E-mail: subscrip@blackwellpub.com: Blackwell Publishing Inc.; 2016;(1372 1 PG-29-41):29–41.
- Broecker F, Russo G, Klumpp J, Moelling K. Stable core virome despite variable microbiome after fecal transfer. *Gut Microbes*. 2016;(PG-1-7):1–7.
- Broecker F, Russo G, Klumpp J, Moelling K. Stable core virome despite variable microbiome after fecal transfer. *Gut Microbes*. 2017;8(3):214-20.
- Brumboiu MI, Poolay Mootien C, Petrus DI, Tzaneva V, Manole FI. The host defense mechanisms and diarrhea with *Clostridium difficile*: Who are the patients? *Eur J Clin Invest*. 2015;45(PG-55):55.
- Bruminhent J, Cawcutt KA, Thongprayoon C, Petterson TM, Kremers WK, Razonable RR. Epidemiology, risk factors, and outcome of *Clostridium difficile* infection in heart and heart-lung transplant recipients. *Clinical Transplantation*. 2017;31 (6) (no pagination)(e12968).
- Budree S, Elliott RJ, Rao S, Njenga M, Ladha A, Allegretti JR, et al. Donor stool processing time: The effect on the intestinal microbiome and clinical outcomes of fecal microbiota transplantation in *clostridium difficile* infection. *Gastroenterology*. 2017;152 (5 Supplement 1):S1006.
- Budree S, Panchal P, Tu E, Kahn S, Kassam Z, Osman M. Access and effectiveness of fecal microbiome transplantation for recurrent *clostridium difficile* infection in the United States pediatric population: A universal stool bank experience. *Journal of Pediatric Gastroenterology and Nutrition*. 2017;64:68-9.
- Budree S, Rao S, Allegretti JR, Fischer M, Kelly CR, Smith M, et al. The association of donor stool consistency by bristol stool scale on microbial profile and clinical outcomes of fecal microbiota transplantation in *clostridium difficile* infection. *Gastroenterology*. 2017;152 (5 Supplement 1):S630.
- Budree S, Tu E, Leith T, Allegretti JR, Rao S, Day R, et al. The association of stool donor diet on microbial profile and clinical outcomes of fecal microbiota transplantation in *clostridium difficile* infection. *Gastroenterology*. 2017;152 (5 Supplement 1):S630-S1.
- Budree S, Wong WF, Tu E, Rao S, Allegretti JR, Fischer M, et al. Do specific bacteria drive clinical cure in fecal microbiota transplantation for *clostridium difficile* infection?: Clinical, microbial and metabolomic characterization of universal FMT donors. *Gastroenterology*. 2017;152 (5 Supplement 1):S349.
- Bush BR, Rogers MB, Firek B, Kufen A, Jackson Z, Morowitz M, et al. Dynamic changes in the intestinal microbiota following fecal microbiota transplantation for refractory inflammatory bowel disease in children. *Gastroenterology*. 2016;1(PG-S541):S541.
- C. B, G.B. G, M. R, E. A-V, E.O. P, Brace C, et al. Microbial composition analysis of *Clostridium difficile* infections in an ulcerative colitis patient treated with multiple fecal microbiota transplantations. *J Crohns*

- Colitis. Netherlands: Brace, Chantalle. Dept. Medicine, GI Diseases Research Unit, Queen's University, Kingston, ON, Canada. Electronic address: chantalle.brace@queensu.ca.; 2014;8(9):1133–7.
- Callejas-Diaz A, Gea-Banacloche JC. Clostridium difficile: Deleterious impact on hematopoietic stem cell transplantation. *Curr Hematol Malig Rep*. 2014;9(1 PG-85-90):85–90.
- Cammarota G, Ianiro G, Magalini S, Gasbarrini A, Gui D. Decrease in surgery for clostridium difficile infection after starting a program to transplant fecal microbiota. *Ann Intern Med*. 2015;163(6 PG-487-488):487–8.
- Cammarota G, Pecere S, Ianiro G, Masucci L, Curro D. Principles of DNA-Based Gut Microbiota Assessment and Therapeutic Efficacy of Fecal Microbiota Transplantation in Gastrointestinal Diseases. *Dig Dis*. 2016;34(3 PG-279-285):279–85.
- Carlet J. The gut is the epicentre of antibiotic resistance. *Antimicrob Resist Infect Control*. 2012;1 (no pagination)(39 PG-).
- Cattaneo C. Gram-positive infections: New and old pathogens in haematological patient. *Haematologica*. 2015;100(PG-191-193):191–3.
- Chehoud C, Dryga A, Hwang Y, Nagy-Szakal D, Hollister EB, Luna RA, et al. Transfer of viral communities between human individuals during fecal microbiota transplantation. *MBio*. 2016;7 (2) (no pagination)(e00322-16 PG-).
- Chen LA, Hourigan S, Radin A, Weidner M, Oliva-Hemker MM, Sears C, et al. Bile acid composition changes over 6 months following fecal microbiota transplantation in children with recurrent C. Difficile infections 2016 ACG presidential poster award. *Am J Gastroenterol*. 2016;111(PG-S453-S454):S453–4.
- Chintalapally R, Kukkadapu T, Parikh S, Mangaonkar AA, Boppidi HR, Kota V, et al. Clostridium difficile infection in adult patients with acute myeloid leukemia: Incidence, recurrence, and outcomes. *J Clin Oncol Conf*. 2015;33(15 SUPPL. 1 PG-).
- Chitnavis M V, Hays RA. Acute-on-chronic neutropenic fever as a complication following fecal microbiota transplantation (FMT) in a patient with shwachman-diamond syndrome. *Am J Gastroenterol*. 2015;110(PG-S492):S492.
- Cho J, Sampathkumar P, Seville MT, Kashyap P. Detecting outcomes from clostridium difficile screening on admission in patients admitted to a bone marrow transplant unit. *Gastroenterology*. 2017;152 (5 Supplement 1):S345.
- Chu ND, Smith MB, Perrotta AR, Kassam Z, Alm EJ. Profiling Living Bacteria Informs Preparation of Fecal Microbiota Transplantations. *PLoS ONE*. 2017;12(1):e0170922.
- Chuong KH, O'Doherty KC, Secko DM. Media Discourse on the Social Acceptability of Fecal Transplants. *Qual Health Res*. 2015;25(10 PG-1359-71):1359–71.
- Cicerone C, Bruno G, Lamonaca L, D'Abramo A, Oliva A, Zingaropoli MA, et al. Fecal microbiota transplantation via enema for recurrent clostridium difficile infection modulates the inflammatory host response and restores intestinal dysbiosis. *Digestive and Liver Disease*. 2017;49 (Supplement 2):e102.

- Cicerone C, Bruno G, Lamonaca L, Trancassini M, Corazziari ES. Fecal microbiota transplantation for recurrent clostridium difficile infection: Transplant protocol by retention enema and preliminary results. *Digestive and Liver Disease*. 2017;49 (Supplement 2):e175.
- Claassen E. Healthy microbiota by probiotics or fecal transplantation prevent diarrhea: Probiotics are beneficial in case of *Clostridium difficile* infection. [Dutch]. *Pharm Weekbl*. 2014;149(10 PG-16-17):16–7.
- Claes IJJ, Vargas Garcia CE, Lebeer S. Novel opportunities for the exploitation of host-microbiome interactions in the intestine. *Curr Opin Biotechnol*. 2015;32(PG-28-34):28–34.
- Cocanour CS. Best strategies in recurrent or persistent *Clostridium difficile* infection. *Surg Infect (Larchmt)*. 2011;12(3 PG-235-239):235–9.
- Cohen NA, Maharshak N. Novel Indications for Fecal Microbial Transplantation: Update and Review of the Literature. *Dig Dis Sci*. 2017;62(5):1131-45.
- Collins SM, Kassam Z, Bercik P. The adoptive transfer of behavioral phenotype via the intestinal microbiota: Experimental evidence and clinical implications. *Curr Opin Microbiol*. 2013;16(3 PG-240-245):240–5.
- Comito D, Cascio A, Romano C. Microbiota biodiversity in inflammatory bowel disease. *Ital J Pediatr*. 2014;40 (1) (no pagination)(32 PG-).
- Costello SP, Tucker EC, La Brooy J, Schoeman MN, Andrews JM. Establishing a fecal microbiota transplant service for the treatment of clostridium difficile infection. *Clin Infect Dis*. 2016;62(7 PG-908-914):908–14.
- Costello SP, Van Der Poorten D, Andrews JM. Fecal microbiota transplantation for recurrent *Clostridium difficile* infection: When regulatory affairs do not keep pace with evidence-based medicine. *Journal of Gastroenterology and Hepatology (Australia)*. 2017;32:156-7.
- Costello SP, Waters O, Bryant RV, Katsikeros R, Makanyanga J, Schoeman M, et al. Short duration, low intensity, pooled fecal microbiota transplantation induces remission in patients with mild/moderately active ulcerative colitis: A randomised controlled trial. *Gastroenterology*. 2017;152 (5 Supplement 1):S198-S9.
- Crabtree S, Gupta J. Knowledge and attitudes towards faecal bacteriotherapy on ITU. *Intensive Care Med*. 2014;1(PG-S106):S106.
- Culligan EP, Sleator RD. Advances in the microbiome: Applications to clostridium difficile infection. *J Clin Med*. 2016;5 (9) (no pagination)(83 PG-).
- Curry SR. *Clostridium difficile*. *Clin Lab Med*. 2017;37(2):341-69.
- Davidovics ZH, Vance K, Etienne N, Hyams JS. Fecal Transplantation Successfully Treats Recurrent D-Lactic Acidosis in a Child with Short Bowel Syndrome. *Journal of Parenteral and Enteral Nutrition*. 2017;41(5):896-7.
- de Groot PF, Frissen MN, de Clercq NC, Nieuwdorp M. Fecal microbiota transplantation in metabolic syndrome: History, present and future. *Gut Microbes*. 2017;8(3):253-67.

- De Leon LM, Watson JB, Kelly CR. Transient flare of ulcerative colitis after fecal microbiota transplantation for recurrent *Clostridium difficile* infection. *Clin Gastroenterol Hepatol*. 2013;11(8 PG-1036-1038):1036–8.
- De Santis S, Cavalcanti E, Mastronardi M, Jirillo E, Chieppa M. Nutritional keys for intestinal barrier modulation. *Front Immunol*. 2015;6 (DEC) (no pagination):612 PG-).
- del Campo-Moreno R, Alarcon-Cavero T, D'Auria G, Delgado-Palacio S, Ferrer-Martinez M. Enfermedades Infecciosas y Microbiología Clínica. 2017;06.
- Dennis M, Salpeter MJ, Hota S. Low awareness but positive attitudes toward fecal transplantation in ontario physicians. *Can J Infect Dis Med Microbiol*. 2015;26(1 PG-30-32):30–2.
- Dinh A, Bouchand F, Le Monnier A. [Current treatment and epidemiology of *Clostridium difficile* infections]. *Rev Med Interne*. 2015;36(9 PG-596-602):596–602.
- Dinh A, Bouchand F, Le Monnier A. Current treatment and epidemiology of *Clostridium difficile* infections. [French]. *Rev Med Interne*. 2015;36(9 PG-596-602):596–602.
- Donnelly SC. Elements: In this month's issue. *Qjm*. 2015;108(5 PG-351-351):351.
- Dore J, Multon MC, Behier JM, participants of Giens Xxxii RTN. The human gut microbiome as source of innovation for health: Which physiological and therapeutic outcomes could we expect? *Therapie*. 2017;72(1):21-38.
- Downs IA, Brandt LJ, Oneto C, Feuerstadt P, Aroniadis OC. Perceptions of fecal microbiota transplantation for diarrhea predominant irritable bowel syndrome. *Am J Gastroenterol*. 2016;111(PG-S1250-S1251):S1250–1.
- Drekonja DM. *Clostridium difficile* infection: Current, forgotten and emerging treatment options. *J Comp Eff Res*. 2014;3(5 PG-547-557):547–57.
- Dryden G. Use of serum-derived bovine immunoglobulin/protein isolate (SBI) to manage refractory ulcerative colitis symptoms and avoid surgery. *Am J Gastroenterol*. 2014;109(PG-S440):S440.
- Dunwoody R, Steel A, Landy J, Simmonds N. *Clostridium difficile* and cystic fibrosis: Management strategies and the role of faecal transplantation. *Paediatric Respiratory Reviews*. 2017.
- DuPont HL. Diagnosis and management of *Clostridium difficile* infection. *Clin Gastroenterol Hepatol*. 2013;11(10 PG-1216-1223):1216–23.
- Ebigbo A, Messmann H. [Challenges of *Clostridium difficile* infection]. *Med Klin Intensivmed Notfmed*. 2013;108(8 PG-624-7):624–7.
- Ebigbo A, Messmann H. Challenges of *Clostridium difficile* infection. [German, English]. *Medizinische Klin - Intensivmed und Notfallmedizin*. 2013;108(8 PG-624-627):624–7.
- El Feghaly RE, Bangar H, Haslam DB. The molecular basis of *Clostridium difficile* disease and host response. *Curr Opin Gastroenterol*. 2015;31(1 PG-24-29):24–9.



- El Feghaly RE, Stauber JL, Deych E, Gonzalez C, Tarr PI, Haslam DB. Markers of intestinal inflammation, not bacterial burden, correlate with clinical outcomes in clostridium difficile infection. *Clin Infect Dis*. 2013;56(12 PG-1713-1721):1713–21.
- El-Nachef N, Piceno YM, Kassam Z, Zydek M, Ablaza AJ, Leith T, et al. Fecal microbiota transplantation is safe and effective in chronic pouchitis patients. *Gastroenterology*. 2017;152 (5 Supplement 1):S1009.
- Elias J, Lichtman J, Sonnenburg J. Quantifying dynamic host-microbiota signatures of antibiotic-associated GI infection: What can the host proteome tell us? *FASEB Journal Conf Exp Biol*. 2015;29(1 Meeting Abstracts PG-).
- Fang Y, Chen J, Yu J, Luo Y, Lou J. The preliminary investigation of fecal microbiota transplantation for pediatric recurrent chronic bowel disease and literary review. *J Pediatr Gastroenterol Nutr*. 2016;62(PG-233-234):233–4.
- Farrell JJ, Martin D, Bogner A, Thompson S V, Taylor AM, Swanson KS, et al. Evolving composition of the human intestinal microbiota following fecal transplantation. *Gastroenterology*. 2016;1(PG-S430):S430.
- Fasullo MJ, Al-Azzawi Y, Abergel J. Microscopic colitis after fecal microbiota transplant. *ACG Case Reports Journal*. 2017;4 (15) (no pagination)(e87).
- Fenner I, Lensing C, Katz A, Petersen H. Clostridium difficile - Diagnosis by culture or PCR? *Int J Med Microbiol*. 2009;299(PG-43):43.
- Ferm S, Varadi N, Fisher C, Gutkin E. Serum-derived bovine immunoglobulin as novel adjunct in complicated clostridium difficile colitis treatment. *ACG Case Reports Journal*. 2017;4 (10) (no pagination)(e64).
- Fischer M, Kao DH, Phelps EL, Smith JD, Roach B, Kassam Z, et al. Should we recommend anti-clostridium difficile antibiotic or probiotic prophylaxis?: Risk of clostridium difficile infection with systemic antimicrobial therapy following successful fecal microbiota transplant. *Gastroenterology*. 2017;152 (5 Supplement 1):S1005.
- Fischer M, Kelly CR, Phelps EL, Wang E, Roach B, Smith JD, et al. Quality of bowel preparation does not affect outcome of fecal microbiota transplantation for the therapy clostridium difficile infection. *Gastroenterology*. 2017;152 (5 Supplement 1):S1004-S5.
- Fischer M, Khan M, Phelps EL, Safdar N, Misch EA, Kaur N, et al. Fecal microbiota transplantation is safe and effective for the treatment of clostridium difficile infection in solid organ transplant recipients. *Gastroenterology*. 2017;152 (5 Supplement 1):S1005.
- Fischer M, Phelps E, Cook G, Sipe B, Rex DK, Xu H. Clostridium difficile pcr testing post fecal microbiota transplantation (FMT) predicts success. *Am J Gastroenterol*. 2015;110(PG-S580):S580.
- Fischer M, Rex DK, Sipe BW. Letter: faecal microbiota transplantation in combination with fidaxomicin to treat severe complicated recurrent Clostridium difficile infection--authors' reply. *Aliment Pharmacol Ther*. 2015;42(8 PG-1031):1031.
- Fischer M, Rex DK, Sipe BW. Letter: Faecal microbiota transplantation in combination with fidaxomicin to treat severe complicated recurrent Clostridium difficile infection - Authors' reply. *Aliment Pharmacol Ther*. 2015;42(8 PG-1031):1031.

- Fischer M, Sipe B, Torbeck M, Xu H, Kassam Z, Allegretti JR. Does fecal microbiota transplantation from an obese donor lead to weight gain? A case series of 70 recipients. *Gastroenterology*. 2017;152 (5 Supplement 1):S1004.
- Fischer M, Torbeck M, Cook G, Mazur S, Phelps E, Sipe B, et al. Weight change after fecal microbiota transplantation (FMT) is not associated with donor body mass index (BMI). *Am J Gastroenterol*. 2015;110(PG-S585):S585.
- Flamaing J. Treatment of *Clostridium difficile* associated diarrhoea: Guidelines. [Dutch]. *Tijdschr Geneesk*. 2015;71(23 PG-1592-1595):1592–5.
- Floch MH, Walker WA, Madsen K, Sanders ME, Macfarlane GT, Flint HJ, et al. Recommendations for probiotic use - 2011 update. *J Clin Gastroenterol*. 2011;45(SUPPL. 3 PG-S168-S171):S168–71.
- Founas A, Wakade Z, Ramachanran P, Ghodoussi A, Tocco J, Moore J, et al. Small bowel obstruction successfully treated with lubiprostone. *Am J Gastroenterol*. 2015;110(PG-S404):S404.
- Freedberg DE, Abrams JA. Recent therapeutic advances in gastroenterology and hepatology. *Adv Ther*. 2013;30(10 PG-855-857):855–7.
- Freedberg DE, Toussaint NC, Ratner AJ, Whittier S, Wang TC, Wang H, et al. Proton pump inhibitors alter specific taxa in the human fecal microbiome: Results of a crossover trial. *Gastroenterology*. 2015;127(PG-S619):S619.
- Friedman-Korn T, Livovsky DM, Maharshak N, Aviv Cohen N, Paz K, Bar-Gil Shitrit A, et al. Fecal Transplantation for Treatment of *Clostridium Difficile* Infection in Elderly and Debilitated Patients. *Digestive Diseases and Sciences*. 2017;1-6.
- Fuentes S, van Nood E, Tims S, Heikamp-de Jong I, ter Braak CJ, Keller JJ, et al. Reset of a critically disturbed microbial ecosystem: faecal transplant in recurrent *Clostridium difficile* infection. *ISME J*. 2014;8(8 PG-1621-1633):1621–33.
- Fujimori S. What are the effects of proton pump inhibitors on the small intestine? *World J Gastroenterol*. 2016;22(22 PG-6817-6819):6817–9.
- Fujimori S. What are the effects of proton pump inhibitors on the small intestine? *World J Gastroenterol*. 2015;21(22 PG-6817-6819):6817–9.
- Galperine T, Sokol H, Guery B. Fecal microbiota transplantation: Do we need harmonization? *Clin Infect Dis*. 2017;64(9):1292.
- Gasbarrini G, Bonvicini F, Gramenzi A. Probiotics history. *J Clin Gastroenterol*. 2016;50(PG-S116-S119):S116–9.
- Gedgudas R, Urba M, Petkevicius V, Jonaitis L, Kiudelis G, Kupcinskas L, et al. First case series of fecal microbiota transplantation for recurrent *clostridium difficile* infection in baltic countries. *United European Gastroenterology Journal*. 2017;5 (5 Supplement 1):A313.
- Gianotti RJ, Moss AC. Fecal microbiota transplantation: From *clostridium difficile* to inflammatory bowel



disease. *Gastroenterology and Hepatology*. 2017;13(4):209-13.

Gibson D, Kendrick S, Simpson E, Costello D, Davis R, Szetela A, et al. Implementation of xenon ultraviolet-C disinfection robot to reduce hospital acquired infections in hematopoietic stem cell transplant population. *Biology of Blood and Marrow Transplantation*. 2017;23 (3 Supplement 1):S367.

Gollwitzer ES, Marsland BJ. Microbiota abnormalities in inflammatory airway diseases - Potential for therapy. *Pharmacol Ther*. 2014;141(1 PG-32-39):32-9.

Gomollon F. [Developments in the treatment of inflammatory bowel disease: 2014 overview]. *Gastroenterol Hepatol*. 2014;37 Suppl 3(PG-14-21):14-21.

Gomollon F. Developments in the treatment of inflammatory bowel disease: 2014 overview. [Spanish]. *Gastroenterol Hepatol*. 2014;37(S3 PG-14-21):14-21.

Gorkiewicz G, Wurm P, Hogenauer C, Spindelbock W. Life-threatening antibiotic-associated enterocolitis and severe dysbiosis in critically ill intensive care unit patients. *Virchows Arch*. 2015;1(PG-S37):S37.

Gotz VP, Rand KH. Medical management of antimicrobial-associated diarrhea and colitis. *Pharmacother J Hum Pharmacol Drug Ther*. 1982;2(2 PG-100-9):100-9.

Goudarzi M, Seyedjavadi SS, Goudarzi H, Mehdizadeh Aghdam E, Nazeri S. Clostridium difficile Infection: Epidemiology, Pathogenesis, Risk Factors, and Therapeutic Options. *Scientifica (Cairo)*. 2014;2014(PG-916826):916826.

Goyal A, Yeh A, Siebold L, Calabro K, Firek B, Bush BR, et al. Clinical efficacy and microbiome findings following fecal microbiota transplant in children with refractory inflammatory bowel disease. *Gastroenterology*. 2017;152 (5 Supplement 1):S959.

Graness N, Swidsinski A, Schusser GF. Equine fecal microbiota in association with systemic use of antimicrobial drugs in horses with acute colitis. *Equine Veterinary Education*. 2017;29:16.

Greathouse KL, Harris CC, Bultman SJ. Dysfunctional families: Clostridium scindens and secondary bile acids inhibit the growth of clostridium difficile. *Cell Metab*. 2015;21(1 PG-9-10):9-10.

Groschel DH. Clostridium difficile infection. *Crit Rev Clin Lab Sci*. 1996;33(3 PG-203-45):203-45.

Groschel DHM. Clostridium difficile infection. *Crit Rev Clin Lab Sci*. 1996;33(3 PG-203-245):203-45.

Habib I, Huq N, Muddana V. Standardized openbiome product as a treatment for clostridium difficile infections: A single center experience. *Gastroenterology*. 2017;152 (5 Supplement 1):S951.

Han S, Lee K, Lee KA, Paik H, Lee J, Kim M, et al. Importance of acquisition of carbapenemase (KPC)-producing enterobacteriaceae in solid organ transplant recipients: A single-center experience. *American Journal of Transplantation*. 2017;17:327.

He Z, Zhang F. Principle, Protocol and Risk Management of Chinese Fecal Microbiota Bank. [Chinese]. *Chinese Journal of Gastroenterology*. 2017;22(4):193-8.

- Hecht GA, Orenstein R, Dubberke ER, Lee C, Khanna S. Lack of association with patient demographics and outcomes in punch CD 2, a randomized controlled trial of RBX2660, a microbiota-based drug for recurrent clostridium difficile infection. *Gastroenterology*. 2017;152 (5 Supplement 1):S951-S2.
- Hecker MT, Ho E, Donskey CJ. Fear of Failure: Engaging Patients in Antimicrobial Stewardship after Fecal Transplantation for Recurrent Clostridium difficile Infection. *Infect Control Hosp Epidemiol*. 2017;38(1 PG-127-129):127–9.
- Hell M, Bernhofer C, Stalzer P, Kern JM, Claassen E. Probiotics in Clostridium difficile infection: Reviewing the need for a multistrain probiotic. *Benef Microbes*. 2013;4(1 PG-39-51):39–51.
- Hove H, Tvede M, Mortensen PB. Antibiotic-associated diarrhoea, Clostridium difficile, and short-chain fatty acids. *Scand J Gastroenterol*. 1996;31(7 PG-688-693):688–93.
- Hrebinko K, Zuckerbraun BS. Clostridium difficile: What the surgeon needs to know. *Seminars in Colon and Rectal Surgery*. 2017.
- Iacob T, Taulescu DF, Dumitrascu DL. Therapy of the postinfectious irritable bowel syndrome: An update. *Clujul Medical*. 2017;90(2):133-8.
- Ianiro G, Masucci L, Simonelli C, Sanguinetti M, Gasbarrini A, Cammarota G. Single-infusion fecal microbiota transplantation is not effective in treating severe clostridium difficile infection. *United European Gastroenterology Journal*. 2017;5 (5 Supplement 1):A155
- iations across the United States: A 10-year nationwide analysis. *Gastrointestinal Endoscopy*. 2017;85 (5 Supplement 1):AB246-AB7.
- Jackson M, Olefson S, MacHan JT, Kelly CR. A high rate of alternative diagnoses in patients referred for presumed clostridium difficile infection. *J Clin Gastroenterol*. 2016;50(9 PG-742-746):742–6.
- Jansen JW. Fecal microbiota transplant vs oral vancomycin taper: Important undiscussed limitations. *Clin Infect Dis*. 2017;64(9):1292-3.
- Jiang ZD, Alexander A, Ke S, Valilis EM, Hu S, Li B, et al. Stability and efficacy of frozen and lyophilized fecal microbiota transplant (FMT) product in a mouse model of Clostridium difficile infection (CDI). *Anaerobe*. 2017;48:110-4.
- Joseph OD, Thompson SV, Bogner A, Martin D, Farrell JJ, Swanson KS, et al. Longitudinal study of the human gastrointestinal microbiota following fecal microbiota transplant (FMT) for clostridium difficile infections. *FASEB Journal Conference: Experimental Biology*. 2017;31(1 Supplement 1).
- Joshi NM, Goodhand J, Alazawi W, Das S, Wilks M, Rampton D. Predicting treatment failure in C. difficile infection: A prospective observational cohort study. *Gut*. 2016;65(PG-A209):A209.
- Jump RL, Pultz MJ, Donskey CJ. Vegetative Clostridium difficile survives in room air on moist surfaces and in gastric contents with reduced acidity: a potential mechanism to explain the association between proton pump inhibitors and C. difficile-associated diarrhea? *Antimicrob Agents Chemother*. 2007;51(8 PG-2883-7):2883–7.

- Jump RLP, Donskey CJ. Clostridium difficile in the Long-Term Care Facility: Prevention and Management Topical Collection on Infectious Diseases in the Elderly. *Curr Geriatr Reports*. 2015;4(1 PG-60-69):60–9.
- Jump RLP, Pultz MJ, Donskey CJ. Vegetative Clostridium difficile survives in room air on moist surfaces and in gastric contents with reduced acidity: A potential mechanism to explain the association between proton pump inhibitors and C. difficile-associated diarrhea? *Antimicrob Agents Chemother*. 2007;51(8 PG-2883-2887):2883–7.
- Juszczuk K, Grudlewska K, Mikucka A, Gospodarek E. Fecal microbiota transplantation - methods of treatment of recurrent Clostridium difficile infections and other diseases. *Postepy Hig Med Dosw (Online)*. 2017;71(0):220-6.
- Kao PC, Han QJ, Liu S, Li XJ, Inman KS, Chia N. Letter to the editor: The surge of type 2 diabetes mellitus in China - An international alert: Physical exercise and low-caloric diet may reduce the risks of type 2 diabetes mellitus and dementia. *Ann Clin Lab Sci*. 2016;46(1 PG-114-118):114–8.
- Karakan T. Fecal microbiota transplantation for treating recurrent hepatic encephalopathy: Ready for clinical application? *Turk J Gastroenterol*. 2017;28(5):425-6.
- Kashani A, Shih DQ. Fecal microbiota transplantation is highly effective for treatment of clostridium difficile infection in patients with inflammatory bowel disease; a meta-analysis. *Gastroenterology*. 2017;152 (5 Supplement 1):S988.
- Kassam Z, Fridman S, Burgess J, Fischer M, Amaratunga K, Edelstein C, et al. The cost-effectiveness of competing strategies for managing multiply recurrent clostridium difficile infection: Examining the impact of universal stool banks and encapsulated fecal microbiota transplantation. *Am J Gastroenterol*. 2015;110(PG-S933-S934):S933–4.
- Kassam Z, Lieberman A, Munoz R, Edelstein C, Osman M, Smith M, et al. The impact of stool banks on access to fecal microbiota transplantation for recurrent clostridium difficile infection in the United States: A geospatial analysis. *Am J Gastroenterol*. 2016;111(PG-S410):S410.
- Kassam Z, Mendolia G, Vo E, Boughari S, Njenga M, Warren K, et al. Microbial emulsion matrices: A novel method to produce stable, orally available capsules for fecal microbiota transplantation to treat clostridium difficile. *Am J Gastroenterol*. 2015;110(PG-S568-S569):S568–9.
- Kato K, Sekizuka T, Sugiyama T, Ishii Y, Kuroda M, Ohkusa T. Characterization of gut microbiome associated with improvement of ulcerative colitis after antibiotic combination therapy using fecal metagenomic analysis. *United European Gastroenterology Journal*. 2017;5 (5 Supplement 1):A264-A5.
- Kazerouni A, Burgess J, Burns LJ, Wein LM. Optimal screening and donor management in a public stool bank. *Microbiome*. 2015;3(PG-75):75.
- Kellermayer R, Balderas M, Nagy-Szakal D, Luna RA, Ihekweazu F, Queliza K, et al. Microbiome and metabolome responses to fecal microbiota transplantation for recurrent clostridium difficile infection in pediatric patients. *Gastroenterology*. 2017;152 (5 Supplement 1):S152.
- Kelly C, De Leon L, Kerstetter D, Okpara N. Barriers to greater utilization of fecal bacteriotherapy for chronic clostridium difficile infection. *Am J Gastroenterol*. 2010;105(PG-S135-S136):S135–6.

- Kelly CR, Kunde SS, Khoruts A. Guidance on preparing an investigational new drug application for fecal microbiota transplantation studies. *Clin Gastroenterol Hepatol*. 2014;12(2 PG-283-288):283–8.
- Kercsak A, Sullivan E, Sikand H. Implementation and outcomes of fecal microbiota transplantation in a four hospital system. *Pharmacotherapy*. 2015;35 (11)(PG-e274):e274.
- Khanna S, Hecht GA, Dubberke ER, Orenstein R, Lee C, Gerding DN. Alterations in microbial diversity are associated with treatment success with RBX2660, a microbiota-based drug for the prevention of recurrent *clostridium difficile* infection: Results from punch CD 2, a randomized doubleblind placebo-controlled trial. *Gastroenterology*. 2017;152 (5 Supplement 1):S46-S7.
- Khanna S, Shin A, Kelly CP. Management of *Clostridium difficile* Infection in Inflammatory Bowel Disease: Expert Review from the Clinical Practice Updates Committee of the AGA Institute. *Clin Gastroenterol Hepatol*. 2017;15(2):166-74.
- Khanna S, Vazquez-Baeza Y, Gonzalez A, Weiss S, Schmidt B, Muniz-Pedrogo DA, et al. Changes in microbial ecology after fecal microbiota transplantation for recurrent *C. difficile* infection affected by underlying inflammatory bowel disease. *Microbiome*. 2017;5(1):55.
- Khoruts A, Staley C, Vaughn BP, Graiziger C, Sadowsky MJ. Treatment of urinary tract infections without affecting the gut microbiota in patients with recurrent *clostridium difficile* infection. *Gastroenterology*. 2016;1(PG-S689):S689.
- Konijeti G, Sauk J, Shrimé M, Ananthakrishnan A. Cost-effectiveness of competing strategies for recurrent *clostridium difficile* infection acg/astrazeneca fellow award. *Am J Gastroenterol*. 2013;108(PG-S473):S473.
- Konijeti GG, Sauk J, Shrimé MG, Gupta M, Ananthakrishnan AN. Cost-effectiveness of competing strategies for management of recurrent *clostridium difficile* infection: A decision analysis. *Clin Infect Dis*. 2014;58(11 PG-1507-1514):1507–14.
- Konturek PC, Dieterich W, Neurath M, Zopf Y. Successful therapy of *clostridium difficile* infection with fecal microbiota transplantation. *Gastroenterology*. 2017;152 (5 Supplement 1):S341.
- Kroner PT, Jirapinyo P, Abougergi MS, Thompson CC. *Clostridium difficile* regional and divisional incidence variations across the United States: a 10-year nationwide analysis. *Gastrointestinal Endoscopy*. 2017. (5 Supplement):AB246-AB247.
- Kucher MA, Goloschapov OV, Moiseev IS, Afanasyev BV. Fecal microbiota transplantation as a method to treat complications after hematopoietic stem cell transplantation. *Cellular Therapy and Transplantation*. 2017;6(1):20-9.
- Kump PK, Krause R, Steininger C, Grochenig HP, Moschen A, Madl C, et al. [Recommendations for the use of faecal microbiota transplantation “stool transplantation”: consensus of the Austrian Society of Gastroenterology and Hepatology (OGGH) in cooperation with the Austrian Society of Infectious Diseases and Tropical Medicine]. *Z Gastroenterol*. 2014;52(12 PG-1485-92):1485–92.
- Laffin M, Millan B, Madsen KL. Fecal microbial transplantation as a therapeutic option in patients colonized with antibiotic resistant organisms. *Gut Microbes*. 2017;8(3):221-4.

- Lagier JC, Aubry C, Delord M, Michelet P, Tissot-Dupont H, Million M, et al. From Expert Protocols to Standardized Management of Infectious Diseases. *Clin Infect Dis*. 2017;65:S12-S9.
- Landy J, Perry-woodford ZL, Clark SK, Hart A. Patients' perspectives of faecal transplantation for pouchitis. *J Crohn's Colitis*. 2012;6(PG-S143):S143.
- Lapointe-Shaw L, Tran KL, Coyte PC, Hancock-Howard RL, Powis J, Poutanen SM, et al. Cost-effectiveness analysis of six strategies to treat recurrent clostridium difficile infection. *PLoS One*. 2016;11 (2) (no pagination)(e0149521 PG-).
- Leber A, Hontecillas R, Abedi V, Tubau-Juni N, Zoccoli-Rodriguez V, Stewart C, et al. Modeling new immunoregulatory therapeutics as antimicrobial alternatives for treating Clostridium difficile infection. *Artif Intell Med*. 2017;78:1-13.
- Leber A, Viladomiu M, Hontecillas R, Abedi V, Philipson C, Hoops S, et al. Systems modeling of interactions between mucosal immunity and the gut microbiome during Clostridium difficile infection. *PLoS One*. 2015;10 (7) (no pagination)(e0134849 PG-).
- Lee C, Kim PT, Smith E. Outcome of fecal microbiota transplantation for recurrent clostridium difficile infection on quality of life. *Gastroenterology*. 2017;152 (5 Supplement 1):S949.
- Lee JC, Lee HY, Kim TK, Kim MS, Park YM, Kim J, et al. Obesogenic diet-induced gut barrier dysfunction and pathobiont expansion aggravate experimental colitis. *PLoS ONE*. 2017;12(11):e0187515.
- Lee STM, Kahn SA, Delmont TO, Shaiber A, Esen OC, Hubert NA, et al. Tracking microbial colonization in fecal microbiota transplantation experiments via genome-resolved metagenomics. *Microbiome*. 2017;5(1):50.
- Leigh DA, Simmons K. Effect of clindamycin and lincomycin therapy on faecal flora. *J Clin Pathol*. 1978;31(5 PG-439-443):439-43.
- Lenhart A, Mittal C, Zierle-Ghosh A, Alangaden G. Is colonization with non-toxigenic clostridium difficile organism protective against toxigenic strains? *Gastroenterology*. 2015;1(PG-S725):S725.
- Lewis BB, Pamer EG. Microbiota-Based Therapies for Clostridium difficile and Antibiotic-Resistant Enteric Infections. *Annual Review of Microbiology*. 2017;71:157-78.
- Lichtenstein GR. Fecal microbiota transplantation: An update. *Gastroenterology and Hepatology*. 2017;13(4):203.
- Lubbert C, Mutters R. [Gastrointestinal infections]. *Internist (Berl)*. 2017;58(2):149-69.
- Lubbert C, Salzberger B, Mossner J. Fecal microbiota transplantation. [German]. *Internist (Berl)*. 2017;58(5):456-68.
- Luo Y, Yang N, Roediger R, Ungaro RC, Grinspan A. Outcomes of fecal microbiota transplantation for clostridium difficile infections in inflammatory bowel disease patients. *Gastroenterology*. 2017;152 (5 Supplement 1):S342.

- Luong Nguyen LB, Osman M, Chiang AL, Edelstein C, Fischer M, Ananthakrishnan AN, et al. The cost-effectiveness of competing strategies for treating severe-complicated clostridium difficile infection: Comparing fecal microbiota transplantation with standard colectomy. *Gastroenterology*. 2016;1(PG-S543):S543.
- Ma GK, Brensinger CM, Wu Q, Lewis JD. Increasing Incidence of Multiply Recurrent Clostridium difficile Infection in the United States: A Cohort Study. *Ann Intern Med*. 2017;167(3):152-8.
- Ma GK, Brensinger CM, Wu Q, Lewis JD. Rising incidence of multiply-recurrent clostridium difficile infection in the United States. *Gastroenterology*. 2017;152 (5 Supplement 1):S340-S1
- Makkawi S, Metz L. Case report: Fecal microbiota transplantation associated with 10 years of disease stability in a patient with secondary progressive multiple sclerosis. *Multiple Sclerosis Journal*. 2017;23 (3 Supplement 1):517.
- Manthey CF, Eckmann L, Fuhrmann V. Therapy for Clostridium difficile infection - any news beyond Metronidazole and Vancomycin? *Expert Rev Clin Pharmacol*. 2017;10(11):1239-50.
- Marshall LL, Peasah S, Stevens GA. Clostridium difficile Infection in Older Adults: Systematic Review of Efforts to Reduce Occurrence and Improve Outcomes. *Consult Pharm*. 2017;32(1):24-41.
- Massachi S, Hay JW. Cost-effectiveness of various clostridium difficile infection (CDI) treatments in patients with recurrent infections. *Value Heal*. 2014;17 (3)(PG-A273-A274):A273-4.
- May T, Mackie RI, Fahey Jr GC, Cremin JC, Garleb KA. Effect of fiber source on short-chain fatty acid production and on the growth and toxin production by clostridium difficile. *Scand J Gastroenterol*. 1994;29(10 PG-916-922):916-22.
- Meinke KW, Hamedani F, Wu S, Balla A, Guzman G. Prototheca zopfii associated diverticulitis in an immunosuppressed host, a case presentation and literature review. *Human Pathology: Case Reports*. 2017;10:43-5.
- Merlo G, Graves N, Brain D, Connelly LB. Economic evaluation of fecal microbiota transplantation for the treatment of recurrent Clostridium difficile infection in Australia. *J Gastroenterol Hepatol*. 2016;31(12 PG-1927-1932):1927-32.
- Merlo G, Graves N, Connelly L. Economic evaluation of fecal microbiota transplantation for the treatment of recurrent clostridium difficile infection in Australia. *Value Heal*. 2015;18 (7)(PG-A628):A628.
- Mertz L. Omics tech, gut-on-a-chip, and bacterial engineering: New approaches for treating inflammatory bowel diseases. *IEEE Pulse*. 2016;7(5 PG-9-12):9-12.
- Metan G, Ture Z, Kaynar L, Berk E, Gursoy S, Alp E, et al. Tigecycline for the treatment of Clostridium difficile infection refractory to metronidazole in haematopoietic stem cell transplant recipients. *J Chemother*. 2015;27(6 PG-354-7):354-7.
- Millan B, Laffin M, Madsen K. Fecal Microbiota Transplantation: Beyond Clostridium difficile. *Current Infectious Disease Reports*. 2017;19 (9) (no pagination)(31).



- Mitchell DK, Van R, Mason EH, Norris DM, Pickering LK. Prospective study of toxigenic *Clostridium difficile* in children given amoxicillin/clavulanate for otitis media. *Pediatr Infect Dis J*. 1996;15(6 PG-514-519):514–9.
- Mitchell I, Shropshire K, Ruel J. *Clostridium difficile* infection and fecal bacteriotherapy. *Gastroenterol Nurs*. 2013;36(1 PG-42-50):42–50.
- Mitchell SW, DeZoysa P, Leis S, Jayewardene AF, Maistry P, Gadalla S, et al. Adverse effects of liquid vs. encapsulated lyophilized fullspectrum microbiota for the treatment of *clostridium difficile* infection. *Gastroenterology*. 2017;152 (5 Supplement 1):S346-S7.
- Mittal C, Hassan S, Abrencillo R, Bajjoka I, Abouljoud M, Patel A, et al. Changing trends of *clostridium difficile* associated diarrhea (CDAD) in liver transplant recipients (LTR) over 15 years. *Transplantation*. 2012;94(PG-549):549.
- Monaghan T, Negm O, MacKenzie B, Hamed M, Shone C, Humphreys DP, et al. High prevalence of subclass-specific binding and neutralising antibodies against *clostridium difficile* toxins in adult cystic fibrosis sera: Possible mode of protection against symptomatic *clostridium difficile* infection. *Gastroenterology*. 2017;152 (5 Supplement 1):S344.
- Moossavi S, Bishehsari F, Ansari R, Vahedi H, Nasser-Moghaddam S, Merat S, et al. Minimum requirements for reporting fecal microbiota transplant trial. *Middle East J Dig Dis*. 2015;7(3 PG-177-180):177–80.
- Morgan D. New antibiotics: What do we need? *Int J Infect Dis*. 2016;45(PG-51):51.
- Moss EL, Falconer SB, Tkachenko E, Wang M, Systrom H, Mahabamunuge J, et al. Long-term taxonomic and functional divergence from donor bacterial strains following fecal microbiota transplantation in immunocompromised patients. *PLoS ONE*. 2017;12(8):e0182585.
- Muenyi V, Kerman DH. Changes in the body mass index (BMI) of patients treated with fecal microbiota transplant (FMT) for recurrent *C. difficile* infection. *Gastroenterology*. 2017;152 (5 Supplement 1):S820-S1.
- Mullish BH, McDonald JA, Kao DH, Allegretti JR, Petrof EO, Pechlivanis A, et al. Understanding the mechanisms of efficacy of fecal microbiota transplantation in the treatment of *clostridium difficile* infection: The potential role of bilemetabolising enzymes. *Gastroenterology*. 2017;152 (5 Supplement 1):S47.
- Mullish BH, McDonald JAK, Thursz MR, Marchesi JR. Fecal microbiota transplant from a rational stool donor improves hepatic encephalopathy: A randomized clinical trial. *Hepatology*. 2017;66(4):1354-5.
- Mussatto CC, Wang J, Koon HW. Orally active cathelicidin mimic ceragenin CSA13 modulates *clostridium difficile*-associated colitis in mice via a modification of intestinal microbiome. *Gastroenterology*. 2017;152 (5 Supplement 1):S347-S8.
- Nanayakkara D, Nanda N. *Clostridium difficile* infection in solid organ transplant recipients. *Current Opinion in Organ Transplantation*. 2017;22(4):314-9.
- Narula N, Kassam Z, Yuan Y, Colombel JF, Ponsioen C, Reinisch W, et al. Systematic Review and Meta-analysis: Fecal Microbiota Transplantation for Treatment of Active Ulcerative Colitis. *Inflammatory Bowel Diseases*. 2017;23(10):1702-9.

- Newman KM, Rank K, Vaughn BP, Khoruts A. Treatment of recurrent clostridium difficile infection using fecal microbiota transplantation in patients with inflammatory bowel disease. *Gastroenterology*. 2017;152 (5 Supplement 1):S343.
- Ng SCC, Wong SH, Lui RN, Cheung K, Ching JYL, Tang W, et al. Vancomycin followed by fecal microbiota transplantation versus vancomycin for initial clostridium difficile infection: An open-label randomised controlled trial. *United European Gastroenterology Journal*. 2017;5 (5 Supplement 1):A314.
- Niccum BA, Stein DJ, Behm BW, Hays RA. Zinc deficiency predicts fecal microbiota transplant failure in recurrent clostridium difficile infection. *Gastroenterology*. 2017;152 (5 Supplement 1):S347.
- Nicholson M, Alexander E, Bartlett M, Becker P, Davidovics Z, Doby E, et al. Young faculty clinical investigator award fecal microbiota transplantation in pediatric clostridium difficile infection, a multi-center study. *Journal of Pediatric Gastroenterology and Nutrition*. 2017;65 (Supplement 2):S219-S20.
- Nozu R, Inoue T, Sato K, Hayashimoto N. Safety evaluation of fecal microbiota transplantation materials for Clostridium difficile infection in common marmosets. *Experimental Animals*. 2017;66:S68.
- Ong GK, Reidy TJ, Huk MD, Lane FR. Clostridium difficile colitis: A clinical review. *Am J Surg*. 2017;213(3):565-71.
- Oreiro MB, De La Guia AL, Nieto JB, De Paz R, Baltasar P, Hernandez D, et al. Fecal calprotectin in allogeneic stem cell transplantation as surrogate marker of gastrointestinal graft versus host disease. *Blood Conf 52nd Annu Meet Am Soc Hematol ASH*. 2010;116(21 PG-). Pestana L, Pardi D, Khanna S. Incidental colonoscopy findings during fecal microbiota transplantation for C. Difficile infection. *Gastroenterology*. 2016;1(PG-S747-S748):S747-8.
- Pamer EG. Microbiota-mediated defense against intestinal infection. *Annals of Hematology*. 2017;96:S43-S4.
- Panchal P, Budree S, Tu E, Kahn SA, Allegretti JR, Fischer M, et al. Pediatric access to fecal microbiota transplantation for recurrent clostridium difficile infection in the United States and the impact of stool banks: A geospatial analysis. *Gastroenterology*. 2017;152 (5 Supplement 1):S849-S50.
- Paramsothy S, Borody TJ, Lin E, Finlayson S, Walsh AJ, Samuel D, et al. Donor Recruitment for Fecal Microbiota Transplantation. *Inflamm Bowel Dis*. 2015;21(7 PG-1600-6):1600-6.
- Paramsothy S, Borody TJ, Lin E, Finlayson S, Walsh AJ, Samuel D, et al. Donor recruitment for fecal microbiota transplantation. *Inflamm Bowel Dis*. 2015;21(7 PG-1600-1606):1600-6.
- Paramsothy S, Walsh A, Borody T, Samuel D, Van Den Bogaerde J, Leong R, et al. Gastroenterologist perceptions of faecal microbiota transplantation. *J Gastroenterol Hepatol*. 2015;30(PG-21):21.
- Paramsothy S, Walsh AJ, Borody T, Samuel D, Van Den Bogaerde J, Leong RWL, et al. Gastroenterologist perceptions of faecal microbiota transplantation. *World J Gastroenterol*. 2015;21(38 PG-10907-10914):10907-14.



- Paramsothy S, Walsh AJ, Borody T, Samuel D, van den Bogaerde J, Leong RW, et al. Gastroenterologist perceptions of faecal microbiota transplantation. *World J Gastroenterol*. 2015;21(38 PG-10907-14):10907–14.
- Park HK, Millan B, Hotte N, Kao DH, Madsen K. Altered phage diversity and increased growth rate of *Escherichia coli* are associated with fecal transplantation failure in patients with *Clostridium difficile* infection. *Gastroenterology*. 2017;152 (5 Supplement 1):S191.
- Park L, Mone A, Price JC, Tzimas D, Hirsh J, Poles MA, et al. Perceptions of fecal microbiota transplantation for *Clostridium difficile* infection: Factors that predict acceptance. *Ann Gastroenterol*. 2017;30(1 PG-83-88):83–8.
- Pechine S, Janoir C, Collignon A. Emerging monoclonal antibodies against *Clostridium difficile* infection. *Expert Opin Biol Ther*. 2017;17(4):415-27.
- Perez E, Lee CH, Petrof EO. A Practical Method for Preparation of Fecal Microbiota Transplantation. *Methods Mol Biol*. 2016;1476(PG-259-67):259–67.
- Perez E, Lee CH, Petrof EO. A practical method for preparation of fecal microbiota transplantation. E-mail: humana@humanapr.com: Humana Press Inc.; 2016;(1476 PG-259-267):259–67.
- Piceno YM, El-Nachef N, Kassam Z, Smith M, Fadrosh D, Lynch K, et al. Fecal microbiota transplantation differentially influences the gut microbiota of *Clostridium difficile* infection and ileal pouch anal anastomosis patients. *Gastroenterology*. 2017;152 (5 Supplement 1):S1006.
- Pinn DM, Aroniadis OC, Brandt LJ. Is fecal microbiota transplantation (FMT) an effective treatment for patients with functional gastrointestinal disorders (FGID)? *Neurogastroenterol Motil*. 2015;27(1 PG-19-29):19–29.
- Pinn DM, Aroniadis OC, Brandt LJ. Is fecal microbiota transplantation the answer for irritable bowel syndrome? A single-center experience. *Am J Gastroenterol*. 2014;109(11 PG-1831-1832):1831–2.
- Plant BJ. *Clostridium Difficile* and other gut infections in patients with pulmonary disease, including in cystic fibrosis. *Pediatr Pulmonol*. 2015;50(PG-113-115):113–5.
- Prayitno N, Akhavan P, Khan M, Willey BM, Hota S, Sales V, et al. Determination of the optimal storage duration and conditions for faecal transplantation (FTX) samples. *Can J Infect Dis Med Microbiol*. 2012;23(PG-3B):3B.
- Prior AR, Kevans D, McDowell L, Cudmore S, Fitzpatrick F. Treatment of *Clostridium difficile* infection: A national survey of clinician recommendations and the use of faecal microbiota transplantation. *J Hosp Infect*. 2016;14(PG-).
- Prior AR, Kevans D, McDowell L, Cudmore S, Fitzpatrick F. Treatment of *Clostridium difficile* infection: a national survey of clinician recommendations and the use of faecal microbiota transplantation. *J Hosp Infect*. 2017;95(4):438-41.
- Quraishi MN, Segal J, Mullish B, McCune VL, Hawkey P, Colville A, et al. National survey of practice of faecal microbiota transplantation for *Clostridium difficile* infection in the UK. *Journal of Hospital Infection*. 2016.

- Quraishi MN, Segal J, Mullish B, McCune VL, Hawkey P, Colville A, et al. National survey of practice of faecal microbiota transplantation for *Clostridium difficile* infection in the UK. *J Hosp Infect*. 2017;95(4):444-5.
- Ray A, Jones CR, Shannon B, Carter S. Donors are universal in the fight against *clostridium difficile*: Results from two trials investigating the safety and efficacy of RBX2660, a microbiota-based drug. *Gastroenterology*. 2017;152 (5 Supplement 1):S950.
- Ren RR, Sun G, Yang YS, Peng LH, Wang SF, Shi XH, et al. Chinese physicians' perceptions of fecal microbiota Transplantation. *World J Gastroenterol*. 2016;22(19 PG-4757-4765):4757-65.
- Rice LB. The complex dynamics of antimicrobial activity in the human gastrointestinal tract. *Trans Am Clin Climatol Assoc*. 2013;124(PG-123-132):123-32.
- Richardson C, Kim P, Lee C, Bersenas A, Weese JS. Comparison of *Clostridium difficile* isolates from individuals with recurrent and single episode of infection. *Anaerobe*. 2015;33(PG-105-108):105-8.
- Saffouri G, Khanna S, Pardi D. Outcomes from rectal vancomycin therapy in patients with severe-complicated *clostridium difficile* infection. *Am J Gastroenterol*. 2013;108(PG-S175):S175.
- Saffouri G, Pardi D, Kashyap P, Khanna S. Body mass index changes after fecal microbiota transplant for recurrent *clostridium difficile* infection. *Am J Gastroenterol*. 2016;111(PG-S103):S103.
- Sammons JS, Gerber JS, Tamma PD, Sandora TJ, Beekmann SE, Polgreen PM, et al. Diagnosis and management of *Clostridium difficile* infection by pediatric infectious diseases physicians. *J Pediatric Infect Dis Soc*. 2014;3(1 PG-43-48):43-8.
- Samuel BP, Crumb TL, Duba MM. What nurses need to know about fecal microbiota transplantation: education, assessment, and care for children and young adults. *J Pediatr Nurs*. 2014;29(4 PG-354-361):354-61.
- Schvartz B, Leveque N. Asymptomatic carriage of gastro-intestinal pathogens in renaltransplant recipients : Epidemiology and risk factors. *Nephrol Dial Transplant*. 2015;30(PG-iii356-iii357):iii356-iii357.
- Sears P, Crook DW, Louie TJ, Miller MA, Weiss K. Fidaxomicin attains high fecal concentrations with minimal plasma concentrations following oral administration in patients with *clostridium difficile* infection. *Clin Infect Dis*. 2012;55(SUPPL.2 PG-S116-S120):S116-20.
- Seril DN, Shen B. *Clostridium difficile* infection in the postcolectomy patient. *Inflamm Bowel Dis*. United States; 2014;20(12):2450-69.
- Shaughnessy MK, Bobr A, Kuskowski MA, Johnston BD, Sadowsky MJ, Khoruts A, et al. Environmental Contamination in Households of Patients with Recurrent *Clostridium difficile* Infection. *Appl Environ Microbiol*. 2016;82(9 PG-2686-92):2686-92.
- Sheehan D, Brown J, Flemer B, Zulquernain SA, Gahan CG, Joyce S, et al. Mechansims underpinning successful faecal microbiota transplantation (FMT) for recurrent *clostridium difficile* infection. *Gastroenterology*. 2017;152 (5 Supplement 1):S47-S8.
- Shen NT, Gold SL, Schneider Y, Cohen-Mekelburg SA, Maw AM, Crawford CV. Probiotic sepsis in patients with

- inflammatory bowel disease; Is it something to worry about? *Gastroenterology*. 2017;152 (5 Supplement 1):S817.
- Sidhu M, van der Poorten D. The gut microbiome. *Aust Fam Physician*. 2017;46(4):206-11.
- Simmerlein R, Basta A, Gosch M. [Clostridium difficile infections in geriatric patients]. *Z Gerontol Geriatr*. 2016;49(8 PG-743-761):743-61.
- Simojoki ST, Kirjavainen V, Rahiala J, Kanerva J. Surveillance cultures in pediatric allogeneic hematopoietic stem cell transplantation. *Pediatr Transplant*. 2014;18(1 PG-87-93):87-93.
- Smith AD, Zhang IT, Schubert AM, Giordano NP, Hastie JE, Cowley SC, et al. MAIT cells: Shaping the microbiome, contributing to Clostridium difficile infection. *Journal of Immunology Conference: 104<sup>th</sup> Annual Meeting of the American Association of Immunologists, AAI*. 2017;198(1 Supplement 1).
- Smith JD, Roach B, Silva M, Louie T, Xu H, Kao DH. Donor body mass index (BMI) does not impact recipient BMI following fecal microbiota transplantation for recurrent clostridium difficile infection. *Gastroenterology*. 2017;152 (5 Supplement 1):S1007.
- Smith MB, Kassam Z, Burgess J, Perrotta AR, Burns LJ, Mendolia GM, et al. The international public stool bank: A scalable model for standardized screening and processing of donor stool for fecal microbiota transplantation. *Gastroenterology*. 2015;1(PG-S211):S211.
- Smith MB, Kelly C, Alm EJ. Policy: How to regulate faecal transplants. *Nature*. 2014;506(7488):290-1.
- Sofi A, Georgescu C, Sodeman T, Nawras A. Physician outlook towards fecal microbiota transplantation in the treatment of recurrent clostridium difficile infection. *Gastroenterology*. 2013;1(PG-S241):S241.
- Sofi A, Nawras A, Sodeman T, Garborg K, Silverman A. Fecal bacteriotherapy works for clostridium difficile infection - A meta-analysis. *Am J Gastroenterol*. 2011;106(PG-S161):S161.
- Sofi AA, Georgescu C, Sodeman T, Nawras A. Physician outlook toward fecal microbiota transplantation in the treatment of Clostridium difficile infection. *Am J Gastroenterol*. 2013;108(10 PG-1661-1662):1661-2.
- Sofi AA, Silverman AL, Khuder S, Garborg K, Westerink JM, Nawras A. Relationship of symptom duration and fecal bacteriotherapy in Clostridium difficile infection-pooled data analysis and a systematic review. *Scand J Gastroenterol*. 2013;48(3 PG-266-73):266-73.
- Solbach P, Dersch P, Bachmann O. Individualized treatment strategies for Clostridium difficile infections. [German]. *Internist (Berl)*. 2017;58(7):675-81.
- Song Y, Garg S, Girotra M, Maddox C, von Rosenvinge EC, Dutta A, et al. Microbiota dynamics in patients treated with fecal microbiota transplantation for recurrent Clostridium difficile infection.[Erratum appears in PLoS One. 2014;9(7):e104471]. *PLoS ONE [Electronic Resour]*. 2013;8(11 PG-e81330):e81330.
- Spalinger M, Gottier C, Hering L, Lang S, Rogler G, Scharl MM. Co-housing DSS treated mice with healthy mice results in faster normalization of the intestinal microbiota and promotes recovery. *Gastroenterology*. 2017;152 (5 Supplement 1):S987.

- Spiceland CM, Saffouri G, Pardi D, Khanna S. Outcomes of fidaxomicin treatment of clostridium difficile infection. *Gastroenterology*. 2016;1(PG-S744):S744.
- Srinivasan I, Tang SJ, Sones JQ. Fecal microbial transplantation. *Gastrointestinal Endoscopy*. 2017;85(5):1107-8.
- Staley C, Hamilton MJ, Vaughn BP, Graiziger C, Newman KM, Kabage A, et al. Successful resolution of recurrent clostridium difficile infection using freeze-dried, encapsulated fecal microbiota. *Gastroenterology*. 2017;152 (5 Supplement 1):S343-S4.
- Staley C, Kelly CR, Brandt LJ, Khoruts A, Sadowsky MJ. Complete microbiota engraftment is not essential for recovery from recurrent *Clostridium difficile* infection following fecal microbiota transplantation. *MBio*. 2016;7 (6) (no pagination)(e01965-16 PG-).
- Staley C, Khoruts A, Sadowsky MJ. Contemporary Applications of Fecal Microbiota Transplantation to Treat Intestinal Diseases in Humans. *Archives of Medical Research*. 2017.
- Staley C, Vaughn BP, Graiziger CT, Sadowsky MJ, Khoruts A. Gut-sparing treatment of urinary tract infection in patients at high risk of *Clostridium difficile* infection. *J Antimicrob Chemother*. 2016;20(PG-20):20.
- Staley C, Vaughn BP, Graiziger CT, Sadowsky MJ, Khoruts A. Gut-sparing treatment of urinary tract infection in patients at high risk of *Clostridium difficile* infection. *J Antimicrob Chemother*. 2017;72(2):522-8.
- Staley C, Vaughn BP, Graiziger CT, Singroy S, Hamilton MJ, Yao D, et al. Community dynamics drive punctuated engraftment of the fecal microbiome following transplantation using freeze-dried, encapsulated fecal microbiota. *Gut Microbes*. 2017;8(3):276-88.
- Steevens CD, Roto D, DeCross AJ. Obese stool donors in fecal microbiota transplantation: Not associated with recipient weight gain! *Gastroenterology*. 2017;152 (5 Supplement 1):S1007-S8.
- Surawicz CM, Brandt LJ, Binion DG, Ananthakrishnan AN, Curry SR, Gilligan PH, et al. Guidelines for diagnosis, treatment, and prevention of *Clostridium difficile* infections. *Am J Gastroenterol*. 2013;108(4):478-98; quiz 499.
- Tan M, Smitasin N, Ong D, Lee AJ, Jureen R, Tambyah P, et al. Bacteraemia following faecal microbiota transplantation for recurrent clostridium difficile infection in an immunosuppressed patient. *Antimicrobial Resistance and Infection Control Conference: International Conference on Prevention and Infection Control, ICPIC*. 2017;6(Supplement 3).
- Tariq R, Khanna S. *Clostridium difficile* infection: Updates in management. *Indian J Gastroenterol*. 2017;36(1):3-10.
- Tariq R, Pardi DS, Weatherly RM, Kammer PP, Khanna S. Outcomes and management of patients with failed fecal microbiota transplantation for recurrent clostridium difficile infection. *Gastroenterology*. 2017;152 (5 Supplement 1):S346.
- Tariq R, Weatherly R, Kammer P, Pardi D, Khanna S. Donor screening experience for fecal microbiota transplantation for patients with recurrent *C. Difficile* infection. *Am J Gastroenterol*. 2016;111(PG-S458):S458.

- Tariq R, Weatherly R, Kammer P, Pardi DS, Khanna S. Donor Screening Experience for Fecal Microbiota Transplantation in Patients With Recurrent *C. difficile* Infection. *J Clin Gastroenterol*. 2016;14(PG-).
- Tariq R, Weatherly RM, Kammer PP, Walker RC, Razonable RR, Pardi DS, et al. Improved urinary tract infections with fecal microbiota transplantation for recurrent *clostridium difficile* infection. *Gastroenterology*. 2017;152 (5 Supplement 1):S815.
- Taur Y, Jenq RR, Perales MA, Littmann ER, Morjaria S, Ling L, et al. The effects of intestinal tract bacterial diversity on mortality following allogeneic hematopoietic stem cell transplantation. *Blood*. 2014;124(7 PG-1174-1182):1174–82.
- Tauxe WM, Dhere T, Ward A, Racska LD, Varkey JB, Kraft CS. Fecal microbiota transplant protocol for *Clostridium Difficile* infection. *Lab Med*. 2015;46(1 PG-e19-e23):e19–23.
- Taylor KN, McHale MT, Saenz CC, Plaxe SC. Diagnosis and treatment of *Clostridium difficile* (*C. diff*) colitis: Review of the literature and a perspective in gynecologic oncology. *Gynecol Oncol*. 2016;23(PG-).
- Taylor KN, McHale MT, Saenz CC, Plaxe SC. Diagnosis and treatment of *Clostridium difficile* (*C. diff*) colitis: Review of the literature and a perspective in gynecologic oncology. *Gynecol Oncol*. 2017;144(2):428-37.
- Thaiss CA, Elinav E. The remedy within: will the microbiome fulfill its therapeutic promise? *Journal of Molecular Medicine*. 2017;95(10):1021-7.
- Tissot F, Maillard MH. [*Clostridium difficile* infections: update on new European recommendations]. *Rev Med Suisse*. 2014;10(427 PG-913-6, 918-9):913–916,918.
- To KB, Napolitano LM. *Clostridium difficile* infection: Update on diagnosis, epidemiology, and treatment strategies. *Surg Infect (Larchmt)*. 2014;15(5 PG-490-502):490–502.
- Trinh SA, Echenique IA, Penugonda S, Angarone MP. Optimal strategies for the diagnosis of community-onset diarrhea in solid organ transplant recipients: Less is more. *Transpl Infect Dis*. 2017;19 (2) (no pagination)(e12673).
- Tschudin-Sutter S, Widmer AF, Perl TM. *Clostridium difficile*: Novel insights on an incessantly challenging disease. *Curr Opin Infect Dis*. 2012;25(4 PG-405-411):405–11.
- Ulmer L, Verma A, Brock J, Iyer R. Fecal microbiota transplant for *C. difficile* colitis from thawed frozen stool and "real world" experience in a community hospital over two years. *Gastroenterology*. 2017;152 (5 Supplement 1):S341.
- Van den Abbeele P, Verstraete W, El Aidy S, Geirnaert A, Van de Wiele T. Prebiotics, faecal transplants and microbial network units to stimulate biodiversity of the human gut microbiome. *Microb Biotechnol*. 2013;6(4 PG-335-340):335–40.
- Varier RU, Biltaji E, Smith KJ, Roberts MS, Jensen MK, LaFleur J, et al. Cost-effectiveness analysis of treatment strategies for initial *Clostridium difficile* infection. *Clin Microbiol Infect*. 2014;20(12 PG-1343-1351):1343–51.

- Varier RU, Biltaji E, Smith KJ, Roberts MS, Jensen MK, LaFleur J, et al. Cost-Effectiveness Analysis of Fecal Microbiota Transplantation for Recurrent *Clostridium difficile* Infection. *Infect Control Hosp Epidemiol*. 2015;36(4 PG-438-444):438–44.
- Varier RU, Biltaji E, Smith KJ, Roberts MS, Kyle Jensen M, LaFleur J, et al. Cost-effectiveness analysis of fecal microbiota transplantation for recurrent *Clostridium difficile* infection. *Infect Control Hosp Epidemiol*. 2015;36(4 PG-438-444):438–44.
- Varier RU, Biltaji EO, Smith KJ, Roberts MS, LaFleur J, Nelson RE. Cost-effectiveness analysis of fecal microbiota transplantation versus vancomycin for recurrent *clostridium difficile* infection. *Gastroenterology*. 2014;1(PG-S250-S251):S250–1.
- Vaughn B, Kahn S, Rubin D, Moss A. Donor stool preparation for fecal transplantation in patients with IBD: Regulatory and financial aspects. *Inflamm Bowel Dis*. 2013;19(PG-S88):S88.
- Vemuri RC, Gundamaraju R, Shinde T, Eri R. Therapeutic interventions for gut dysbiosis and related disorders in the elderly: Antibiotics, probiotics or faecal microbiota transplantation? *Beneficial Microbes*. 2017;8(2):179-92.
- Verna EC, Macesic N, Annavajhala M, Giddins M, Stump S, Brown RS, et al. Dynamic adaptations of intestinal microbiota after liver transplantation. *Hepatology*. 2017;66 (Supplement 1):116A.
- Vestermarck CA, Singla MB, Rodriguez B, Armbruster SP. Salmonella-associated *clostridium difficile* infection presenting as new onset ascites. *Am J Gastroenterol*. 2016;111(PG-S617):S617.
- Vyas D, Aekka A, Vyas A. Fecal transplant policy and legislation. *World J Gastroenterol*. 2015;21(1 PG-6-11):6–11.
- Waye A, Atkins K, Kao D. Cost averted with timely fecal microbiota transplantation in the management of recurrent *clostridium difficile* infection in Alberta, Canada. *J Clin Gastroenterol*. 2016;50(9 PG-747-753):747–53.
- Weil AA, Hohmann EL. Fecal microbiota transplant: Benefits and risks. *Open Forum Infect Dis*. 2015;2 (1) (no pagination)(ofv005 PG-).
- Weingarden AR, Vaughn BP. Intestinal microbiota, fecal microbiota transplantation, and inflammatory bowel disease. *Gut Microbes*. 2017;8(3):238-52.
- Wilson D, Rahni D, Kelly C. Safety outcomes after fecal microbiota transplantation (FMT) For *C. Difficile* Infection (CDI). *Am J Gastroenterol*. 2014;109(PG-S207):S207.
- Wilson M, Ritz N, Singh S, Lin HC. Successful fecal microbiota transplant depends on the gut microbial community of the recipient to be already perturbed. *Gastroenterology*. 2017;152 (5 Supplement 1):S1039.
- Wolf-Meyer MJ. Normal, Regular and Standard: Scaling the Body through Fecal Microbial Transplants. *Med Anthropol Q*. 2016;30(PG-30):30.
- Wolf-Meyer MJ. Normal, Regular, and Standard: Scaling the Body through Fecal Microbial Transplants. *Med Anthropol Q*. 2017;31(3):297-314.



- Wurm P, Spindelboeck W, Krause R, Plank J, Fuchs G, Bashir M, et al. Antibiotic-Associated Apoptotic Enterocolitis in the Absence of a Defined Pathogen: The Role of Intestinal Microbiota Depletion. *Crit Care Med*. 2017;45(6):e600-e6.
- Xi D, Michail S. Fecal microbiota transplantation in children does not significantly alter body mass index. *Gastroenterology*. 2017;152 (5 Supplement 1):S648.
- Yakob L, Riley T V, Paterson DL, Marquess J, Clements AC. Assessing control bundles for *Clostridium difficile*: a review and mathematical model. *Emerg Microbes Infect*. 2014;3(6 PG-e43):e43.
- Yakob L, Riley T V, Paterson DL, Marquess J, Clements ACA. Assessing control bundles for *Clostridium difficile*: A review and mathematical model. *Emerg Microbes Infect*. 2014;3 (no pagination)(e43 PG-).
- Yamazaki Y, Kawarai S, Morita H, Kikusui T, Iriki A. Faecal transplantation for the treatment of *Clostridium difficile* infection in a marmoset. *BMC Vet Res*. 2017;13(1):150.
- Yang Z, Wang X, Bu C. Fecal microbiota transplant for Crohn's disease: A prospective, randomized study in chinese population. *United European Gastroenterology Journal*. 2017;5 (5 Supplement 1):A112-A3.
- Yeh A, Morowitz MJ. Probiotics and fecal microbiota transplantation in surgical disorders. *Seminars in Colon and Rectal Surgery*. 2017.
- Young VB. Treatment With Fecal Microbiota Transplantation: The Need for Complete Methodological Reporting for Clinical Trials. *Ann Intern Med*. 2017;167(1):61-2.
- Zeitz J, Bissig M, Barthel C, Biedermann L, Scharl S, Pohl D, et al. Patients' views on fecal microbiota transplantation: an acceptable therapeutic option in inflammatory bowel disease? *Eur J Gastroenterol Hepatol*. 2016;22(PG-).
- Zeitz J, Bissig M, Barthel C, Biedermann L, Scharl S, Pohl D, et al. Patients' views on fecal microbiota transplantation: an acceptable therapeutic option in inflammatory bowel disease? *Eur J Gastroenterol Hepatol*. 2017;29(3):322-30.
- Zellmer C, De Wolfe TJ, Van Hoof S, Blakney R, Safdar N. Patient Perspectives on Fecal Microbiota Transplantation for *Clostridium Difficile* Infection. *Infect Dis Ther*. 2016;5(2 PG-155-164):155-64.
- Zhang F, Luo W, Shi Y, Fan Z, Ji G. Should we standardize the 1,700-year-old fecal microbiota transplantation. *Am J Gastroenterol*. 2012;107(11 PG-1755):1755.
- Zhu YL, Guo XH, Zhang LF, Qin YM. A case of fecal microbiota transplantation for treatment of ulcerative colitis. [Chinese]. *World Chinese Journal of Digestology*. 2017;25(14):1321-6.
- Zhu YM, Li L. New recognition of gut microbiota and related diseases. [Chinese]. *World Chinese Journal of Digestology*. 2017;25(23):2095-101.
- Zipursky JS, Sidorsky TI, Freedman C a., Sidorsky MN, Kirkland KB. Patient attitudes toward the use of fecal microbiota transplantation in the treatment of recurrent *Clostridium difficile* infection. *Clin Infect Dis*. 2012;55(12):1652-8.

Zipursky JS, Sidorsky TI, Freedman CA, Sidorsky MN, Kirkland KB. Physician attitudes toward the use of fecal microbiota transplantation for the treatment of recurrent *Clostridium difficile* infection. *Can J Gastroenterol Hepatol*. 2014;28(6 PG-319-324):319–24.

Zowall H, Brewer C, Deutsch A. A model of *clostridium difficile* infection: Dynamic transmission between hospitals , long-term care facilities and communities. *Value Heal*. 2014;17 (3)(PG-A280-A281):A280–1.

Zowall H, Brewer C, Deutsch A. Cost-effectiveness of fecal microbiota transplant in treating *clostridium difficile* infection in Canada. *Value Heal*. 2014;17 (7)(PG-A676):A676.

Zowall H, Brewer C, Deutsch A. Projected cost savings of introducing fecal microbiota transplant treatment for *clostridium difficile* infection in Canada. *Value Heal*. 2015;18 (3)(PG-A238):A238.

Zucca M, Scutera S, Savoia D. Novel avenues for *Clostridium difficile* infection drug discovery. *Expert Opin Drug Discov*. 2013;8(4 PG-459-477):459–77.

Zuo T, Wong SH, Lam K, Lui R, Cheung K, Tang W, et al. Bacteriophage transfer during faecal microbiota transplantation in *Clostridium difficile* infection is associated with treatment outcome. *Gut*. 2017;15.

## **D.2. Non-CDI indications:D.2.1. Abstracts not fulfilling selection criteria:**

Bajaj JS, Kassam Z, Fagan A, Gavis E, Liu EJ, Kheradman R, et al. Fecal microbiota transplant using a precision medicine approach is safe, associated with lower hospitalization risk and improved cognitive function in recurrent hepatic encephalopathy. *Gastroenterology*. 2017;152 (5 Supplement 1):S906.

El-Nachef N, Piceno YM, Kassam Z, Zydek M, Ablaza AJ, Leith T, et al. Fecal microbiota transplantation is safe and effective in chronic pouchitis patients. *Gastroenterology*. 2017;152 (5 Supplement 1):S1009.

Hefazi M, Patnaik MM, Hogan WJ, Litzow MR, Pardi DS, Khanna S. Safety and efficacy of fecal microbiota transplantation for recurrent *clostridium* infection in patients with hematologic malignancies. *Blood Conference: 58th Annual Meeting of the American Society of Hematology, ASH*. 2016;128(22).

Ianiro G, Masucci L, Valerio L, Nagel D, D'Aversa F, Poto R, et al. Fecal microbiota transplantation for recurrent *C. Difficile* infection: Analysis of factors associated with the need for multiple fecal infusions. *United European Gastroenterology Journal*. 2016;4 (5 Supplement 1):A96-A7.

Karakan T, Ibis M, Cindoruk M, Sargin ZG, Alizadeh N. Faecal microbiota transplantation as a rescue therapy for steroid-dependent and/or nonresponsive patients with ulcerative colitis: A pilot study. *Journal of Crohn's and Colitis*. 2016;10:S425-S6.

Masaoka T, Yamane T, Mizuno S, Mori K, Hirata K, Matsushita M, et al. Safety and efficacy of fecal microbiota transplantation on functional bowel disorders-a pilot study. *Neurogastroenterology and Motility*. 2016;28 (Supplement 1):96.

Paramsothy S, Kaakoush N, Kamm MA, Faith J, Clemente J, Walsh A, et al. Faecal microbiota transplantation (FMT) in ulcerative colitis (UC) is associated with specific bacterial changes: Stool and colonic mucosa 16s microbiota analysis from the randomised controlled focus study. *United European Gastroenterology Journal*.



2016;4 (5 Supplement 1):A30-A1.

Paramsothy S, Kamm M, Walsh A, Van Den Bogaerde J, Samuel D, Leong R, et al. Multi-donor intense faecal microbiota transplantation is an effective treatment for resistant ulcerative colitis: A randomised placebo-controlled trial. *Journal of Crohn's and Colitis*. 2016;10:S14.

Paramsothy S, Kamm MA, Walsh AJ, Van Den Bogaerde J, Samuel D, Leong RWL, et al. Multi donor intense faecal microbiota transplantation is an effective treatment for resistant ulcerative colitis: A randomised placebocontrolled trial and microbiota analysis. *Journal of Gastroenterology and Hepatology (Australia)*. 2016;31:143.

Rossen N, Fuentes S, Van Der Spek M, Tijssen J, Hartman J, Duflo A, et al. Faecal microbiota transplantation in Ulcerative Colitis: A randomised controlled trial. *Journal of Crohn's and Colitis*. 2015;9:S2.

#### **D.2.2. Case series not fulfilling selection criteria:**

Cui B, Feng Q, Wang H, Wang M, Peng Z, Li P, et al. Fecal microbiota transplantation through mid-gut for refractory Crohn's disease: Safety, feasibility, and efficacy trial results. *Journal of Gastroenterology and Hepatology (Australia)*. 2015;30(1):51-8.

Cui B, Li P, Xu L, Zhao Y, Wang H, Peng Z, et al. Step-up fecal microbiota transplantation strategy: A pilot study for steroid-dependent ulcerative colitis. *Journal of Translational Medicine*. 2015;13 (1) (no pagination)(298).

David B, Batista R, Michelon H, Lepointeur M, Bouchand F, Lepeule R, et al. Is faecal microbiota transplantation an option to eradicate highly drug-resistant enteric bacteria carriage? *Journal of Hospital Infection*. 2017;95(4):433-7.

Fang YH, Chen J, Yu JD, Luo YY, Lou JG. The preliminary investigation of faecal microbiota transplantation for paediatric recurrent chronic bowel diseases and literature review. *Hong Kong Journal of Paediatrics*. 2017;22(4):199-203.

Grewal CS, Sood A, Mehta V, Mahajan R. Role of fecal microbiota transplantation in steroid dependant ulcerative colitis: A prospective observational study. *Indian Journal of Gastroenterology*. 2016;35 (1 Supplement):A39.

Grewal CS, Sood A, Mehta V, Sood N, Midha V, Mahajan R, et al. Role of fecal microbiota transplantation in patients with steroid dependant ulcerative colitis. *American Journal of Gastroenterology*. 2016;111:S1252-S3.

Ishikawa D, Osada T, Haga K, Kodani T, Shibuya T, Watanabe S. Combination therapy of fresh faecal microbial transplantation and antibiotics for ulcerative colitis. *Journal of Crohn's and Colitis*. 2016;10:S335-S6.

Jacob V, Crawford C, Cohen-Mekelburg S, Viladomiu M, Putzel GG, Schneider Y, et al. Single Delivery of High-Diversity Fecal Microbiota Preparation by Colonoscopy Is Safe and Effective in Increasing Microbial Diversity in Active Ulcerative Colitis. *Inflammatory Bowel Diseases*. 2017;23(6):903-11.

Karakan T, Ibis M, Gok Sargn Z. Faecal microbiota transplantation (FMT) as a rescue therapy for steroid-dependent and/or nonresponsive patients with ulcerative colitis (UC): A pilot study. *United European Gastroenterology Journal*. 2016;4 (5 Supplement 1):A268.

Kunde S, Pham A, Bonczyk S, Crumb T, Duba M, Conrad H, et al. Safety, tolerability, and clinical response after fecal transplantation in children and young adults with ulcerative colitis. *Journal of Pediatric Gastroenterology and Nutrition*. 2013;56(6):597-601.

Midha V, Singh A, Grewal CS, Mahajan R, Mehta V, Sood A. Efficacy and safety of fecal microbiota therapy in ulcerative colitis: Early experience. *Journal of Gastroenterology and Hepatology*. 2017;32 (Supplement 3):150-1.

Nishida A, Imaeda H, Ohno M, Inatomi O, Bamba S, Sugimoto M, et al. Efficacy and safety of single fecal microbiota transplantation for Japanese patients with mild to moderately active ulcerative colitis. *Journal of Gastroenterology*. 2017;52(4):476-82.

Suskind DL, Singh N, Nielson H, Wahbeh G. Fecal microbial transplant via nasogastric tube for active pediatric ulcerative colitis. *Journal of Pediatric Gastroenterology and Nutrition*. 2015;60(1):27-9.

Uygun A, Ozturk K, Demirci H, Oger C, Avci IY, Turker T, et al. Fecal microbiota transplantation is a rescue treatment modality for refractory ulcerative colitis. *Medicine (United States)*. 2017;96 (16) (no pagination)(e6479).

Wei Y, Gong J, Zhu W, Tian H, Ding C, Gu L, et al. Pectin enhances the effect of fecal microbiota transplantation in ulcerative colitis by delaying the loss of diversity of gut flora. *BMC Microbiology*. 2016;16(1):1-9.

Wei Y, Zhu W, Gong J, Guo D, Gu L, Li N, et al. Fecal microbiota transplantation improves the quality of life in patients with inflammatory bowel disease. *Gastroenterology Research and Practice*. 2015;2015 (no pagination)(517597).

### **D.2.3. Narrative reviews:**

Biehl L. Fecal microbiota transfer. *Transfusion Medicine and Hemotherapy*. 2017;44 (Supplement 1):22.

Borody TJ, Warren EF, Leis SM, Surace R, Ashman O, Siarakas S. Bacteriotherapy using fecal flora: Toying with human motions. *Journal of Clinical Gastroenterology*. 2004;38(6):475-83.

Colman RJ, Rubin DT. Fecal microbiota transplantation as therapy for inflammatory bowel disease: A systematic review and meta-analysis. *Journal of Crohn's and Colitis*. 2014;8(12):1569-81.

### **D.2.4. Miscellaneous, not relevant:**

Aarbiou J, Leeming DJ, Cruwys S, Gudmann NS, Brockbank S, Young A, et al. A comparison of compounds with claimed anti-fibrotic activity in a novel human fibroblast to myofibroblast transition assay using IPF derived patient material. *American Journal of Respiratory and Critical Care Medicine Conference: American Thoracic Society International Conference, ATS*. 2017;195(no pagination).

- Abbas SH, Abdulridha MK, Najeb AA. Potential benefit of curcumin adjuvant therapy to the standard *Helicobacter pylori* eradication therapy in patients with peptic ulcer disease. *Asian Journal of Pharmaceutical and Clinical Research*. 2017;10(5):313-7.
- Adachi JA, DuPont HL. Rifaximin: A novel nonabsorbed rifamycin for gastrointestinal disorders. *Clinical Infectious Diseases*. 2006;42(4):541-7.
- Adam B, Koldehoff M, Ditschkowski M, Gromke T, Hlinka M, Trenchel R, et al. Endoscopic and Histological Findings Are Predicted by Fecal Calprotectin in Acute Intestinal Graft-Versus-Host-Disease. *Digestive Diseases and Sciences*. 2016;61(7):2019-26.
- Agachan F, Pfeifer J, Joo JS, Nogueras JJ, Weiss EG, Wexner SD. Results of perineal procedures for the treatment of rectal prolapse. *American Surgeon*. 1997;63(1):9-12.
- Ahmed AR, Watanabe H, Aoki J, Shinozaki T, Takagishi K. Schwannoma of the extremities: The role of PET in preoperative planning. *European Journal of Nuclear Medicine*. 2001;28(10):1541-51.
- Al-Bayati I, Saadi M, Elhanafi S, McCallum RW. Effectiveness of Bulking Agent (Solesta) Therapy in Fecal Incontinence in Patients Refractory to Conventional Therapies. *American Journal of the Medical Sciences*. 2017.
- Almeida AG, Mesquita Gabriel H, Coutinho CA, Sargento L, David C, Oliveira J, et al. Myocardial perfusion and angioplasty. Comparison of myocardial contrast echocardiography and scintigraphy. [Portuguese, English]. *Revista Portuguesa de Cardiologia*. 2002;21(7-8):859-68.
- Amini M, Khedmat H, Yari F. Eradication rate of *Helicobacter pylori* in dyspeptic patients. *Medical Science Monitor*. 2005;11(4):CR193-CR5.
- Amini-Bavil-Olyaei S, Trautwein C, Tacke F. Hepatitis E vaccine: Current status and future prospects. *Future Virology*. 2009;4(2):143-54.
- Andersen ML, Fallentin E, Lauridsen CA, Kjaer MS, Clemmesen O, Larsen FS, et al. Evaluation of blood perfusion in liver cirrhosis by dynamic contrast enhanced computed tomography. *Hepatology*. 2017;66 (Supplement 1):343A.
- Anonymous. Abstracts of the 10th Congress of ECCO. *Journal of Crohn's and Colitis Conference: 10th Congress of the European Crohn's and Colitis Organisation, ECCO*. 2015;9(no pagination).
- Arguedas MR, Fallon MB. Hepatitis A. *Current Treatment Options in Gastroenterology*. 2004;7(6):443-50.
- Arnold PM, Carandang GC, Zabner R, Irwin ME. Randomized controlled trial of selective bowel decontamination for prevention of infections following liver transplantation. *Clinical Infectious Diseases*. 1996;22(6):997-1003.
- Asari SO, Nakajima T, Kojima K, Miyauchi A, Saitou JI, Saga Y, et al. FMT-PET analysis in gene therapy for AADC deficiency. *Clinical Neurology*. 2016;56:S268.

- Ashraf W, Park F, Lof J, Quigley EM. Effects of psyllium therapy on stool characteristics, colon transit and anorectal function in chronic idiopathic constipation. *Aliment Pharmacol Ther.* 1995;9(6):639-47.
- Badin RA, Binley K, Van Camp N, Jan C, Gourlay J, Stewart H, et al. Advancing a state of the art gene therapy for parkinson's disease. *Molecular Therapy.* 2015;23:S79-S80.
- Bajaj JS, Kassam Z, Fagan A, Gavis EA, John B, Fuchs M, et al. Fecal microbiota transplantation from a rationally selected donor is safe in patients with recurrent hepatic encephalopathy: Preliminary data from a randomized trial. *Hepatology.* 2016;64 (1 Supplement 1):717A.
- Bajaj JS, Sikaroodi M, White M, Fagan A, Gilles HC, Heuman DM, et al. Liver transplant significantly improves gut microbial dysbiosis and microbial diversity in cirrhotic patients. *Hepatology.* 2016;64 (1 Supplement 1):492A-3A.
- Bariol C, Meagher AP, Vickers CR, Byrnes DJ, Edwards PD, Hing M, et al. Thalidomide for inflammatory bowel disease: Early studies on the safety and efficacy of thalidomide for symptomatic inflammatory bowel disease. *Journal of Gastroenterology and Hepatology (Australia).* 2002;17(2):135-9.
- Bariol C, Meagher AP, Vickers CR, Byrnes DJ, Edwards PD, Hing M, et al. Early studies on the safety and efficacy of thalidomide for symptomatic inflammatory bowel disease. *J Gastroenterol Hepatol.* 2002;17(2):135-9.
- Barnes D, Park KT, Smith M, Kassam Z. Feasibility of a competitively selected universal donor fecal microbiota transplantation protocol and characterization of post-transplant microbiota modification. *Journal of Pediatric Gastroenterology and Nutrition.* 2016;63:S142-S3.
- Basu PP, Krishnaswamy N, Korapati R, Tammiseti S, Shah NJ, Hampole H, et al. A new ultra short regimen with dexamprazole, moxifloxacin, amoxicillin, nitazoxanide, and doxycycline (DeMAND) in eradication of *Helicobacter pylori*: An open-label randomized clinical trial. *International Journal of Infectious Diseases.* 2010;14:S52.
- Beecher B, Glassner P, Malchau H, Kwon YM. A concise minimum eight year follow-up of proximally porous-coated tapered titanium femoral stem in primary total hip arthroplasty. *International Orthopaedics.* 2012;36(8):1561-5.
- Belin A, Prost PL, Mercadier G, Grolhier G, Lablanche JM. Comparative study of verapamil LI 120 mg 3 times a day and verapamil LP 120 mg twice a day in stable exertional angina. A multicentre study. [French]. *Annales de Cardiologie et d'Angéiologie.* 1995;44(7):365-71.
- Belin A, Prost PL, Mercadier G, Grolhier G, Lablanche JM. [Comparative study of verapamil LI 120 mg 3 times a day and verapamil LP 120 mg twice a day in stable exertion-induced angina. A multicenter study]. *Ann Cardiol Angeiol (Paris).* 1995;44(7):365-71.
- Berg KJ, Lundby B, Reinton V, Nordal KP, Rootwelt K, Smith HJ. Gadodiamide in renal transplant patients: Effects on renal function and usefulness as a glomerular filtration rate marker. *Nephron.* 1996;72(2):212-7.
- Bharucha AE. Outcome Measures for Fecal Incontinence: Anorectal Structure and Function. *Gastroenterology.* 2004;126(1):S90-S8.

- Bhatnagar S, Bahl R, Sharma PK, Kumar GT, Saxena SK, Bhan MK. Zinc with oral rehydration therapy reduces stool output and duration of diarrhea in hospitalized children: A randomized controlled trial. *Journal of Pediatric Gastroenterology and Nutrition*. 2004;38(1):34-40.
- Bosi A, Fanci R, Pecile P, Guidi S, Saccardi R, Vannucchi AM, et al. Aztreonam versus colistin-neomycin for selective decontamination of the digestive tract in patients undergoing bone marrow transplantation: a randomized study. *J Chemother*. 1992;4(1):30-4.
- Bowden R, Murali K, Lambert K, Smyth M, Lonergan M. Chronic use of sodium polystyrene sulfonate (resonium) enables wider implementation of renin-angiotensinaldosterone inhibition in chronic kidney disease patients. *Nephrology*. 2017;22:67.
- Boyle BJ, Long WB, Balistreri WF, Widzer SJ, Huang N. Effect of cimetidine and pancreatic enzymes on serum and fecal bile acids and fat absorption in cystic fibrosis. *Gastroenterology*. 1980;78(5 Pt 1):950-3.
- Brenner D, Hiergeist A, Adis C, Gessner A, Ludolph A, Weishaupt J. The fecal microbiome ALS patients. *Amyotrophic Lateral Sclerosis and Frontotemporal Degeneration*. 2017;18 (Supplement 2):198.
- Brittnacher MJ, Heltshe SL, Hayden HS, Radey MC, Weiss EJ, Damman CJ, et al. GUTSS: An Alignment-Free Sequence Comparison Method for Use in Human Intestinal Microbiome and Fecal Microbiota Transplantation Analysis. *PLoS ONE [Electronic Resource]*. 2016;11(7):e0158897.
- Burigo T, Fagundes RLM, Trindade EBSDM, Vasconcelos HCFF. Bifidogenic effect of fructooligosaccharides in the intestinal flora of patients with hematological neoplasia. [Portuguese]. *Revista de Nutricao*. 2007;20(5):491-7.
- Butcher KS, Parsons M, MacGregor L, Barber PA, Chalk J, Bladin C, et al. Refining the perfusion-diffusion mismatch hypothesis. *Stroke*. 2005;36(6):1153-9.
- Butcher KS, Parsons M, MacGregor L, Barber PA, Chalk J, Bladin C, et al. Refining the perfusion-diffusion mismatch hypothesis. *Stroke*. 2005;36(6):1153-9.
- Byrne TA, Persinger RL, Young LS, Ziegler TR, Wilmore DW. A new treatment for patients with short-bowel syndrome. Growth hormone, glutamine, and a modified diet. *Annals of Surgery*. 1995;222(3):243-54; discussion 54-5.
- Cai CJ, Li MR, Yi SH, Wang GS, Lu MQ, Chen GH. [Application of somatostatin combined with oral vancomycin in the treatment of intestinal obstruction after liver transplantation]. *Zhonghua Wei Chang Wai Ke Za Zhi*. 2008;11(4):335-8.
- Canullo L, Quaranta A, Teles RP. The microbiota associated with implants restored with platform switching: A preliminary report. *Journal of Periodontology*. 2010;81(3):403-11.
- Cao Z, Li Z, Liu Y, Mo R, Ren P, Chen L, et al. The role of bacterial infection (BI) in decompensated cirrhosis patients with or without acute-on-chronic liver failure (ACLF). *Hepatology International*. 2017;11 (1 Supplement 1):S527-S8.

- Carter NJ, Keating GM. Micafungin: A review of its use in the prophylaxis and treatment of invasive *Candida* infections in pediatric patients. *Pediatric Drugs*. 2009;11(4):271-91.
- Carter R, Hemingway D, Cooke TG, Pickard R, Poon FW, McKillop JA, et al. A prospective study of six methods for detection of hepatic colorectal metastases. *Ann R Coll Surg Engl*. 1996;78(1):27-30.
- Cataldo PA, O'Brien S, Osler T. Transanal endoscopic microsurgery: A prospective evaluation of functional results. *Diseases of the Colon and Rectum*. 2005;48(7):1366-71.
- Cello JP, Grendell JH, Basuk P, Simon D, Weiss L, Wittner M, et al. Effect of octreotide on refractory AIDS-associated diarrhea. A prospective, multicenter clinical trial. *Annals of Internal Medicine*. 1991;115(9):705-10.
- Ceran N, Mert D, Kocdogan FY, Erdem I, Adalati R, Ozyurek S, et al. A randomized comparative study of single-dose fosfomycin and 5-day ciprofloxacin in female patients with uncomplicated lower urinary tract infections. *Journal of Infection and Chemotherapy*. 2010;16(6):424-30.
- Chakrabarti S, Collingham KE, Stevens RH, Pillay D, Fegan CD, Milligan DW. Isolation of viruses from stools in stem cell transplant recipients: A prospective surveillance study. *Bone Marrow Transplantation*. 2000;25(3):277-82.
- Cheetham M, Brazzelli M, Norton C, Glazener CM. Drug treatment for faecal incontinence in adults. *Cochrane database of systematic reviews (Online)*. 2003(3):CD002116.
- Chen YL, Cui XH, Li JL. A transposition of iliopsoas in replacement of pelvic floor for incontinence of urination and/or defecation in children. [Chinese]. *Zhonghua wai ke za zhi [Chinese journal of surgery]*. 1994;32(12):724-6.
- Chiaravalloti ND, Tulskey DS, Glosser G. Validation of the WMS-III Facial Memory subtest with the Graduate Hospital Facial Memory Test in a sample of right and left anterior temporal lobectomy patients. *J Clin Exp Neuropsychol*. 2004;26(4):484-97.
- Cho CS, Dayton MT, Thompson JS, Koltun WA, Heise CP, Harms BA. Proctocolectomy-ileal pouch-anal anastomosis for ulcerative colitis after liver transplantation for primary sclerosing cholangitis: A multi-institutional analysis. *Journal of Gastrointestinal Surgery*. 2008;12(7):1221-6.
- Cho WS, Chae C. Expression of nitric oxide synthase 2 and cyclooxygenase-2 in swine experimentally infected with *Actinobacillus pleuropneumoniae*. *Vet Pathol*. 2004;41(6):666-72.
- Chouinard LE, Schoeller DA, Watras AC, Clark RR, Close RN, Buchholz AC. Bioelectrical impedance vs. four-compartment model to assess body fat change in overweight adults. *Obesity*. 2007;15(1):85-92.
- Christine CW, Starr PA, Larson PS, Eberling JL, Jagust WJ, Hawkins RA, et al. Safety and tolerability of putaminal AADC gene therapy for Parkinson disease. *Neurology*. 2009;73(20):1662-9.
- Cichowski SB, Dunivan GC, Rogers RG, Murrietta AM, Komesu YM. Standard compared with mnemonic counseling for fecal incontinence: A randomized controlled trial. *Obstetrics and Gynecology*. 2015;125(5):1063-70.



- 1  
2  
3  
4 Ciurea SO, Saliba RM, Hamerschlag N, Karduss Aurueta AJ, Bassett R, Fernandez-Vina M, et al. Fludarabine,  
5 melphalan, thiotepe and anti-thymocyte globulin conditioning for unrelated cord blood transplant. *Leuk*  
6 *Lymphoma*. 2012;53(5):901-6.  
7  
8 Clerici C, Setchell KD, O'Connell N, Gentili G, Rusticali G, Aversa F, et al. Effect of ursodeoxycholic acid on  
9 hypertransaminasaemia and bile acid composition in patients undergoing bone marrow transplantation--a  
10 double-blind randomized control study. *Italian Journal of Gastroenterology*. 1996;28(4):191-8.  
11  
12  
13 Clerici C, Setchelli KDR, O'Connell N, Gentili G, Rusticali G, Aversa F, et al. Effect of ursodeoxycholic acid on  
14 hypertransaminasaemia and bile acid composition in patients undergoing bone marrow transplantation - A  
15 double-blind randomized control study. *Italian Journal of Gastroenterology*. 1996;28(4):191-8.  
16  
17  
18 Cocchiara G, Calderone F, Luna E, Virzi C, Agrusa A, Romano G, et al. Endoscopic treatment of colorectal  
19 polyps in a digestive endoscopy outpatient department. [Italian]. *Chirurgia italiana*. 2004;56(5):669-73.  
20  
21  
22 Cohen HS, Kimball KT. Usefulness of some current balance tests for identifying individuals with disequilibrium  
23 due to vestibular impairments. *Journal of Vestibular Research: Equilibrium and Orientation*. 2008;18(5-  
24 6):295-303.  
25  
26  
27 Collins MG, Teo E, Cole SR, Chan CY, McDonald SP, Russ GR, et al. Screening for colorectal cancer and  
28 advanced colorectal neoplasia in kidney transplant recipients: cross sectional prevalence and diagnostic  
29 accuracy study of faecal immunochemical testing for haemoglobin and colonoscopy. *Bmj*. 2012;345:e4657.  
30  
31  
32 Congilosi SM. The artificial anal sphincter. *Perspectives in Colon and Rectal Surgery*. 2000;13(1):41-51.  
33  
34 Copelan EA, Bechtel TP, Klein JP, Klein JL, Tutschka P, Kapoor N, et al. Controlled trial of orally administered  
35 immunoglobulin following bone marrow transplantation. *Bone Marrow Transplantation*. 1994;13(1):87-91.  
36  
37  
38 Corpetti G, Rosignoli MT, Dionisio P. Comparative bioavailability study of two oral formulations of ibuprofen.  
39 *Arzneimittel-Forschung/Drug Research*. 1998;48(4):392-5.  
40  
41  
42 Cox GJ, Matsui SM, Lo RS, Hinds M, Bowden RA, Hackman RC, et al. Etiology and outcome of diarrhea after  
43 marrow transplantation: a prospective study. *Gastroenterology*. 1994;107(5):1398-407.  
44  
45 Cranen K, Groothuis-Oudshoorn CG, Vollenbroek-Hutten MM, M.J IJ. Toward Patient-Centered  
46 Telerehabilitation Design: Understanding Chronic Pain Patients' Preferences for Web-Based Exercise  
47 Telerehabilitation Using a Discrete Choice Experiment. *Journal of medical Internet research*. 2017;19(1):e26.  
48  
49  
50 Cranen K, Groothuis-Oudshoorn CG, Vollenbroek-Hutten MM, MJ IJ. Toward Patient-Centered  
51 Telerehabilitation Design: Understanding Chronic Pain Patients' Preferences for Web-Based Exercise  
52 Telerehabilitation Using a Discrete Choice Experiment. *Journal of Medical Internet Research*. 2017;19(1):e26.  
53  
54  
55 Culbert P, Gillett H, Ferguson A. Highly effective oral therapy (polyethylene glycol/electrolyte solution) for  
56 faecal impaction and severe constipation. *Clinical Drug Investigation*. 1998;16(5):355-60.  
57  
58  
59 Culbert P, Gillett H, Ferguson A. Highly effective new oral therapy for faecal impaction. *British Journal of*  
60 *General Practice*. 1998;48(434):1599-600.



- Culkin DJ, Ramsey CE. Urethrectal fistula: Transanal, transsphincteric approach with locally based pedicle interposition flaps. *Journal of Urology*. 2003;169(6):2181-3.
- Curran MP. Bimatoprost: A review of its use in open-angle glaucoma and ocular hypertension. *Drugs and Aging*. 2009;26(12):1049-71.
- Damman CJ, Brittnacher MJ, Westerhoff M, Hayden HS, Radey M, Hager KR, et al. Low level engraftment and improvement following a single colonoscopic administration of fecal microbiota to patients with ulcerative colitis. *PLoS ONE*. 2015;10 (8) (no pagination)(e0133925).
- Davis SC, Yadav JS, Barrow SD, Robertson BK. Gut microbiome diversity influenced more by the Westernized dietary regime than the body mass index as assessed using effect size statistic. *MicrobiologyOpen*. 2017;6 (4) (no pagination)(e00476).
- De Caro G, Gaiani F, Duranti S, Fugazza A, Madia C, Milani C, et al. The role of bifidobacteria in ulcerative colitis: Preliminary results. *American Journal of Gastroenterology*. 2016;111:S325-S6.
- de Castro CG, Jr., Ganc AJ, Ganc RL, Petrolli MS, Hamerschlack N. Fecal microbiota transplant after hematopoietic SCT: report of a successful case. *Bone Marrow Transplantation*. 2015;50(1):145.
- De Groot PF, Kahn MT, Backhed F, Nieuwdorp M. Faecal microbiota transfer from donors post bariatric surgery does not improve insulin sensitivity in metabolic syndrome subjects. *Diabetologia*. 2016;59 (1 Supplement 1):S172-S3.
- DeJesus OT. Positron-labeled DOPA analogs to image dopamine terminals. *Drug Development Research*. 2003;59(2):249-60.
- Demetriades D, Murray JA, Chan L, Ordonez C, Bowley D, Nagy KK, et al. Penetrating colon injuries requiring resection: Diversion or primary anastomosis? An AAST prospective multicenter study. *Journal of Trauma - Injury, Infection and Critical Care*. 2001;50(5):765-75.
- Demetriades D, Murray JA, Chan LS, Ordonez C, Bowley D, Nagy KK, et al. Handsewn versus stapled anastomosis in penetrating colon injuries requiring resection: a multicenter study. *J Trauma*. 2002;52(1):117-21.
- Demetriades D, Murray JA, Chan LS, Ordonez C, Bowley D, Nagy KK, et al. Handsewn versus stapled anastomosis in penetrating colon injuries requiring resection: A multicenter study. *Journal of Trauma - Injury, Infection and Critical Care*. 2002;52(1):117-21.
- Depauw S, Bosch G, Hesta M, Whitehouse-Tedd K, Hendriks WH, Kaandorp J, et al. Fermentation of animal components in strict carnivores: A comparative study with cheetah fecal inoculum. *Journal of Animal Science*. 2012;90(8):2540-8.
- Dessinioti C. Managing adverse reactions to HPI. *Journal of the European Academy of Dermatology and Venereology*. 2017;31:16.
- Dhillon S. Argatroban: A review of its use in the management of heparin-induced thrombocytopenia.

American Journal of Cardiovascular Drugs. 2009;9(4):261-82.

Di Giulio G, Lupo L, Tirelli A, Vinci R, Rotondo A, Angelelli G. Blood flow assessment with Doppler color ultrasonography in primary and secondary tumors of the liver. [Italian]. *La Radiologia medica*. 1997;93(3):225-9.

Di Giulio G, Lupo L, Tirelli A, Vinci R, Rotondo A, Angelelli G. [Blood flow assessment with Doppler color ultrasonography in primary and secondary tumors of the liver]. *Radiol Med (Torino)*. 1997;93(3):225-9.

Dillon MT, Tubbs RS, Adunka MC, King ER, Hillman TA, Adunka OF, et al. Round window stimulation for conductive and mixed hearing loss. *Otology & neurotology: official publication of the American Otological Society, American Neurotology Society [and] European Academy of Otology and Neurotology*. 2014;35(9):1601-8.

Ding T, Telesco S, Monast C, Brodmerkel C, Yatsunenko T, Das A, et al. The gut microbiome differentiates clinical phenotypes in moderate to severe crohn's disease: Results from the certifi study. *Canadian Journal of Gastroenterology and Hepatology Conference*. 2016(pagination).

Ding T, Telesco S, Monast CS, Brodmerkel C, Yatsunenko T, Das A, et al. The gut microbiome differentiates clinical phenotypes in moderate to severe crohn's disease: Results from the CERTIFI study. *United European Gastroenterology Journal*. 2015;1):A133-A4.

Doering TM, Reaburn PR, Borges NR, Cox GR, Jenkins DG. The Effect of Higher Than Recommended Protein Feedings Post-Exercise on Recovery Following Downhill Running in Masters Triathletes. *Int J Sport Nutr Exerc Metab*. 2017;27(1):76-82.

Doi K, Kanzaki S, Kumakawa K, Usami S, Iwasaki S, Yamanaka N, et al. Evaluation of the Effectiveness and Safety in a Multi-center Clinical Trial of VIBRANT SOUNDBRIDGE in Japan. [Japanese]. *Nihon Jibiinkoka Gakkai kaiho*. 2015;118(12):1449-58.

Doi K, Kanzaki S, Kumakawa K, Usami S, Iwasaki S, Yamanaka N, et al. [Evaluation of the Effectiveness and Safety in a Multi-center Clinical Trial of VIBRANT SOUNDBRIDGE in Japan]. *Nippon Jibiinkoka Gakkai Kaiho*. 2015;118(12):1449-58.

Doki N, Suyama M, Sasajima S, Ota J, Igarashi A, Mimura I, et al. Clinical impact of pre-transplant gut microbial diversity on outcomes of allogeneic hematopoietic stem cell transplantation. *Ann Hematol*. 2017;96(9):1517-23.

Downs IA, Brandt LJ, Oneto C, Feuerstadt P, Aroniadis OC. Perceptions of fecal microbiota transplantation for diarrhea predominant irritable bowel syndrome. *American Journal of Gastroenterology*. 2016;111:S1250-S1.

Duman N, Utkutan S, Ozkan H, Ozdogan S. Are the stool characteristics of preterm infants affected by infant formulas? *The Turkish journal of pediatrics*. 2000;42(2):138-44.

Dummer R, Migden M. Long-term effects of sonidegib on tumor burden: 30-month results from the phase 2 randomized bolt trial. *Annals of Oncology*. 2017;28 (Supplement 5):v436.

- Dutheil F, Lac G, Courteix D, Dore E, Chapier R, Roszyk L, et al. Treatment of Metabolic syndrome by combination of physical activity and diet needs an optimal protein intake: A randomized controlled trial. *Nutrition Journal*. 2012;11 (1) (no pagination)(72).
- Eberling JL, Jagust WJ, Christine CW, Starr P, Larson P, Bankiewicz KS, et al. Results from a phase I safety trial of hAADC gene therapy for Parkinson disease. *Neurology*. 2008;70(21):1980-3.
- Edenfield AL, Amundsen CL, Wu JM, Levin PJ, Siddiqui NY. Posterior tibial nerve stimulation for the treatment of fecal incontinence: A systematic review. *Female Pelvic Medicine and Reconstructive Surgery*. 2013;19:S29.
- Eguchi S, Takatsuki M, Hidaka M, Soyama A, Ichikawa T, Kanematsu T. Perioperative synbiotic treatment to prevent infectious complications in patients after elective living donor liver transplantation: A prospective randomized study. *American Journal of Surgery*. 2011;201(4):498-502.
- Ehrenpreis ED, Chang D, Eichenwald E. Pharmacotherapy for fecal incontinence: A review. *Diseases of the Colon and Rectum*. 2007;50(5):641-9.
- El-Nachef N, Kassam Z, Piceno YM, Ablaza AJ, Zydek M, Elliott RJ, et al. Does rifaximin prior to fecal microbiota transplantation improve clinical outcomes compared to microbiome restoration alone in ulcerative colitis? A cohort study evaluating the impact of non-absorbable antibiotic pretreatment. *Gastroenterology*. 2017;152 (5 Supplement 1):S1008-S9.
- El-Nachef N, Piceno YM, Kassam Z, Ablaza AJ, Zydek M, Fadrosch D, et al. The role of fecal microbiota transplantation in ulcerative colitis and crohn's disease: Results from a parallel inflammatory bowel disease cohort study. *Gastroenterology*. 2017;152 (5 Supplement 1):S1008.
- Enck P, Daublin G, Lubke HJ, Strohmeyer G. Long-term efficacy of biofeedback training for fecal incontinence. *Dis Colon Rectum*. 1994;37(10):997-1001.
- Enck P, Daublin G, Lubke HJ, Strohmeyer G, Stein BL, Gordon PH. Long-term efficacy of biofeedback training for fecal incontinence. *Diseases of the Colon and Rectum*. 1994;37(10):997-1001.
- Espigado I, Aguilar-Guisado M, Martin-Pena A, Gudiol C, Falantes J, Vazquez L, et al. Discontinuing antibacterial therapy after apyrexia and clinical stability regardless of neutrophil count in febril neutropenia is safe and reduces exposition to antibiotics (howlong randomized trial). *Haematologica*. 2017;102:330-1.
- Evans RC, Shim Wong V, Morris AI, Rhodes JM. Treatment of corticosteroid-resistant ulcerative colitis with heparin - A report of 16 cases. *Alimentary Pharmacology and Therapeutics*. 1997;11(6):1037-40.
- Evans S, Daly A, Davies P, Macdonald A. Fibre content of enteral feeds for the older child. *Journal of Human Nutrition and Dietetics*. 2009;22(5):414-21.
- Fachner J, Gold C, Erkkila J. Music therapy modulates fronto-temporal activity in rest-EEG in depressed clients. *Brain Topogr*. 2013;26(2):338-54.
- Ferrara G, Sancin L, Bibalo C, Tommasini A, Taddio A, Pastore S. Faecal calprotectin as screening tool to identify inflammatory bowel disease among juvenile idiopathic patients: Results from a monocentric Italian study. *Pediatric Rheumatology*. 2017;15:118.

- 1  
2  
3  
4 Ferrecchia CE, Hobbs TR. A technique for orally administered fecal bacteriotherapy to treat chronic diarrhea  
5 in rhesus macaques (*macaca mulatta*). *Journal of the American Association for Laboratory Animal Science*.  
6 2012;51 (5):655.  
7
- 8  
9 Ferrecchia CE, Hobbs TR. Efficacy of oral fecal bacteriotherapy in rhesus macaques (*Macaca mulatta*) with  
10 chronic diarrhea. *Comp Med*. 2013;63(1):71-5.  
11
- 12  
13 Ferrucci PF, Zucca E. Primary gastric lymphoma pathogenesis and treatment: What has changed over the  
14 past 10 years? *British Journal of Haematology*. 2007;136(4):521-38.  
15
- 16  
17 Fineberg SE, Rathbun MJ, Hufferd S, Fineberg NS, Spradlin CT, Galloway JA, et al. Immunologic aspects of  
18 human proinsulin therapy. *Diabetes*. 1988;37(3):276-80.  
19
- 20  
21 Fishpool SJ, Amato-Watkins A, Hayhurst C. Free middle turbinate mucosal graft reconstruction after primary  
22 endoscopic endonasal pituitary surgery. *Eur Arch Otorhinolaryngol*. 2017;274(2):837-44.  
23
- 24  
25 Fong SS, Guo X, Cheng YT, Liu KP, Tsang WW, Yam TT, et al. A Novel Balance Training Program for Children  
26 With Developmental Coordination Disorder: A Randomized Controlled Trial. *Medicine*. 2016;95(16):e3492.  
27
- 28  
29 Fong SS, Guo X, Liu KP, Ki WY, Louie LH, Chung RC, et al. Task-Specific Balance Training Improves the Sensory  
30 Organisation of Balance Control in Children with Developmental Coordination Disorder: A Randomised  
31 Controlled Trial. *Scientific reports*. 2016;6:20945.  
32
- 33  
34 Fong SSM, Guo X, Cheng YTY, Liu KPY, Tsang WWN, Yam TTT, et al. A novel balance training program for  
35 children with developmental coordination disorder a randomized controlled trial. *Medicine (United States)*.  
36 2016;95 (16) (no pagination)(e3492).  
37
- 38  
39 Ford CD, Reilly W, Wood J, Classen DC, Burke JP. Oral antimicrobial prophylaxis in bone marrow transplant  
40 recipients: Randomized trial of ciprofloxacin versus ciprofloxacin-vancomycin. *Antimicrobial Agents and  
41 Chemotherapy*. 1998;42(6):1402-5.  
42
- 43  
44 Francini LC, Vazquez-Montes M, Buclin T, Perera R, Dunand M, Grouzmann E, et al. Pediatric reference  
45 intervals for plasma free and total metanephrines established with a parametric approach: relevance to the  
46 diagnosis of neuroblastoma. *Pediatr Blood Cancer*. 2015;62(4):587-93.  
47
- 48  
49 Freedman SF, Holgado S, Enyedi LB, Toth CA. Management of ocular torsion and diplopia after macular  
50 translocation for age-related macular degeneration: Prospective clinical study. *American Journal of  
51 Ophthalmology*. 2003;136(4):640-8.  
52
- 53  
54 Fritsch C, Lang K, Bolsen K, Lehmann P, Ruzicka T. Congenital erythropoietic porphyria. *Skin Pharmacology  
55 and Applied Skin Physiology*. 1998;11(6):347-57.  
56
- 57  
58 Garcia-Olmo D, Garcia-Arranz M, Herreros D. Expanded adipose-derived stem cells for the treatment of  
59 complex perianal fistula including Crohn's disease. *Expert Opinion on Biological Therapy*. 2008;8(9):1417-23.  
60
- 61  
62 Garcia-Plaza A, Arenas JI, Belda O, Diago A, Dominguez A, Fernandez C, et al. [A multicenter clinical trial. Zinc  
63 acexamate versus famotidine in the treatment of acute duodenal ulcer. Study Group of Zinc acexamate (new

UP doses)]. *Revista Espanola de Enfermedades Digestivas*. 1996;88(11):757-62.

Garcia-Plaza A, Arenas JI, Belda O, Diago A, Dominguez A, Fernandez C, et al. A multicentric trial of zinc acexamate versus famotidine in the treatment of acute duodenal ulcer. [Spanish]. *Revista Espanola de Enfermedades Digestivas*. 1996;88(11):757-62.

Gaughran F, Stahl D, Ismail K, Atakan Z, Lally J, Gardner-Sood P, et al. Improving physical health and reducing substance use in psychosis - randomised control trial (IMPACT RCT): Study protocol for a cluster randomised controlled trial. *BMC Psychiatry*. 2013;13 (no pagination)(263).

Gaughran F, Stahl D, Ismail K, Atakan Z, Lally J, Gardner-Sood P, et al. Improving physical health and reducing substance use in psychosis--randomised control trial (IMPACT RCT): study protocol for a cluster randomised controlled trial. *BMC Psychiatry*. 2013;13:263.

Gebhard DJ, Price J, Kennedy CE, Akcan-Arikan A. Staging of cardiorenal syndrome for outcome prediction in pediatric acute decompensated heart failure. *Intensive Care Medicine Experimental Conference: 29th Annual Congress of the European Society of Intensive Care Medicine, ESICM*. 2016;4(no pagination).

Gelissen F, Voelker M, Schwabe R, Besch D, Aisenbrey S, Szurman P, et al. Full macular translocation versus photodynamic therapy with verteporfin in the treatment of neovascular age-related macular degeneration: 1-year results of a prospective, controlled, randomised pilot trial (FMT-PDT). *Graefes Archive for Clinical and Experimental Ophthalmology*. 2007;245(8):1085-95.

Geller RB, Gilmore CE, Dix SP, Lin LS, Topping DL, Davidson TG, et al. Randomized trial of loperamide versus dose escalation of octreotide acetate for chemotherapy-induced diarrhea in bone marrow transplant and leukemia patients. *American Journal of Hematology*. 1995;50(3):167-72.

Girgis NI, Butler T, Frenck RW, Sultan Y, Brown FM, Tribble D, et al. Azithromycin versus ciprofloxacin for treatment of uncomplicated typhoid fever in a randomized trial in Egypt that included patients with multidrug resistance. *Antimicrobial Agents and Chemotherapy*. 1999;43(6):1441-4.

Giuliano M, Pantosti A, Gentile G, Venditti M, Arcese W, Martino P. Effects on oral and intestinal microfloras of norfloxacin and pefloxacin for selective decontamination in bone marrow transplant patients. *Antimicrob Agents Chemother*. 1989;33(10):1709-13.

Goel A, Aggarwal R. Prevention of hepatitis E: Another step forward. *Future Microbiology*. 2011;6(1):23-7.

Goel H, Szczepanczyk K, Bindal P, Shukla P, Tendler B, Latif S. Pituitary germinoma in adult man masquerading as pituitary apoplexy: Perils of delayed diagnosis. *Endocrine Reviews Conference: 99th Annual Meeting of the Endocrine Society, ENDO*. 2017;38(3 Supplement 1).

Gomollon F. Treatment of inflammatory bowel diseases. [Spanish]. *Gastroenterologia y Hepatologia*. 2015;38:13-9.

Gosselin KB, Feldman HA, Sonis AL, Bechard LJ, Kellogg MD, Gura K, et al. Serum citrulline as a biomarker of gastrointestinal function during hematopoietic cell transplantation in children. *Journal of Pediatric Gastroenterology and Nutrition*. 2014;58(6):709-14.

- Goyal A, Chu A, Calabro K, Firek B, Bush B, Morowitz M. Safety and efficacy of fecal microbiota transplant in children with inflammatory bowel disease. *Journal of Pediatric Gastroenterology and Nutrition*. 2016;63:S212.
- Goyal A, Kufen A, Jackson Z, Morowitz M. A study of fecal microbiota transplantation in pediatric patients with inflammatory bowel disease. *Inflammatory Bowel Diseases*. 2016;22:S74.
- Grandjean P, Acker M, Madoff R, Williams NS, Woloszko J, Kantor C. Dynamic myoplasty: Surgical transfer and stimulation of skeletal muscle for functional substitution or enhancement. *Journal of Rehabilitation Research and Development*. 1996;33(2):133-44.
- Gregory CR, Gourley IM, Cain GR, Patz JD, Imondi KA, Martin JA. Mizoribine serum levels associated with enterotoxicity in the dog. *Transplantation*. 1991;51(4):877-81.
- Gruss HJ. Macrogol 3350: Treatment of choice in severe cases of chronic constipation and faecal impaction. [German]. *Coloproctology*. 1998;20(4):161-7.
- Gu L, Ding C, Tian H, Yang B, Zhang X, Hua Y, et al. Serial frozen fecal microbiota transplantation in the treatment of chronic intestinal pseudo-obstruction: A preliminary study. *Journal of Neurogastroenterology and Motility*. 2017;23(2):289-97.
- Guay DRP. Drug forecast - The peptide deformylase inhibitors as antibacterial agents. *Therapeutics and Clinical Risk Management*. 2007;3(4):513-25.
- Guiot HF, Biemond J, Klasen E, Gratama JW, Kramps JA, Zwaan FE. Protein loss during acute graft-versus-host disease: diagnostic and clinical significance. *European journal of haematology*. 1987;38(2):187-96.
- Hadengue A, Spahr L. From bench to bedside: Is the road trickier in alcoholic liver disease? *Alcoholism: Clinical and Experimental Research*. 2010;34:53A.
- Hahn S, Kim Y, Garner P. Reduced osmolarity oral rehydration solution for treating dehydration due to diarrhoea in children: Systematic review. *British Medical Journal*. 2001;323(7304):81-5.
- Hakkinen K, Pakarinen A, Hannonen P, Hakkinen A, Airaksinen O, Valkeinen H, et al. Effects of strength training on muscle strength, cross-sectional area, maximal electromyographic activity, and serum hormones in premenopausal women with fibromyalgia. *Journal of Rheumatology*. 2002;29(6):1287-95.
- Halibasic E, Fuerst E, Heiden D, Japtok L, Diesner SC, Hillebrand P, et al. Significantly reduced plasma levels of the bioactive sphingolipid S1P in lung transplanted cystic fibrosis patients are associated with gastrointestinal symptoms. *Allergy: European Journal of Allergy and Clinical Immunology*. 2017;72:195.
- Hansson J, Hauschild A, Kunstfeld R, Jacques Grob J, Dreno B, Mortier L, et al. Vismodegib (VISMO), a hedgehog pathway inhibitor (HPI), in advanced basal cell carcinoma (aBCC): STEVIE study primary analysis in 1215 patients (pts). *Journal of Clinical Oncology Conference*. 2016;34(no pagination).
- Hasper D, Schefold JC, Baumgart DC. Management of severe abdominal infections. *Recent Patents on Anti-Infective Drug Discovery*. 2009;4(1):57-65.



- Haveman LM, De Jager W, Van Loon AM, Claas ECJ, Prakken BJ, Bierings M. Different cytokine signatures in children with localized and invasive adenovirus infection after stem cell transplantation. *Pediatric Transplantation*. 2010;14(4):520-8.
- Hellinger WC, Yao JD, Alvarez S, Blair JE, Cawley JJ, Paya CV, et al. A randomized, prospective, double-blinded evaluation of selective bowel decontamination in liver transplantation. *Transplantation*. 2002;73(12):1904-9.
- Hemingway DM, Cooke TG, Warren H, Bessent RG, McKillop JH, McArdle CS. Dynamic hepatic scintigraphy in colorectal cancer. *Nucl Med Commun*. 1995;16(10):867-9.
- Hendrickson R, Ryan J, Dandridge L, Andrews W, Daniel J, Fischer R, et al. Conservative management of pneumatosis intestinalis after pediatric liver transplantation. *American Journal of Transplantation*. 2017;17:605.
- Hermes F, Haudebourg L, Bagot M, Dutriaux C, Grob JJ, Guillot B, et al. Follow-up of patients with complete remission of locally advanced basal cell carcinoma treated with vismodegib after treatment discontinuation: A retrospective multicentric French study. *Journal of Clinical Oncology Conference*. 2017;35(15 Supplement 1).
- Hickman C, Wells D, Gwinnett D, Wilkinson T, Christiansen S, Olliana O, et al. Euploid rate sensitivity to laboratory culture environment: A blind, prospective, randomised, sibling study. *Human Reproduction*. 2016;31:i216-i8.
- Ho KS, Ho YH. Controlled, randomized trial of island flap anoplasty for treatment of trans-sphincteric fistula-in-ano: Early results. *Techniques in Coloproctology*. 2005;9(2):166-8.
- Holger Johnsen P, Mazzawi T, El-Salhy M, Hausken T, Goll R, Valle PC. Effect of faecal microbiota transplantation on the enteroendocrine cells of the colon in patients with Irritable Bowel Syndrome (IBS): Double blinded-placebo controlled study. *Neurogastroenterology and Motility*. 2017;29:71.
- Holster S, Brummer RJ, Repsilber D, König J. Fecal microbiota transplantation in irritable bowel syndrome and a randomized placebo-controlled trial. *Gastroenterology*. 2017;152 (5 Supplement 1):S101-S2.
- Holvoet T, Boelens J, Joossens M, Raes J, De Vos M, De Looze D. Fecal microbiota transplantation in irritable bowel syndrome with bloating: Results from a prospective pilot study. *Gastroenterology*. 2015;1):S963-S4.
- Hoppe B, Beck B, Gatter N, von Unruh G, Tischer A, Hesse A, et al. *Oxalobacter formigenes*: a potential tool for the treatment of primary hyperoxaluria type 1. *Kidney Int*. 2006;70(7):1305-11.
- Horrocks E, Bremner SA, Stevens N, Norton C, Eldridge S, Knowles CH. Double blind randomised controlled trial of Percutaneous Tibial Nerve Stimulation (PTNS) VS. sham electrical stimulation in the treatment of faecal incontinence. *Gastroenterology*. 2015;1):S177.
- Horrocks E, Bremner SA, Stevens N, Norton C, O'Connell PR, Eldridge S, et al. Double blind randomised controlled trial of percutaneous tibial nerve stimulation for the treatment of faecal incontinence in adults. *Gut*. 2015;64:A4.



- Hoverstad T, Carlstedt-Duke B, Lingaas E, Norin E, Saxerholt H, Steinbakk M, et al. Influence of oral intake of seven different antibiotics on faecal short-chain fatty acid excretion in healthy subjects. *Scandinavian Journal of Gastroenterology*. 1986;21(8):997-1003.
- Hsu YF, Huang YZ, Lin YY, Tang CW, Liao KK, Lee PL, et al. Intermittent theta burst stimulation over ipsilesional primary motor cortex of subacute ischemic stroke patients: A pilot study. *Brain Stimulation*. 2013;6(2):166-74.
- Hu XY, Zhang Y, Chen G, Zhong S, Fan XJ. A prospective cohort study on the influence of high doses of herbs for clearing heat and resolving stasis on survival rates in patients with hepatitis B-related acute-on-chronic liver failure. *Journal of Chinese Integrative Medicine*. 2012;10(2):176-85.
- Huang W, Wan X. Update of critical care medicine 2013. [Chinese]. *Zhonghua Wei Zhong Bing Ji Jiu Yi Xue*. 2014;26(1):3-10.
- Huang Z, Huang Y, Peng K, Li X, Cheng X, Zhao R, et al. The mutation of interleukin-10/interleukin-10 receptors and clinical characterization of Chinese children with very early onset inflammatory bowel disease: A survey of Chinese very early onset inflammatory bowel disease study group. *Journal of Pediatric Gastroenterology and Nutrition*. 2016;63:S376.
- Hudson CO, Karp DR, Pratt T, Northington GM. Anticholinergic therapy and fecal incontinence symptoms in patients with dual incontinence: A pilot study. *Female Pelvic Medicine and Reconstructive Surgery*. 2016;22 (5 Supplement 1):S108.
- Huijgens PC, Simoons-Smit AM, Van Loenen AC, Prooy E, Van Tinteren H, Ossenkoppele GJ, et al. Fluconazole versus itraconazole for the prevention of fungal infections in haemato-oncology. *Journal of Clinical Pathology*. 1999;52(5):376-80.
- Husain M, Khan RN, Rehmani B, Haris H. Omental patch technique for the ileal perforation secondary to typhoid fever. *Saudi journal of gastroenterology: official journal of the Saudi Gastroenterology Association*. 2011;17(3):208-11.
- Ibanez-Cervantes G, Bello-Lopez JM, Fernandez-Sanchez V, Dominguez-Mendoza CA, Acevedo-Alfaro LI. Prevalence of bacterial contamination in platelet concentrates at the National Center of Blood Transfusion (Mexico). *Transfusion Clinique et Biologique*. 2017;24(2):56-61.
- Inoue T, Koyama K, Oriuchi N, Alyafei S, Yuan Z, Suzuki H, et al. Detection of malignant tumors: Whole-body PET with fluorine 18 alpha-methyl tyrosine versus FDG - Preliminary study. *Radiology*. 2001;220(1):54-62.
- Inoue T, Koyama K, Oriuchi N, Alyafei S, Yuan Z, Suzuki H, et al. Detection of malignant tumors: whole-body PET with fluorine 18 alpha-methyl tyrosine versus FDG--preliminary study. *Radiology*. 2001;220(1):54-62.
- Inoue T, Shibasaki T, Oriuchi N, Aoyagi K, Tomiyoshi K, Amano S, et al. 18F alpha-methyl tyrosine PET studies in patients with brain tumors. *Journal of Nuclear Medicine*. 1999;40(3):399-405.
- Inoue T, Shibasaki T, Oriuchi N, Aoyagi K, Tomiyoshi K, Amano S, et al. <sup>18</sup>F alpha-methyl tyrosine PET studies in patients with brain tumors. *Journal of Nuclear Medicine*. 1999;40(3):399-405.

- Ishihara S, Kaji T, Kawamura A, Rumi MA, Sato H, Okuyama T, et al. Diagnostic accuracy of a new non-invasive enzyme immunoassay for detecting *Helicobacter pylori* in stools after eradication therapy. *Aliment Pharmacol Ther.* 2000;14(5):611-4.
- Shikawa D, Sasaki T, Osada T, Kuwahara-Arai K, Haga K, Shibuya T, et al. Changes in intestinal microbiota following combination therapy with fecal microbial transplantation and antibiotics for ulcerative colitis. *Inflammatory Bowel Diseases.* 2017;23(1):116-25.
- Ivanyi JL, Plander M, Szendrei T, Toth C. Prevention and treatment of invasive fungal infections in patients with hematological malignancies-results from a single hematological centre. *Haematologica.* 2016;101:765.
- Jaafari A, Boukhriss B, Selmi K, Bencheikh M, Boussabah E, Benyoussef S. [Echocardiography under perfusion with dobutamine. Experience of Tunisian cardiologic service. About 70 cases]. *Tunis Med.* 2004;82(4):373-6.
- Jalanka J, Salonen A, Salojärvi J, Ritari J, Immonen O, Marciani L, et al. Effects of bowel cleansing on the intestinal microbiota. *Gut.* 2015;64(10):1562-8.
- Jaworski A, Mitchell SW, Wong C, Gadalla S, Borody TJ. Patient with relapsing *C. difficile* successfully treated with lyophilised encapsulated faecal microbiota transplant product. *Journal of Gastroenterology and Hepatology (Australia).* 2016;31:161.
- Jian Z, Hatib F, Pinsky M. Prevalence of hypotension and prediction of hypotension in intensive care unit. *Critical Care Conference: 37th International Symposium on Intensive Care and Emergency Medicine Belgium.* 2017;21(1 Supplement 1).
- Johnsen PH, Hilpusch F, Cavanagh JP, Sande Leikanger I, Kolstad C, Valle PC, et al. Fecal transplantation in Irritable Bowel Syndrome (IBS): An RCT. *Neurogastroenterology and Motility.* 2017;29:135.
- Jolly S, Lobo A. Neuraxial analgesia in the laboring parturient with arnold-chiari type i malformation-relief of pain in uncharted terrain? *Regional Anesthesia and Pain Medicine Conference: 41st Annual Regional Anesthesiology and Acute Pain Medicine Meeting of the American Society of Regional Anesthesia and Pain Medicine, ASRA.* 2016;41(5).
- Jones C, Shannon B. Placebo responders in a randomised controlled trial of rbx2660 for recurrent *c. difficile* infection: Predictive value of 16 s rRNA microbiome analysis. *United European Gastroenterology Journal.* 2016;4 (5 Supplement 1):A652.
- Jones MP, Talley NJ, Nuyts G, Dubois D. Lack of objective evidence of efficacy of laxatives in chronic constipation. *Digestive Diseases and Sciences.* 2002;47(10):2222-30.
- Joshi NM, Goodhand J, Alazawi W, Das S, Wilks M, Rampton D. Predicting treatment failure in *C. difficile* infection: A prospective observational cohort study. *Gut.* 2016;65:A209.
- Journois D, Safran D, Castelain MH, Chanu D, Drevillon C, Barrier G. [Comparison of the antithrombotic effects of heparin, enoxaparin and prostacycline in continuous hemofiltration]. *Ann Fr Anesth Reanim.* 1990;9(4):331-7.
- Joyce MR, Hull TL. Endoanal Advancement Flaps in the Management of Complex Anorectal Fistulas. *Seminars*

in Colon and Rectal Surgery. 2009;20(1):24-31.

Jung K, Kang BK, Kim JY, Shin KS, Lee CS, Song DS. Effects of epidermal growth factor on atrophic enteritis in piglets induced by experimental porcine epidemic diarrhoea virus. *Vet J*. 2008;177(2):231-5.

Kaido T, Shimamura T, Sugawara Y, Sadamori H, Shirabe K, Yamamoto M, et al. Multicentre, randomised, placebocontrolled trial of extract of Japanese herbal medicine Daikenchuto to prevent bowel dysfunction after adult liver transplantation (DKB 14 Study). *BMJ Open*. 2015;5 (9) (no pagination)(e008356).

Kaido T, Shimamura T, Sugawara Y, Sadamori H, Shirabe K, Yamamoto M, et al. Multicentre, randomised, placebo-controlled trial of extract of Japanese herbal medicine Daikenchuto to prevent bowel dysfunction after adult liver transplantation (DKB 14 Study). *BMJ Open*. 2015;5(9):e008356.

Kajbafzadeh A. The dream of functional organ engineering and preclinical transplantation. *Iranian Journal of Biotechnology*. 2017;ISSUE):43-5.

Kajbafzadeh AM. Tissue engineering in pediatric urology reconstruction. *International Journal of Urology*. 2012;19:250.

Kakihana K, Fujioka Y, Suda W, Najima Y, Kuwata G, Sasajima S, et al. Fecal microbiota transplantation for patients with steroid-resistant acute graft-versus-host disease of the gut. *Blood*. 2016;128(16):2083-8.

Kallarackal GU, Ansari EA, Amos N, Martin JC, Lane C, Camilleri JP. A comparative study to assess the clinical use of Fluorescein Meniscus Time (FMT) with Tear Break up Time (TBUT) and Schirmer's tests (ST) in the diagnosis of dry eyes. *Eye*. 2002;16(5):594-600.

Kanauchi O, Suga T, Tochiara M, Hibi T, Naganuma M, Homma T, et al. Treatment of ulcerative colitis by feeding with germinated barley foodstuff: First report of a multicenter open control trial. *Journal of Gastroenterology*. 2002;37(SUPPL. 14):67-72.

Kang DW, Adams JB, Gregory AC, Borody T, Chittick L, Fasano A, et al. Microbiota Transfer Therapy alters gut ecosystem and improves gastrointestinal and autism symptoms: An open-label study. *Microbiome*. 2017;5 (1) (no pagination)(10).

Kang SB, Lee TG. Muscle regeneration: Research for the treatment of fecal incontinence. [Korean]. *Journal of the Korean Society of Coloproctology*. 2010;26(1):1-7.

Kao SS, Wu DC, Tsay FWT, Tsai KW, Hsu PI. A randomized controlled study comparing 14-day reverse hybrid and bismuth quadruple therapies for helicobacter pylori infection and impacts on clarithromycin resistance of gut microbiota. *Gastroenterology*. 2017;152 (5 Supplement 1):S183.

Kato K, Sekizuka T, Sugiyama T, Ishii Y, Kuroda M, Ohkusa T. Characterization of gut microbiome associated with improvement of ulcerative colitis after antibiotic combination therapy using fecal metagenomic analysis. *United European Gastroenterology Journal*. 2017;5 (5 Supplement 1):A264-A5.

Kaufmann S, Horger T, Oelker A, Kloth C, Nikolaou K, Schulze M, et al. Characterization of hepatocellular carcinoma (HCC) lesions using a novel CT-based volume perfusion (VPCT) technique. *European Journal of Radiology*. 2015;84(6):1029-35.

- Kawecki D, Chmura A, Pacholczyk M, Lagiewska B, Adadynski L, Wasiak D, et al. Bacterial infections in the early period after liver transplantation: Etiological agents and their susceptibility. *Medical Science Monitor*. 2009;15(12):CR628-CR37.
- Keshaw H, Foong KS, Forbes A, Day RM. Perianal fistulae in Crohn's disease: Current and future approaches to treatment. *Inflammatory Bowel Diseases*. 2010;16(5):870-80.
- Khoruts A. Implementation of colorectal cancer guidelines. *Pediatric Pulmonology*. 2016;51:184.
- Kim CH, Oh Y, Han K, Seo HW, Kim D, Kang I, et al. Expression of secreted mucins (MUC2, MUC5AC, MUC5B, and MUC6) and membrane-bound mucin (MUC4) in the lungs of pigs experimentally infected with *Actinobacillus pleuropneumoniae*. *Res Vet Sci*. 2012;92(3):486-91.
- Kirk KF, Kousgaard SJ, Nielsen HL, Nielsen H, Thorlacius-Ussing O. Faecal transplant for the treatment of chronic pouchitis-A randomised, placebo-controlled, clinical trial. *Colorectal Disease*. 2017;19 (Supplement 2):143.
- Kisiel JB, Taylor WR, Allawi H, Yab TC, Simonson JA, Devens ME, et al. Detection of colorectal cancer and polyps in patients with inflammatory bowel disease by novel methylated stool DNA markers. *Gastroenterology*. 2014;1:S440-S1.
- Knowles CH, Horrocks EJ, Bremner SA, Stevens N, Norton C, O'Connell PR, et al. Percutaneous tibial nerve stimulation versus sham electrical stimulation for the treatment of faecal incontinence in adults (CONFIDENT): A double-blind, multicentre, pragmatic, parallel-group, randomised controlled trial. *The Lancet*. 2015;386(10004):1640-8.
- Knowles CH, Horrocks EJ, Bremner SA, Stevens N, Norton C, O'Connell PR, et al. Percutaneous tibial nerve stimulation versus sham electrical stimulation for the treatment of faecal incontinence in adults (CONFIDENT): a double-blind, multicentre, pragmatic, parallel-group, randomised controlled trial. *Lancet*. 2015;386(10004):1640-8.
- Ko CY, Tong J, Lehman RE, Shelton AA, Schrock TR, Welton ML. Biofeedback is effective therapy for fecal incontinence and constipation. *Arch Surg*. 1997;132(8):829-33; discussion 33-4.
- Komura T, Miura K, Shirasaka T, Ohnuma S, Shimada M, Kajiwarra T, et al. Usefulness of alternate-day administration of S-1 and leucovorin in a xenograft mouse model of colorectal cancer: a shorter drug-free interval leads to more efficient antitumor effects. *International Journal of Clinical Oncology*. 2015;20(1):117-25.
- Komura T, Ohnuma S, Miura K, Shirasaka T, Kajiwarra T, Kudoh K, et al. Usefulness of alternate-day administration of S-1 and leucovorin in a xenograft mouse model of colorectal cancer: A shorter drug-free interval leads to more efficient antitumor effects. *Cancer Research Conference: 105th Annual Meeting of the American Association for Cancer Research, AACR*. 2014;74(19 SUPPL. 1).
- Kovatcheva-Datchary P, Nilsson A, Akrami R, Lee YS, De Vadder F, Arora T, et al. Dietary Fiber-Induced Improvement in Glucose Metabolism Is Associated with Increased Abundance of *Prevotella*. *Cell Metabolism*. 2015;22(6):971-82.

- Koyama D, Murata M, Hanajiri R, Okuno S, Kamoshita S, Julamanee J, et al. REG3A polymorphism is associated with the incidence of extensive chronic Gvhd after allogeneic BMT. Blood Conference: 58th Annual Meeting of the American Society of Hematology, ASH. 2016;128(22).
- Kramer P, Peters B, Schubert S, Photiadis J, Berger F, Ovroutski S. Treatment strategies for protein-losing enteropathy in Fontan patients. *Cardiology in the Young*. 2016;26:S54.
- Krammer HJ, Kamper H, Von Bunau R, Zieseniss E, Stange C, Schlieger F, et al. Probiotic drug therapy with *E. coli* strain Nissle 1917 (EcN): Results of a prospective study of the records of 3807 patients. [German]. *Zeitschrift fur Gastroenterologie*. 2006;44(8):651-6.
- Krammer HJ, Kamper H, von Bunau R, Zieseniss E, Stange C, Schlieger F, et al. [Probiotic drug therapy with *E. coli* strain Nissle 1917 (EcN): results of a prospective study of the records of 3,807 patients]. *Zeitschrift fur Gastroenterologie*. 2006;44(8):651-6.
- Kumakawa K, Kanzaki S, Usami S, Iwasaki S, Yamanaka N, Doi K, et al. Multicenter Clinical Study of Vibrant Soundbridge in Japan: Analysis of Subjective Questionnaires. [Japanese]. *Nihon Jibiinkoka Gakkai kaiho*. 2015;118(11):1309-18.
- Kumakawa K, Kanzaki S, Usami S, Iwasaki S, Yamanaka N, Doi K, et al. [Multicenter Clinical Study of Vibrant Soundbridge in Japan: Analysis of Subjective Questionnaires]. *Nippon Jibiinkoka Gakkai Kaiho*. 2015;118(11):1309-18.
- Kumar R, Maynard CL, Eipers P, Goldsmith KT, Ptacek T, Grubbs JA, et al. Colonization potential to reconstitute a microbe community in patients detected early after fecal microbe transplant for recurrent *C. difficile*. *BMC Microbiology*. 2016;16:5.
- Kushnir J, Sadeh A. Assessment of brief interventions for nighttime fears in preschool children. *Eur J Pediatr*. 2012;171(1):67-75.
- Lacima G, Pera M, Amador A, Escaramis G, Pique JM. Long-term results of biofeedback treatment for faecal incontinence: A comparative study with untreated controls. *Colorectal Disease*. 2010;12(8):742-9.
- Lad Y, Badin RA, Binley K, Van Camp N, Jan C, Gourlay J, et al. OXB-102: An enhanced gene therapy for Parkinson's disease. *Human Gene Therapy*. 2015;26 (10):A82.
- Ladas EJ, Bhatia M, Chen L, Sandler E, Petrovic A, Berman DM, et al. The safety and feasibility of probiotics in children and adolescents undergoing hematopoietic cell transplantation. *Bone Marrow Transplantation*. 2016;51(2):262-6.
- Lamere B, Wendt ER, Kanwar B, Lynch SV. Investigating the microbiome in a phase 1B study of andecaliximab in ulcerative colitis. *United European Gastroenterology Journal*. 2017;5 (5 Supplement 1):A263.
- Lang PJ, Schlegel PG, Meisel R, Schulz AS, Greil J, Bader P, et al. TCR-alpha/beta and CD19 depleted haploidentical stem cell transplantation following reduced intensity conditioning in children: First results of a prospective multicenter phase I/II clinical trial. Blood Conference: 58th Annual Meeting of the American Society of Hematology, ASH. 2016;128(22).



- Lee K, Byun B, Kim B, Lim I, Choi C, Youn S, et al. Microdose study for amino acid imaging using D-<sup>18</sup>F]FMT PET in human brains. *European Journal of Nuclear Medicine and Molecular Imaging*. 2017;44 (2 Supplement 1):S527.
- Lee KC, Byun BH, Kim BI, Lim I, Choi CW, Youn SM, et al. A phase 0 study for amino acid imaging using D-<sup>18</sup>F]FMT PET in human brains. *Journal of Labelled Compounds and Radiopharmaceuticals*. 2017;60:S182.
- Lee SH, Carey S, Dubey R, Matz R. Intervention program in college instrumental musicians, with kinematics analysis of cello and flute playing: a combined program of yogic breathing and muscle strengthening-flexibility exercises. *Med Probl Perform Art*. 2012;27(2):85-94.
- Lenisa L, Espin-Basany E, Rusconi A, Mascheroni L, Escoll-Rufino J, Lozoya-Trujillo R, et al. Anal fistula plug is a valid alternative option for the treatment of complex anal fistula in the long term. *International Journal of Colorectal Disease*. 2010;25(12):1487-93.
- Leong L, Choo J, Serisier D, Rogers G. Long-term erythromycin therapy affects microbiota composition and antibiotic resistance gene prevalence in the oropharynx of bronchiectasis patients. *Respirology*. 2015;20:29.
- Leung LY, Lim HK, Abell MW, Zimmerman JJ. Pharmacokinetics and metabolic disposition of sirolimus in healthy male volunteers after a single oral dose. *Ther Drug Monit*. 2006;28(1):51-61.
- Leuschner U, Guldutuna S, Imhof M, Hubner K, Benjaminov A, Leuschner M. Effects of ursodeoxycholic acid after 4 to 12 years of therapy in early and late stages of primary biliary cirrhosis. *Journal of Hepatology*. 1994;21(4):624-33.
- Lewis JD, Reinisch W, Bressler B, Parikh A, Yang H, Rosario M, et al. Faecal calprotectin reductions in patients achieving mucosal healing with vedolizumab induction therapy in GEMINI 1. *Journal of Crohn's and Colitis*. 2016;10:S206-S8.
- Lim TY, Pavlidis P, Pirani T, Gulati S, Samaan M, Chung-Faye G, et al. Vedolizumab in primary and autoimmune sclerosing cholangitis associated inflammatory bowel disease pre and post liver transplantation: A case series. *Gut*. 2016;65:A89.
- Lin E, Jaworski A, Furnari V, Wong C, Bull M, Chapman B, et al. Twelve week storage trial of microbial viability in lyophilized and frozen fecal microbiota preparations. *Gastroenterology*. 2015;1:S962.
- Lin WY, Wang SJ, Yeh SH. Hepatic perfusion index in evaluating treatment effect of transcatheter hepatic artery embolization in patients with hepatocellular carcinoma. *Neoplasma*. 1995;42(2):89-92.
- Lista F, Redondo C, Meilan E, Garcia-Tello A, Ramon de Fata F, Angulo JC. Efficacy and safety of fosfomycin-trometamol in the prophylaxis for transrectal prostate biopsy. Prospective randomized comparison with ciprofloxacin. *Actas urológicas españolas*. 2014;38(6):391-6.
- Lorenz F, Marklund S, Werner M, Palmqvist R, Wahlin BE, Wahlin A. Fecal calprotectin as a biomarker of intestinal graft versus host disease after allogeneic hematopoietic stem cell transplantation. *Scientific reports*. 2015;5:7920.

- Luber RP, Kariyawasam VC, Dawson LP, Munari SC, Gibson PR, Sparrow MP, et al. Combination therapy with infliximab and a thiopurine vs. infliximab monotherapy in Crohn's disease. *Journal of Gastroenterology and Hepatology (Australia)*. 2016;31:141-2.
- Luke M, Ziemssen F, Bartz-Schmidt KU, Gelissen F. Quality of life in a prospective, randomised pilot-trial of photodynamic therapy versus full macular translocation in treatment of neovascular age-related macular degeneration - A report of 1 year result. *Graefe's Archive for Clinical and Experimental Ophthalmology*. 2007;245(12):1831-6.
- Luke M, Ziemssen F, Bartz-Schmidt KU, Gelissen F. Quality of life in a prospective, randomised pilot-trial of photodynamic therapy versus full macular translocation in treatment of neovascular age-related macular degeneration--a report of 1 year results. *Graefes Arch Clin Exp Ophthalmol*. 2007;245(12):1831-6.
- Luke M, Ziemssen F, Voker M, Altpeter E, Beutel J, Besch D, et al. Full macular translocation (FMT) versus photodynamic therapy (PDT) with verteporfin in the treatment of neovascular age-related macular degeneration: 2-year results of a prospective, controlled, randomised pilot trial (FMT-PDT). *Graefe's Archive for Clinical and Experimental Ophthalmology*. 2009;247(6):745-54.
- Luke M, Ziemssen F, Volker M, Altpeter E, Beutel J, Besch D, et al. Full macular translocation (FMT) versus photodynamic therapy (PDT) with verteporfin in the treatment of neovascular age-related macular degeneration: 2-year results of a prospective, controlled, randomised pilot trial (FMT-PDT). *Graefes Arch Clin Exp Ophthalmol*. 2009;247(6):745-54.
- Madoff RD. Surgical Treatment Options for Fecal Incontinence. *Gastroenterology*. 2004;126(1):S48-S54.
- Mady FM, Abou-Taleb AE, Khaled KA, Yamasaki K, Iohara D, Taguchi K, et al. Evaluation of carboxymethyl-beta-cyclodextrin with acid function: Improvement of chemical stability, oral bioavailability and bitter taste of famotidine. *International Journal of Pharmaceutics*. 2010;397(1-2):1-8.
- Makhlough A, Fakheri H, Hojati S, Hosseini V, Bari Z. A comparison between hybrid therapy and standard triple therapy for *Helicobacter pylori* eradication in patients with uremia: A randomized clinical trial. *Middle East Journal of Digestive Diseases*. 2016;8(1):39-43.
- Mann PA, McNicholas PM, Chau AS, Patel R, Mendrick C, Ullmann AJ, et al. Impact of antifungal prophylaxis on colonization and azole susceptibility of *Candida* species. *Antimicrob Agents Chemother*. 2009;53(12):5026-34.
- Mansour-Ghanaei F, Shafaghi A, Fallah M. The effect of metronidazole in treating human fascioliasis. *Medical Science Monitor*. 2003;9(10):PI127-PI30.
- Mariotti G, Quaranta A, Merli M, Paterno Holtzman L, Piemontese M. Chronic periodontitis and cardiovascular disease: A controlled clinical trial. *European Journal of Inflammation*. 2013;11(2):459-67.
- Marquez HP, Karalli A, Haubenreisser H, Mathew RP, Alkadhi H, Brismar TB, et al. Computed tomography perfusion imaging for monitoring transarterial chemoembolization of hepatocellular carcinoma. *European Journal of Radiology*. 2017;91:160-7.



- Mathews V, Srivastava A, George B, Korula A, Perumalla S, Abubacker FN, et al. Multi-drug resistant organisms are common in fecal surveillance cultures and do not predict bacteremia but correlate with poorer outcomes in patients undergoing allogeneic stem cell transplants. *Blood Conference: 58th Annual Meeting of the American Society of Hematology, ASH*. 2016;128(22).
- Maughan J, Parkin A, Smith AH, Barker MC, Robinson PJ, Finan P, et al. Hepatic perfusion index: a multicentre trial. *Nucl Med Commun*. 1992;13(3):161-7.
- Mazique DC. Anchors away: Anchoring bias, confirmation bias, and pulmonary emboli. *Journal of General Internal Medicine*. 2017;32 (2 Supplement 1):S447.
- McLauchlin J, Amar CFL, Pedraza-Diaz S, Mieli-Vergani G, Hadzic N, Davies EG. Polymerase chain reaction-based diagnosis of infection with *Cryptosporidium* in children with primary immunodeficiencies. *Pediatric Infectious Disease Journal*. 2003;22(4):329-34.
- McNeil SA, Malani PN, Chenoweth CE, Fontana RJ, Magee JC, Punch JD, et al. Vancomycin-resistant enterococcal colonization and infection in liver transplant candidates and recipients: a prospective surveillance study. *Clinical Infectious Diseases*. 2006;42(2):195-203.
- Metafuni E, Giammarco S, De Ritis D, Rossi M, Bacigalupo A, Sica S, et al. Comparison between serum and fecal calprotectin as marker of graft-versus-host disease. *Haematologica*. 2016;101:868-9.
- Michot F, Lefebure B, Bridoux V, Gourcerol G, Kianifard B, Leroi AM, et al. Artificial anal sphincter for severe fecal incontinence implanted by a transvaginal approach: Experience with 32 patients treated at one institution. *Diseases of the Colon and Rectum*. 2010;53(8):1155-60.
- Mielcarek M, Furlong T, Storer BE, Green ML, Carpenter PA, McDonald GB, et al. Efficacy and safety of lower-dose glucocorticoids for initial treatment of acute graft-versus-host disease: A randomized controlled trial. *Blood Conference: 55th Annual Meeting of the American Society of Hematology, ASH*. 2013;122(21).
- Migden M. Sonidegib's duration of response: Results from the pivotal phase 2 BOLT study over 30 months in patients with locally advanced basal cell carcinoma. *Journal of the European Academy of Dermatology and Venereology*. 2017;31:39.
- Miyazaki K, Collins DJ, Walker-Samuel S, Taylor JN, Padhani AR, Leach MO, et al. Quantitative mapping of hepatic perfusion index using MR imaging: A potential reproducible tool for assessing tumour response to treatment with the antiangiogenic compound BIBF 1120, a potent triple angiokinase inhibitor. *European Radiology*. 2008;18(7):1414-21.
- Mizukami K, Sonoda A, Okimoto T, Kodama M, Murakami K. An open-label prospective randomized multicentre study of daily granulocyte and monocyte adsorptive apheresis as compared with intensive treatment in patients with active ulcerative colitis. *Journal of Crohn's and Colitis*. 2015;9:S352.
- Modiba MCM, Koto Z, Lowan TA, Magano S, Segal I, Esser J, et al. Distal splenorenal shunt for non-cirrhotic variceal bleeding in black South Africans. *South African Journal of Surgery*. 1994;32(3):87-90.
- Moen MD, McKeage K, Plosker GL, Siddiqui MAA. Imatinib: A review of its use in chronic myeloid leukaemia. *Drugs*. 2007;67(2):299-320.

- Molina JM, Tourneur M, Sarfati C, Chevret S, De Gouvello A, Gobert JG, et al. Fumagillin treatment of intestinal microsporidiosis. *New England Journal of Medicine*. 2002;346(25):1963-9.
- Molina JM, Tourneur M, Sarfati C, Chevret S, de Gouvello A, Gobert JG, et al. Fumagillin treatment of intestinal microsporidiosis. *New England Journal of Medicine*. 2002;346(25):1963-9.
- Moller A, Iwasaki K, Kawamura A, Teramura Y, Shiraga T, Hata T, et al. The disposition of <sup>14</sup>C-labeled tacrolimus after intravenous and oral administration in healthy human subjects. *Drug Metab Dispos*. 1999;27(6):633-6.
- Montebugnoli L, Venturi M, Cervellati F, Servidio D, Vocale C, Pagan F, et al. Peri-Implant Response and Microflora in Organ Transplant Patients 1 Year after Prosthetic Loading: A Prospective Controlled Study. *Clinical implant dentistry and related research*. 2015;17(5):972-82.
- Mortimer K, Brown A, Feary J, Jagger C, Lewis S, Antoniak M, et al. Dose-ranging study for trials of therapeutic infection with necator *Americanus* in humans. *American Journal of Tropical Medicine and Hygiene*. 2006;75(5):914-20.
- Muller T, Buttner T, Gholipour AF, Kuhn W. Coenzyme Q<sub>10</sub> supplementation provides mild symptomatic benefit in patients with Parkinson's disease. *Neuroscience Letters*. 2003;341(3):201-4.
- Muller T, Buttner T, Gholipour AF, Kuhn W. Coenzyme Q<sub>10</sub> supplementation provides mild symptomatic benefit in patients with Parkinson's disease. *Neuroscience Letters*. 2003;341(3):201-4.
- Muraji T, Nishijima E, Higashimoto Y, Tsugawa C. Biliary atresia: current management and outcome. *The Tohoku journal of experimental medicine*. 1997;181(1):155-60.
- Muramatsu SI. A phase i study of aromatic l-amino acid decarboxylase gene therapy for parkinson's disease. *Journal of Gene Medicine*. 2014;16 (7-8):218.
- Muramatsu SI. In vivo imaging in cell and gene therapy for parkinson's disease. *Journal of Gene Medicine*. 2014;16 (7-8):214.
- Muramatsu SI, Fujimoto KI, Kato S, Asari S, Mizukami H, Ikeguchi K, et al. Aadc gene therapy for parkinson's disease: Four years of follow-up. *Journal of Gene Medicine*. 2014;16 (7-8):220.
- Muramatsu SI, Fujimoto KI, Kato S, Mizukami H, Asari S, Ikeguchi K, et al. A phase i study of aromatic l-amino acid decarboxylase gene therapy for parkinson's disease. *Molecular Therapy*. 2010;18(9):1731-5.
- Naeini AE, Sharifi M, Shahidi S, Taheri S, Seirafian S, Taheri D, et al. Intestinal fungal and parasitic infections in kidney transplant recipients: a multi-center study. *Saudi journal of kidney diseases and transplantation: an official publication of the Saudi Center for Organ Transplantation, Saudi Arabia*. 2012;23(4):677-83.
- Nagappan V, Deresinski S. Posaconazole: A broad-spectrum triazole antifungal agent. *Clinical Infectious Diseases*. 2007;45(12):1610-7.
- Nagel R, Cuttall L, Stensvold CR, Mills PC, Bielefeldt-Ohmann H, Traub RJ. Blastocystis subtypes in

- symptomatic and asymptomatic family members and pets and response to therapy. *Internal Medicine Journal*. 2012;42(11):1187-95.
- Nahmias C, Wahl L, Chirakal R, Firnau G, Garnett ES. A probe for intracerebral aromatic amino-acid decarboxylase activity: Distribution and kinetics of [<sup>18</sup>F]6-fluoro-L-m-tyrosine in the human brain. *Movement Disorders*. 1995;10(3):298-304.
- Nordgaard I, Hove H, Clausen MR, Mortensen PB. Colonic production of butyrate in patients with previous colonic cancer during long-term treatment with dietary fibre (*Plantago ovata* seeds). *Scandinavian Journal of Gastroenterology*. 1996;31(10):1011-20.
- Norton C, Chelvanayagam S, Wilson-Barnett J, Redfern S, Kamm MA. Randomized Controlled Trial of Biofeedback for Fecal Incontinence. *Gastroenterology*. 2003;125(5):1320-9.
- Obradovic V, Artiko V, Radevic B, Dapcevic B, Petrovic N. Single injection hepatic radionuclide angiography and hepatobiliary scintigraphy in the evaluation of liver transplant function. *Nucl Med Rev Cent East Eur*. 2004;7(1):21-5.
- O'Connell MJ, Schutt AJ, Moertel CG, Rubin J, Hahn RG, Scott M. A randomized clinical trial of combination chemotherapy in advanced colorectal cancer. *American journal of clinical oncology*. 1987;10(4):320-4.
- Ogholikhan S, Franciscovich A, Mogul D. PoopMD, a mobile application to screen for biliary atresia, accurately identifies Acholic Stools in the Field. *Hepatology*. 2016;64 (1 Supplement 1):152A.
- Oh Y, Ha Y, Han K, Seo HW, Kang I, Park C, et al. Expression of Leucocyte Function-associated Antigen-1 and Intercellular Adhesion Molecule-1 in the Lungs of Pigs Infected with *Actinobacillus pleuropneumoniae*. *Journal of Comparative Pathology*. 2013;148(2-3):259-65.
- Orlowska E, Czubkowski P, Motyl I, Klewicka E, Libudzisz Z, Socha P. The clinical effect and changes of microflora under probiotic supplementation in children with biliary atresia-a randomized controlled trial. *United European Gastroenterology Journal*. 2015;1):A349.
- Orr DW, Myint H, Murphy R. Probiotic supplementation after Very Low Calorie Diet does not aid improvement of the metabolic syndrome or maintenance of weight loss post Liver Transplant. A randomised double-blind placebo controlled trial. *Hepatology*. 2016;64 (1 Supplement 1):113A-4A.
- Ortiz M, Schnabel K, Teut M, Rotter G, Binting S, Cree M, et al. Complementary and integrative medicine in nursing homes-results of a prospective, exploratory, comparative, two-armed cohort study from the residents' perspective. *BMC Complementary and Alternative Medicine Conference: World Congress Integrative Medicine and Health*. 2017;17(Supplement 1).
- Orvain C, Moles-Moreau MP, Francois S, Mercier M, Moal F, Hamel JF, et al. Miconazole mucoadhesive buccal tablet in high-dose therapy with autologous stem cell transplantation (HDT/ASCT)-induced mucositis. *Supportive Care in Cancer*. 2015;23(2):359-64.
- Palaoro LG, Araujo VP, Matos SL, Alves CLGF, Brito VN, Cunha FS, et al. Effects of long term testosterone administration and gonadectomy on gonadotropin secretion in female to male transsexuals. *Endocrine Reviews Conference: 97th Annual Meeting and Expo of the Endocrine Society, ENDO*. 2015;36(no

1  
2  
3 pagination).

4  
5 Pallotta N, Rubinetto MP, Zaccaro C, Gizzi G, Villani V, Barbara L. Calcium polycarbophil in clinical practice.  
6 Treatment of constipation. [Italian]. *Minerva Gastroenterologica e Dietologica*. 1993;39(4):175-8.

7  
8 Pallotta N, Rubinetto MP, Zaccaro C, Gizzi G, Villani V, Barbara L. [Calcium polycarbophil in clinical practice.  
9 The therapy of constipation]. *Minerva Gastroenterologica e Dietologica*. 1993;39(4):175-8.

10  
11  
12 Palou J, Angulo JC, Ramon De Fata F, Garcia-Tello A, Gonzalez-Enguita C, Boada A, et al. Randomized  
13 comparative study for the assessment of a new therapeutic schedule of fosfomycin trometamol in  
14 postmenopausal women with uncomplicated lower urinary tract infection. [Spanish]. *Actas Urologicas*  
15 *Espanolas*. 2013;37(3):147-55.

16  
17  
18 Palou J, Angulo JC, Ramon de Fata F, Garcia-Tello A, Gonzalez-Enguita C, Boada A, et al. [Randomized  
19 comparative study for the assessment of a new therapeutic schedule of fosfomycin trometamol in  
20 postmenopausal women with uncomplicated lower urinary tract infection]. *Actas Urologicas Espanolas*.  
21 2013;37(3):147-55.

22  
23  
24 Paramsothy S, Borody T, Lin E, Finlayson S, Walsh A, Samuel D, et al. Obstacles to donor recruitment for  
25 faecal microbiota transplantation: Experiences from the focus study. *American Journal of Gastroenterology*.  
26 2014;109:S188.

27  
28  
29 Paramsothy S, Borody T, Lin E, Finlayson S, Walsh A, Samuel D, et al. Obstacles to donor recruitment for  
30 faecal microbiota transplantation-Experiences from the FOCUS study. *Journal of Gastroenterology and*  
31 *Hepatology (Australia)*. 2014;29:135.

32  
33  
34 Paramsothy S, Borody TJ, Lin E, Finlayson S, Walsh AJ, Samuel D, et al. Donor Recruitment for Fecal  
35 Microbiota Transplantation. *Inflammatory Bowel Diseases*. 2015;21(7):1600-6.

36  
37  
38 Paramsothy S, Kaakoush NO, Kamm MA, Faith JJ, Clemente JC, Walsh AJ, et al. Faecal microbiota  
39 transplantation (FMT) in ulcerative colitis is associated with specific bacterial changes: Stool and colonic  
40 mucosa 16S microbiota analysis from the randomised controlled FOCUS study. *Journal of Gastroenterology*  
41 *and Hepatology (Australia)*. 2016;31:125-6.

42  
43  
44 Park EJ, Kang J, Baik SH. Treatment of Faecal incontinence using allogeneic-adipose-derived mesenchymal  
45 stem cells: A study protocol for a pilot randomised controlled trial. *BMJ Open*. 2016;6 (2) (no  
46 pagination)(e010450).

47  
48  
49 Parnetti L. Clinical pharmacokinetics of drugs for Alzheimer's disease. *Clinical Pharmacokinetics*.  
50 1995;29(2):110-29.

51  
52  
53 Paul D, Gokarn AG, Bhat V, Bonda A, Zanwar S, Mathew L, et al. Impact of surveillance stool culture guided  
54 selection of antibiotics in allogeneic hematopoietic stem cell transplant patients. *Blood Conference: 58th*  
55 *Annual Meeting of the American Society of Hematology, ASH*. 2016;128(22).

56  
57  
58 Pawlowska J, Klewicka E, Czubkowski P, Motyl I, Jankowska I, Libudzisz Z, et al. Effect of *Lactobacillus casei*  
59 DN-114001 Application on the Activity of Fecal Enzymes in Children After Liver Transplantation.  
60 *Transplantation Proceedings*. 2007;39(10):3219-21.

- Peng Z, Xiang J, He Z, Zhang T, Xu L, Cui B, et al. Colonic transendoscopic enteral tubing: A novel way of transplanting fecal microbiota. *Endoscopy International Open*. 2016;4(6):E610-E3.
- Perlick DA, Miklowitz DJ, Lopez N, Chou J, Calvin C, Adzhiasvili V, et al. Family-focused treatment for caregivers of patients with bipolar disorder. *Bipolar Disord*. 2010;12(6):627-37.
- Persson GR, Samuelsson E, Lindahl C, Renvert S. Mechanical non-surgical treatment of peri-implantitis: A single-blinded randomized longitudinal clinical study. II. Microbiological results. *Journal of Clinical Periodontology*. 2010;37(6):563-73.
- Pertile G, Claes C. Macular translocation with 360 degree retinotomy for management of age-related macular degeneration with subfoveal choroidal neovascularization. *American Journal of Ophthalmology*. 2002;134(4):560-5.
- Pfundstein J, Roghmann MC, Schwalbe RS, Qaiyumi SQ, McCarter Jr RJ, Keay S, et al. A randomized trial of surgical antimicrobial prophylaxis with and without vancomycin in organ transplant patients. *Clinical Transplantation*. 1999;13(3):245-52.
- Pfundstein J, Roghmann MC, Schwalbe RS, Qaiyumi SQ, McCarter RJ, Jr., Keay S, et al. A randomized trial of surgical antimicrobial prophylaxis with and without vancomycin in organ transplant patients. *Clinical Transplantation*. 1999;13(3):245-52.
- Philips CA, Shasthry SM, Pande A, Jamwal KD, Chandel SS, Kumar G, et al. Fecal microbiota transplantation (FMT) improves outcome and survival in steroid ineligible severe alcoholic hepatitis-A randomized control trial (NCT 02458079). *Hepatology*. 2016;64 (1 Supplement 1):706A.
- Philips CA, Shasthry SM, Pande A, Jamwal KD, Khillan V, Hussain MS, et al. Outcomes of fecal microbiota transplantation in steroid ineligible severe alcoholic hepatitis-A randomized control trial (NCT02458079). *Indian Journal of Gastroenterology*. 2016;35 (1 Supplement):A68-A9.
- Philpott-Howard JN, Wade JJ, Mufti GJ, Brammer KW, Ehninger G. Randomized comparison of oral fluconazole versus oral polyenes for the prevention of fungal infection in patients at risk of neutropenia. Multicentre Study Group. *Journal of Antimicrobial Chemotherapy*. 1993;31(6):973-84.
- Philpott-Howard JN, Wade JJ, Mufti GJ, Brammer KW, Ehninger G, Tanzer J, et al. Randomized comparison of oral fluconazole versus oral polyenes for the prevention of fungal infection in patients at risk of neutropenia. *Journal of Antimicrobial Chemotherapy*. 1993;31(6):973-84.
- Pipkin KM, Hagey JV, Rayburn MC, Chigerwe M. A randomized clinical trial evaluating metabolism of colostral and plasma derived immunoglobulin G in Jersey bull calves. *Journal of veterinary internal medicine / American College of Veterinary Internal Medicine*. 2015;29(3):961-6.
- Price CE, Cox S, Rode H. The use of diverting colostomies in paediatric peri-anal burns: Experience in 45 patients. *South African Journal of Surgery*. 2013;51(3):102-5.
- Pulungsih SP, Punjabi NH, Rafli K, Rifajati A, Kumala S, Simanjuntak CH, et al. Standard WHO-ORS versus reduced-osmolarity ORS in the management of cholera patients. *Journal of Health, Population and Nutrition*.



2006;24(1):107-12.

Qian C, Decot V, Wang Y, Cai HL, Venard V, Jeulin H, et al. Adoptive immunotherapy of refractory systemic adenovirus infections after allogeneic umbilical cord blood (UCB) or peripheral blood stem cell transplantation. *Bone Marrow Transplantation*. 2015;50:S120.

Queenan KM, Stewart ML, Smith KN, Thomas W, Fulcher RG, Slavin JL. Concentrated oat beta-glucan, a fermentable fiber, lowers serum cholesterol in hypercholesterolemic adults in a randomized controlled trial. *Nutrition Journal*. 2007;6 (no pagination)(6).

Quirke P. Simplicity and complexity-improving outcomes in bowel cancer. *Journal of Pathology*. 2013;231:S6.

Quraishi N, McMillan M, Widlak M, Nell L, Quraishi K, Pathmakanthan S, et al. Patient perception towards faecal microbiota transplantation for treatment of inflammatory bowel disease. *United European Gastroenterology Journal*. 2014;1):A383.

Quraishi N, McMillan M, Widlak M, Nell L, Quraishi K, Pathmakanthan S, et al. Patient perception towards faecal microbiota transplantation for treatment of Inflammatory Bowel Disease. *Journal of Crohn's and Colitis*. 2015;9:S252.

Rams TE, Degener JE, van Winkelhoff AJ. Antibiotic resistance in human peri-implantitis microbiota. *Clinical oral implants research*. 2014;25(1):82-90.

Ranganathan N, Friedman EA, Tam P, Rao V, Ranganathan P, Dheer R. Probiotic dietary supplementation in patients with stage 3 and 4 chronic kidney disease: A 6-month pilot scale trial in Canada. *Current Medical Research and Opinion*. 2009;25(8):1919-30.

Rathmann N, Kara K, Budjan J, Henzler T, Smakic A, Schoenberg SO, et al. Parenchymal liver blood volume and dynamic volume perfusion CT measurements of hepatocellular carcinoma in patients undergoing transarterial chemoembolization. *Anticancer Research*. 2017;37(10):5681-5.

Ratto C, Buntzen S, Aigner F, Altomare DF, Heydari A, Donisi L, et al. Multicentre observational study of the Gatekeeper for faecal incontinence. *The British journal of surgery*. 2016;103(3):290-9.

Rayes A, Morrow AL, Payton LR, Lake KE, Lane A, Davies SM. A Genetic Modifier of the Gut Microbiome Influences the Risk of Graft-versus-Host Disease and Bacteremia After Hematopoietic Stem Cell Transplantation. *Biol Blood Marrow Transplant*. 2016;22(3):418-22.

Rebello D, Yen E, Lio P, Kelly CR. Unexpected benefits: Hair growth in two alopecia patients after fecal microbiota transplant. *American Journal of Gastroenterology*. 2016;111:S623-S4.

Reinhardt K, Foell D, Vogl T, Mezger M, Wittkowski H, Fend F, et al. Monocyte-induced development of Th17 cells and the release of S100 proteins are involved in the pathogenesis of graft-versus-host disease. *J Immunol*. 2014;193(7):3355-65.

Reinshagen M, Stallmach A. Multidonor intensive faecal microbiota transplantation for active ulcerative colitis: A randomised placebo-controlled trial. [German]. *Zeitschrift fur Gastroenterologie*. 2017;55(8):779-80.

- Reintgen D, Pendas S, Jakub J, Swor G, Giuliano R, Bauer J, et al. National trials involving lymphatic mapping for melanoma: The multicenter selective lymphadenectomy trial, the SunBelt melanoma trial, and the Florida melanoma trial. *Seminars in Oncology*. 2004;31(3):363-73.
- Renvert S, Lindahl C, Renvert H, Persson GR. Clinical and microbiological analysis of subjects treated with Branemark or AstraTech implants: A 7-year follow-up study. *Clinical Oral Implants Research*. 2008;19(4):342-7.
- Riko K, Pichora-Fuller MK, Alberti PW. Clinical evaluation of a two-channel amplitude compression hearing aid. *Laryngoscope*. 1986;96(11):1226-30.
- Riley DK, Pavia AT, Beatty PG, Denton D, Carroll KC. Surveillance cultures in bone marrow transplant recipients: Worthwhile or wasteful? *Bone Marrow Transplantation*. 1995;15(3):469-73.
- Robaeys G, Cassiman D, Verslype C, Monbaliu D, Aerts R, Pirenne J, et al. Successful Conversion From Mycophenolate Mofetil to Enteric-Coated Mycophenolate Sodium (Myfortic) in Liver Transplant Patients With Gastrointestinal Side Effects. *Transplantation Proceedings*. 2009;41(2):610-3.
- Rodrigues JFV, Rayo J, Vicente J, Carolino E, Figueiredo S, Vieira L. Influence of the geometry and positioning of the regions of interest in the transplanted renogram. *European Journal of Nuclear Medicine and Molecular Imaging*. 2017;44 (2 Supplement 1):S896-S7.
- Rohrenbach J, Matthess A, Maier R, Von Bunau R. Treatment of children with E. coli strain Nissle 1917. Results of a prospective data collection with 668 patients. [German]. *Padiatrische Praxis*. 2009;73(4):645-52.
- Romaniszyn M, Rozwadowska N, Malcher A, Kolanowski T, Walega P, Kurpisz M. Implantation of autologous muscle-derived stem cells in treatment of fecal incontinence: results of an experimental pilot study. *Techniques in Coloproctology*. 2015;19(11):685-96.
- Romano G, Cocchiara G, Calderone F, Luna E, Virzi C, Agrusa A, et al. [Endoscopic treatment of colorectal polyps in a digestive endoscopy outpatient department]. *Chirurgia Italiana*. 2004;56(5):669-73.
- Rongen MJ, Adang EM, van der Hoop AG, Baeten CG. One-step vs two-step procedure in dynamic graciloplasty. *Colorectal Disease*. 2001;3(1):51-7.
- Rongen MJGM, Adang EMM, Gerritsen van der Hoop A, Baeten CGMI. One step vs two-step procedure in dynamic graciloplasty. *Colorectal Disease*. 2001;3(1):51-7.
- Rossen N, Bart A, Verhaar N, Van Nood E, Kootte R, De Groot P, et al. Low prevalence of blastocystis SP in active ulcerative colitis patients. *Gastroenterology*. 2014;1:S-371.
- Rossen N, Bart A, Verhaar N, Van Nood E, Kootte R, De Groot P, et al. Low prevalence of Blastocystis sp. In active ulcerative colitis patients. *Journal of Crohn's and Colitis*. 2014;8:S349.
- Roth B, Birkhauser FD, Zehnder P, Burkhard FC, Thalmann GN, Studer UE. Readaptation of the peritoneum following extended pelvic lymphadenectomy and cystectomy has a significant beneficial impact on early



- postoperative recovery and complications: Results of a prospective randomized trial. *European Urology*. 2011;59(2):204-10.
- Rump JA, Arndt R, Arnold A, Bendick C, Dichtelmuller H, Franke M, et al. Treatment of diarrhoea in human immunodeficiency virus-infected patients with immunoglobulins from bovine colostrum. *Clin Investig*. 1992;70(7):588-94.
- Runde V, Ross S, Trenchel R, Lagemann E, Basu O, Renzing-Kohler K, et al. Adenoviral infection after allogeneic stem cell transplantation (SCT): report on 130 patients from a single SCT unit involved in a prospective multi center surveillance study. *Bone Marrow Transplantation*. 2001;28(1):51-7.
- Rzepecki P, Barzal J, Oborska S. Blood and marrow transplantation and nutritional support. *Supportive Care in Cancer*. 2010;18(SUPPL. 2):S57-S65.
- Sabharwal S, Abraham JM, Grand R, Mascarenhas M. "ins and outs" of constipation and DIOS. *Pediatric Pulmonology*. 2016;51:147.
- Samy E, Wu Y, Higginbotham G, Grenningloh R, Xu D. The alphav integrin inhibitor abituzumab inhibits myofibroblast differentiation. *Arthritis and Rheumatology Conference: American College of Rheumatology/Association of Rheumatology Health Professionals Annual Scientific Meeting, ACR/ARHP*. 2017;69(Supplement 10).
- Santini B, Antonelli M, Battistini A, Bertasi S, Collura M, Esposito I, et al. Comparison of two enteric coated microsphere preparations in the treatment of pancreatic exocrine insufficiency caused by cystic fibrosis. *Digestive and Liver Disease*. 2000;32(5):406-11.
- Santos JL, Carvalho E, Bezerra JA. Advances in biliary atresia: From patient care to research. *Brazilian Journal of Medical and Biological Research*. 2010;43(6):522-7.
- Santosham M, Burns BA, Reid R, Letson GW, Duncan B, Powlesland JA, et al. Glycine-based oral rehydration solution: reassessment of safety and efficacy. *Journal of Pediatrics*. 1986;109(5):795-801.
- Sanz Y, Santacruz A, Gauffin P. Gut microbiota in obesity and metabolic disorders. *Proceedings of the Nutrition Society*. 2010;69(3):434-41.
- Sarveazad A, Newstead GL, Mirzaei R, Joghataei MT, Bakhtiari M, Babahajian A, et al. A new method for treating fecal incontinence by implanting stem cells derived from human adipose tissue: preliminary findings of a randomized double-blind clinical trial. *Stem Cell Research and Therapy*. 2017;8 (1) (no pagination)(40).
- Sasaki M, Shimozaoto A, Ogasawara N, Funaki Y, Ebi M, Hijikata Y, et al. Transglucosidase improves the bowel movements in type 2 diabetes mellitus patients: A randomized double-blind, placebo-controlled study. *Gastroenterology*. 2017;152 (5 Supplement 1):S1012-S3.
- Scales SJ, de Sauvage FJ. Mechanisms of Hedgehog pathway activation in cancer and implications for therapy. *Trends in Pharmacological Sciences*. 2009;30(6):303-12.
- Schneider HJ, Pickel J, Stalla GK. Typical female 2nd-4th finger length (2D:4D) ratios in male-to-female transsexuals-possible implications for prenatal androgen exposure. *Psychoneuroendocrinology*.

2006;31(2):265-9.

Seekatz AM, Aas J, Gessert CE, Rubin TA, Saman DM, Bakken JS, et al. Recovery of the gut microbiome following fecal microbiota transplantation. *mBio*. 2014;5(3):e00893-14.

Seo Y, Hawkins R, Christine C, Larson P, Bankiewicz K. In vivo quantitative PET/MR imaging of gene expression in Parkinson's Disease. *Journal of Nuclear Medicine Conference: Society of Nuclear Medicine and Molecular Imaging Annual Meeting, SNMMI*. 2015;56(no pagination).

Shah K, Jacobs A, Breakefield XO, Weissleder R. Molecular imaging of gene therapy for cancer. *Gene Therapy*. 2004;11(15):1175-87.

Shimoyama T, Sawaya M, Ishiguro A, Hanabata N, Yoshimura T, Fukuda S. Applicability of a rapid stool antigen test, using monoclonal antibody to catalase, for the management of *Helicobacter pylori* infection. *Journal of Gastroenterology*. 2011;46(4):487-91.

Shiogai T, Koshimura M, Uebo C, Makino M, Mizuno T, Nakajima K. Acetazolamide vasoreactivity in persistent vegetative state and vascular dementia evaluated by transcranial harmonic perfusion imaging and Doppler sonography. *Acta neurochirurgica*. 2003;Supplement. 87:63-9.

Shiogai T, Morisaka A, Takayasu N, Yoshikawa K, Mizuno T, Nakagawa M, et al. Quantitative evaluation of cerebrovascular reactivity in brain tissue by a refill kinetic method of transcranial ultrasonic perfusion imaging: a comparison with Doppler sonography. *Acta neurochirurgica*. 2005;Supplement. 95:183-90.

Shukla A. Those spots on his penis: It is bannayan riley ruvalcaba syndrome!! *Pediatric Dermatology*. 2017;34:S141-S2.

Sidhu SS, Goyal O, Kishore H, Sidhu S. New paradigms in management of alcoholic hepatitis: a review. *Hepatology International*. 2017:1-13.

Simonetti F, Fortunato S, Rousseau M, Tascini C, Menichetti F, Stefanelli A, et al. Oral gentamicin therapy for carbapenem-resistant *klebsiella pneumoniae* infections in hematologic patients: A single center experience. *Haematologica*. 2016;101:485.

Siproudhis L, Morcet J, Laine F. Elastomer implants in faecal incontinence: A blind, randomized placebo-controlled study. *Alimentary Pharmacology and Therapeutics*. 2007;25(9):1125-32.

Smith RC, Lindenmayer JP, Hu Q, Kelly E, Viviano TF, Cornwell J, et al. Effects of olanzapine and risperidone on lipid metabolism in chronic schizophrenic patients with long-term antipsychotic treatment: A randomized five month study. *Schizophrenia Research*. 2010;120(1-3):204-9.

Smith SD, Jackson RJ, Hannakan CJ, Wadowsky RM, Tzakis AG, Rowe MI. Selective decontamination in pediatric liver transplants. A randomized prospective study. *Transplantation*. 1993;55(6):1306-9.

Somsouk M, Vujkovic-Cvijin I, Pao M, Hunt P, McCune M. Safety of fecal microbial transplantation during treated hiv infection. *American Journal of Gastroenterology*. 2015;110:S578-S9.

Spence C, Verleden S, Einarsson G, Yserbyt J, Lee AJ, Van Herck A, et al. Impact of azithromycin on the post-

lung transplant microbiota. *Thorax*. 2017;72 (Supplement 3):A15.

Staikuniene N, Valantinas J, Pukalskas A, Simoliuniene R, Karciauskaite D. The effect of synbiotic and prokinetic on intestinal permeability, endotoxemia and childpugh score in patients with liver cirrhosis: A prospective cohort study. *United European Gastroenterology Journal*. 2016;4 (5 Supplement 1):A536-A7.

Staley C, Kelly CR, Brandt LJ, Khoruts A, Sadowsky MJ. Characterization of fecal microbiota in response to heterologous versus autologous (placebo) fecal microbial transplantation: Results from a dualcenter, randomized, placebo-controlled trial. *Gastroenterology*. 2016;1):S542.

Stockfleth E. Sonidegib tolerability across 30-months: Results from the phase 2 randomized BOLT trial. *Journal of Investigative Dermatology*. 2017;137 (10 Supplement 2):S285.

Stockfleth E. Sonidegib duration of response across 30 months: Results from the randomized phase 2 BOLT trial. *Journal of Investigative Dermatology*. 2017;137 (10 Supplement 2):S284.

Stojkovic M, Stojkovic M, Artiko V, Zuvela M, Lekic N, Petrovic M, et al. Is identification of malignant lesions of the liver and of hemangiomas possible by Doppler ultrasonography and radionuclide angiography? *Hellenic J Nucl Med*. 2011;14(1):38-42.

Storey RF, Bliden KP, Ecob R, Karunakaran A, Butler K, Wei C, et al. Earlier recovery of platelet function after discontinuation of treatment with ticagrelor compared with clopidogrel in patients with high antiplatelet responses. *J Thromb Haemost*. 2011;9(9):1730-7.

Stratigos AJ. Update on systemic treatment of basal cell carcinoma. *Journal of the European Academy of Dermatology and Venereology*. 2017;31:16.

Strik AS, Brandse JF, Koelink P, Wildenberg M, De Vries A, Van Den Brink G, et al. Underestimation of fecal loss of infliximab due to proteolysis. *Gastroenterology*. 2017;152 (5 Supplement 1):S388-S9.

Subramaniam K, Watthayalage R, Neeman T, Pavli P. Is anti-TNF therapeutic drug monitoring of value in IBD patients in clinical remission? *Journal of Gastroenterology and Hepatology (Australia)*. 2016;31:149.

Subramanian S, Huq S, Yatsunenko T, Haque R, Mahfuz M, Alam MA, et al. Persistent gut microbiota immaturity in malnourished Bangladeshi children. *Nature*. 2014;510(7505):417-21.

Sudan D. Small bowel transplantation: Current status and new developments in allograft monitoring. *Current Opinion in Organ Transplantation*. 2005;10(2):124-7.

Sulkowski JP, Nacion KM, Deans KJ, Minneci PC, Levitt MA, Mousa HM, et al. Sacral nerve stimulation: a promising therapy for fecal and urinary incontinence and constipation in children. *J Pediatr Surg*. 2015;50(10):1644-7.

Sunshine A, Mulhern SA, Olson N, Elkind A, Almas M, Sikes C. Comparative sensitivity of stopwatch methodology and conventional pain assessment measures for detecting early response to triptans in migraine: Results of a randomized, open-label pilot study. *Clinical Therapeutics*. 2006;28(8):1107-15.

Suskind DL, Brittnacher MJ, Wahbeh G, Shaffer ML, Hayden HS, Qin X, et al. Fecal microbial transplant effect

- on clinical outcomes and fecal microbiome in active Crohn's disease. *Inflammatory Bowel Diseases*. 2015;21(3):556-63.
- Suzuki H, Watanabe H, Shinozaki T, Yanagawa T, Suzuki R, Takagishi K. Positron emission tomography imaging of musculoskeletal tumors in the shoulder girdle. *Journal of Shoulder and Elbow Surgery*. 2004;13(6):635-47.
- Suzuki R, Watanabe H, Yanagawa T, Sato J, Shinozaki T, Suzuki H, et al. PET evaluation of fatty tumors in the extremity: possibility of using the standardized uptake value (SUV) to differentiate benign tumors from liposarcoma. *Ann Nucl Med*. 2005;19(8):661-70.
- Sylvester FA, Turner D, Draghi A, 2nd, Uuosoe K, McLernon R, Koproske K, et al. Fecal osteoprotegerin may guide the introduction of second-line therapy in hospitalized children with ulcerative colitis. *Inflammatory Bowel Diseases*. 2011;17(8):1726-30.
- Tamandl D, Waneck F, Sieghart W, Unterhumer S, Kolblinger C, Baltzer P, et al. Early response evaluation using CT-perfusion one day after transarterial chemoembolization for HCC predicts treatment response and long-term disease control. *European Journal of Radiology*. 2017;90:73-80.
- Tan JJY, Chan M, Tjandra JJ. Evolving therapy for fecal incontinence. *Diseases of the Colon and Rectum*. 2007;50(11):1950-67.
- Tanpowpong P, Prachasitthisak N, Treepongkaruna S, Lertudomphonwanit C, Boonsathorn S, Angkathunyakul N, et al. Stool cytomegalovirus polymerase chain reaction for the diagnosis of cytomegalovirus causing gastrointestinal disease in immunocompromised children. *Journal of Pediatric Gastroenterology and Nutrition*. 2016;63:S13.
- Thin NN, Taylor SJ, Bremner SA, Emmanuel AV, Hounsborne N, Williams NS, et al. Randomized clinical trial of sacral versus percutaneous tibial nerve stimulation in patients with faecal incontinence. *The British journal of surgery*. 2015;102(4):349-58.
- Thin NN, Taylor SJ, Bremner SA, Emmanuel AV, Hounsborne N, Williams NS, et al. Randomized clinical trial of sacral versus percutaneous tibial nerve stimulation in patients with faecal incontinence. *Br J Surg*. 2015;102(4):349-58.
- Tischer S, Schultze-Florey R, Heim A, Picksak G, Mynarek M, Sauer M, et al. Monitoring of adenovirus-specific T cells after HSCT in children: Equal detection of hexon and penton-specific T cells. *Transfusion Medicine and Hemotherapy*. 2016;43:62.
- Tjellstrom A, Luetje CM, Hough JV, Arthur B, Hertzmann P, Katz B, et al. Acute human trial of the floating mass transducer. *Ear Nose Throat J*. 1997;76(4):204-6.
- Tornatore L, Acton G, Adams N, Campbell EA, Kelly J, Szydlo RM, et al. Cancer-selective targeting of the NF-kappaB survival pathway in multiple myeloma with the GADD45beta/MKK7 inhibitor, DTP3. *Blood*. 2015;126(23):868.
- Totman JJ, O'Gorman R L, Kane PA, Karani JB. Comparison of the hepatic perfusion index measured with gadolinium-enhanced volumetric MRI in controls and in patients with colorectal cancer. *Br J Radiol*.

2005;78(926):105-9.

Tsai F, Coyle WJ. The microbiome and obesity: Is obesity linked to our gut flora? *Current Gastroenterology Reports*. 2009;11(4):307-13.

Tsunashima D, Kawamura A, Murakami M, Sawamoto T, Undre N, Brown M, et al. Assessment of tacrolimus absorption from the human intestinal tract: open-label, randomized, 4-way crossover study. *Clinical Therapeutics*. 2014;36(5):748-59.

Tsunashima D, Kawamura A, Murakami M, Sawamoto T, Undre N, Brown M, et al. Assessment of tacrolimus absorption from the human intestinal tract: Open-label, randomized, 4-way crossover study. *Clinical Therapeutics*. 2014;36(5):748-59.

Turki AT, Basu O, Ditschkowski M, Trenchel R, Beelen DW, Steckel NK. Ileostomy as feasible treatment option for patients with severe refractory graft versus host disease of the gastrointestinal tract after allogeneic stem cell transplantation. *Oncology Research and Treatment*. 2016;39:167.

Valkeinen H, Hakkinen K, Pakarinen A, Hannonen P, Hakkinen A, Airaksinen O, et al. Muscle hypertrophy, strength development, and serum hormones during strength training in elderly women with fibromyalgia. *Scandinavian Journal of Rheumatology*. 2005;34(4):309-14.

Van Beurden YH, Budding AE, Terveer EM, Keller JJ, Kuijper EJ, Vandenbroucke-Grauls CMJE, et al. Fecal microbiota transplantation for patients with post-infectious or antibiotic-induced irritable bowel syndrome: Results from a prospective pilot study. *United European Gastroenterology Journal*. 2017;5 (5 Supplement 1):A566.

van Kraaij MG, Dekker AW, Verdonck LF, van Loon AM, Vinje J, Koopmans MP, et al. Infectious gastroenteritis: an uncommon cause of diarrhoea in adult allogeneic and autologous stem cell transplant recipients. *Bone Marrow Transplantation*. 2000;26(3):299-303.

Varsano I, Eidlitz-Marcus T, Nussinovitch M, Elian I. Comparative efficacy of ceftriaxone and ampicillin for treatment of severe shigellosis in children. *Journal of Pediatrics*. 1991;118(4 I):627-32.

Vaughn BP, Vatanen T, Allegretti JR, Bai A, Xavier RJ, Korzenik J, et al. Increased Intestinal Microbial Diversity Following Fecal Microbiota Transplant for Active Crohn's Disease. *Inflammatory Bowel Diseases*. 2016;22(9):2182-90.

Vlasplolder F, de Zeeuw G, Rozenberg-Arska M, Egyedi P, Verhoef J. The influence of flucloxacillin and amoxicillin with clavulanic acid on the aerobic flora of the alimentary tract. *Infection*. 1987;15(4):241-4.

Vrieze A, Holleman F, Serlie MJ, Ackermans MT, Dallinga-Thie GM, Groen AK, et al. Metabolic effects of transplanting gut microbiota from lean donors to subjects with metabolic syndrome. *Diabetologia*. 2010;53:S44.

Vrieze A, Out C, Fuentes S, Jonker L, Reuling I, Kootte RS, et al. Impact of oral vancomycin on gut microbiota, bile acid metabolism, and insulin sensitivity. *Journal of Hepatology*. 2014;60(4):824-31.

Wagner B. Azathioprine and allopurinol-a deadly combination. *Pharmacotherapy*. 2016;36 (12):e293.

- Wahl LM, Chen JJ, Thompson M, Chirakal R, Nahmias C. The time course of metabolites in human plasma after 6-<sup>18</sup>F-fluoro-L-m-tyrosine administration. *European Journal of Nuclear Medicine*. 1999;26(11):1407-12.
- Wang B, Feng Q, Ye X, Zeng S. The experience and technique in laparoscopic portoenterostomy for biliary atresia. *J Laparoendosc Adv Surg Tech A*. 2014;24(5):350-3.
- Wasserman EI, Hidalgo M, Hornedo J, Cortes-Funes H. Octreotide (SMS 201-995) for hematopoietic support-dependent high-dose chemotherapy (HSD-HDC)-related diarrhoea: Dose finding study and evaluation of efficacy. *Bone Marrow Transplantation*. 1997;20(9):711-4.
- Watanabe H, Inoue T, Shinozaki T, Yanagawa T, Ahmed AR, Tomiyoshi K, et al. PET imaging of musculoskeletal tumours with fluorine-18 alpha-methyltyrosine: Comparison with fluorine-18 fluorodeoxyglucose PET. *European Journal of Nuclear Medicine*. 2000;27(10):1509-17.
- Waugh J, Keating GM, Plosker GL, Easthope S, Robinson DM. Pioglitazone: A review of its use in type 2 diabetes mellitus. *Drugs*. 2006;66(1):85-109.
- Weber E, Doppelmayr M. Kinesthetic motor imagery training modulates frontal midline theta during imagination of a dart throw. *International Journal of Psychophysiology*. 2016;110:137-45.
- Wei Y, Gong J, Zhu W, Guo D, Gu L, Li N, et al. Fecal microbiota transplantation restores dysbiosis in patients with methicillin resistant *Staphylococcus aureus* enterocolitis. *BMC infectious diseases*. 2015;15:265.
- Wilk CM, Weber I, Rachmuhl C, Seidl K, Burgel AH, Muller AMS, et al. Prevalence and eradication of colonizing *staphylococcus aureus* in patients undergoing allogeneic hematopoietic cell transplantation. *Blood Conference: 58th Annual Meeting of the American Society of Hematology, ASH*. 2016;128(22).
- Wu H, Esteve E, Tremaroli V, Khan MT, Caesar R, Manneras-Holm L, et al. Metformin alters the gut microbiome of individuals with treatment-naïve type 2 diabetes, contributing to the therapeutic effects of the drug. *Nat Med*. 2017;23(7):850-8.
- Xi D, Michail S. Fecal microbiota transplantation in children does not significantly alter body mass index. *Journal of Pediatric Gastroenterology and Nutrition*. 2017;65 (Supplement 2):S73-S4.
- Yahyaoui R, Esteva I, Haro-Mora JJ, Almaraz MC, Morcillo S, Rojo-Martinez G, et al. Effect of long-term administration of cross-sex hormone therapy on serum and urinary uric acid in transsexual persons. *J Clin Endocrinol Metab*. 2008;93(6):2230-3.
- Yang Z, Wang X, Bu C. Fecal microbiota transplant for Crohn's disease: A prospective, randomized study in chinese population. *United European Gastroenterology Journal*. 2017;5 (5 Supplement 1):A112-A3.
- Yao Q, Huang Q, Cao Y, Qian P, Chen H. Porcine interferon-gamma protects swine from foot-and-mouth disease virus (FMDV). *Vet Immunol Immunopathol*. 2008;122(3-4):309-11.
- Ye X, Van JN, Munoz FM, Revell PA, Kozinetz CA, Krance RA, et al. Noroviruses as a Cause of Diarrhea in Immunocompromised Pediatric Hematopoietic Stem Cell and Solid Organ Transplant Recipients. *American*



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Journal of Transplantation. 2015;15(7):1874-81.

Young VB. Treatment with fecal microbiota transplantation: The need for complete methodological reporting for clinical trials. *Annals of Internal Medicine*. 2017;167(1):61-2.

Yu C, Benhammou JN, Goyal D, Oh D, Wang L, Jacobs J, et al. High protein dietary intervention improves body mass index (BMI) and reduces the NAFLD fibrosis score (NFS) in veterans with obesity. *American Journal of Gastroenterology*. 2016;111:S349.

Zaza G, Dalla Gassa A, Granata S, Felis G, Lupo A. Impact of the maintenance immunosuppressive therapy on the fecal microbiome of renal transplant recipients: Comparison between an everolimus-versus a standard tacrolimus-based regimen. *Nephrology Dialysis Transplantation*. 2017;32 (Supplement 3):iii408.

Zhang C, Yin A, Li H, Wang R, Wu G, Shen J, et al. Dietary Modulation of Gut Microbiota Contributes to Alleviation of Both Genetic and Simple Obesity in Children. *EBioMedicine*. 2015;2(8):968-84.

Zhang C, Yin A, Li H, Wang R, Wu G, Shen J, et al. Dietary Modulation of Gut Microbiota Contributes to Alleviation of Both Genetic and Simple Obesity in Children. *EBioMedicine*. 2015;2(8):968-84.

Zhang FM, Wang HG, Wang M, Cui BT, Fan ZN, Ji GZ. Fecal microbiota transplantation for severe enterocolonic fistulizing Crohn's disease. *World Journal of Gastroenterology*. 2013;19(41):7213-6.

Zhang T, Xiang J, Cui B, He Z, Li P, Chen H, et al. Cost-effectiveness analysis of fecal microbiota transplantation for inflammatory bowel disease. *Oncotarget*. 2017;8(51):88894-903.

Zheng Y, Lee J, Masand A, Dadhania D, Thangamani M, Suthanthiran M. Tacrolimus precision medicine: Antibiotics increase intra-patient variability in tacrolimus trough concentrations in kidney transplant recipients. *American Journal of Transplantation*. 2016;16:776.

Zhu J, Zhang F, Zhou J, Li H. Assessment of therapeutic response in Crohn's disease using quantitative dynamic contrast enhanced MRI (DCE-MRI) parameters. *Medicine*. 2017;96(32).

Ziemssen F, Luke M, Bartz-Schmidt KU, Gelissen F. Time-dependent effects on contrast sensitivity, near and distance acuity: Difference in functional parameters? (Prospective, randomized pilot trial of photodynamic therapy versus full macular translocation). *Graefes Archive for Clinical and Experimental Ophthalmology*. 2008;246(5):653-9.



## Appendix E. Peer review

### Healthcare Infection Society

**Consultation – The use of faecal microbiota transplant as treatment for recurrent or refractory *Clostridium difficile* infection and other potential indications: joint British Society of Gastroenterology (BSG) and Healthcare Infection Society (HIS) guidelines.**

**Closing date: 5pm on 18 January 2018**

Organisation	Royal College of General Practitioners
Title (e.g. Dr, Mr, Ms, Prof)	Drs
Name	Clinical Adviser: Kevin Barrett Medical Director: Matthew Hoghton
Job title or role	As above
Address and post code	30 Euston square, London, Nw1 2FB
Telephone number	0203 188 7688
Email address	clinicaladvisers@rcgp.org.uk
<b>Please note:</b> comments will only be accepted electronically on this proforma.	

Please provide comments on the draft guideline on the form below, putting each new comment in a new row. When feeding back, please note the section you are commenting on (for example, section 1 Introduction and line number). If your comment relates to the guideline as a whole then please put 'general'. Add extra rows if required.

Section	Comments	Working group response
A	This is an important consultation of an important treatment for recurrent or refractory CDI. The recommendations are sensible and will help produce a universal service to patients across the UK.	Thank you for your comment.
B	Hudson et al doi: 10.1128/CMR.00049-16Clin. Microbiol. Rev. January 2017 vol. 30 no. 1 191-2311 January 2017 review suggests that faecal microbiotca transplant in the United States is used not only in refractory or recurrent Clostridium Difficile (CDI) but also in initial CDI and Ulcerative colitis	We cannot find mention of FMT use as treatment for initial CDI in this review. Updated searches have identified a small RCT evaluating the use of FMT as treatment for first CDI (Camacho-Ortiz <i>et al</i> , 2017), and this is now evaluated by the working group within the guideline ( <b>Section 8.1.1.3</b> ). All published RCTs evaluating the use of the FMT as treatment for ulcerative colitis have been reviewed by the working group within the guideline ( <b>Section 8.6.2</b> ).
C	There is a lack of GP representation on the working group (5.6) and this is reflected in the consultation with a lack of a suggested referral pathway for community based patients	We agree that the implications of this guideline for primary care were not well-described, and we have strengthened this within the guideline. In particular, we have more strongly highlighted the responsibility of microbiology staff in clinical laboratories to liaise proactively with primary care teams regarding the possibility of FMT when recurrent positive stool samples are received from the community on a particular patient ( <b>Section 8.7.1</b> ).
D	There has also been a reported case of the development of obesity following FMT from an overweight donor but this has not been substantiated in other studies. The BMI restriction on donors (8.3.2) may restrict donors.	The recruitment of suitable donors is relatively restrictive by necessity since FMT is an unlicensed and poorly-studied medicinal product. There is a growing literature base demonstrating an association between a high or low BMI and perturbation of the structure and/or function of the gut microbiota and subclinical chronic inflammation. The implications of this for the safety and efficacy of FMT are not well-defined. The suggested BMI range does not make it prohibitively difficult to find suitable donors. As such, the working group believes that their existing recommendation is reasonable.

Section	Comments	Working group response
E	It would be useful to have a standard UK pre and post questionnaire for patients to standardise recording (8.1.2.3)	We agree that the introduction of standardised questionnaires would have clear potential advantages for clinical care and/ or research. We now discuss this further in <b>Section 10</b> , 'further research'.
F	It may useful to consider measuring the microbiol strains of donors to monitor the impact of combinations of specific microbial strains to understand the undefined nature of faecal preparations	We agree of the importance of this, and this is now discussed in more detail in <b>Section 10</b> , 'further research'.
G	The lack of universal definitions of cures (8.1.2.4) is likely to hamper future studies	We agree with this comment. <b>Section 10</b> , 'further research' has been amended accordingly. Furthermore, we expect that the attention generated by this guideline will highlight this inadequacy.
H	With the introduction of the clinical term SNOMECT across primary care in 2018 and secondary care in 2020 it is important to record faecal microbiota transplant so that long term sequaelae can be measured and patients can be potentially contacted in the future.	We agree that there should be specific procedure codes for FMT (according to route of administration), so that this can be accurately recorded in the patient's medical record. This would also lay the foundation for a future HRG code and tariff for the procedure which is not currently funded by CCGs. Members of the working group are in discussion with NHS England about this.

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Healthcare Infection Society

**Consultation – The use of faecal microbiota transplant as treatment for recurrent or refractory *Clostridium difficile* infection and other potential indications: joint British Society of Gastroenterology (BSG) and Healthcare Infection Society (HIS) guidelines.**

Closing date: 5pm on January 2018

Organisation	NHS Highland
Title (e.g. Dr, Mr, Ms, Prof)	Dr
Name	Alex Cochrane
Job title or role	Consultant Microbiology and Infectious Diseases
Address and post code	Raigmore Hospital, Perth Road, Inverness IV2 3UJ
Telephone number	01463 704000
Email address	Alexandra.cochrane@nhs.net
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Section	Comments	Working group response
8.1.1.1	I dont think you should limit FMT for first recurrence to those with specific risk factors. If clinicians wish to use FMT rather than fidaxomicin for the first recurrence on cost effectiveness grounds then that is reasonable. Suggest that you recommend FMT may be offered for the first or second or subsequent recurrences.	As FMT is currently an unlicensed medicinal product with poorly-studied long term sequelae, the working group considered that it should generally be reserved for patients who have had more than three episodes of infection. There are no studies directly comparing its effectiveness with some of the newer agents such as fidaxomicin or bezlotoxumab, hence this recommendation is made on the basis of safety. However, the working group felt that it may be reasonable in certain patient groups (with ongoing risk factors for further

Section	Comments	Working group response
		recurrence) to offer FMT after the second episode. Cost effectiveness analysis was outside the remit of the working group.
8.1.1.3 (ii)	I disagree that patients should have previously been treated with extended/pulsed vancomycin or fidaxomicin before being offered FMT. You dont present any evidence to show that these antibiotic treatment is superior to FMT. Where FMT is the preferred treatment for the first recurrence it is quite likely that the patient will not have had a prolonged or tapered course, and this should not be a barrier to giving FMT which as you say is highly efficacious.	As above, there are no studies comparing FMT to fidaxomicin or bezlotoxumab, and only one study comparing a vancomycin taper to FMT (Hota <i>et al</i> , 2017). The safety profile of these medications is well-established from large randomised controlled trials, whilst randomised studies involving FMT have tended to be smaller, and have more variable patient follow-up. As such, on the balance of safety, the working group agreed that antimicrobial/antitoxin therapy associated with reduced CDI recurrence should be considered prior to FMT. Reflecting the uncertainties in this area within the reviewed literature, the relevant recommendation is 'conditional' rather than 'strong'.
8.1.1.3 (iii)	You dont cite any evidence that fidaxomicin or bezlotoxumab have better cure rates than FMT. My practice has been not to use fidaxomicin in life threatening C. difficile due to lack of evidence of efficacy in this setting, though I may be out of date with this.	Pre-planned subgroup analysis of patients with severe CDI in a randomised trial demonstrated a significantly lower recurrence rate when treated with fidaxomicin (13.0%, $n=12/92$ ) than when treated with vancomycin (26.6%, $n=29/209$ ) (Louie <i>et al</i> , 2011); this finding was replicated in another randomised controlled trial, with 8.3% ( $n=4/48$ ) and 32.6% ( $n=14/43$ ) experiencing a recurrence respectively (Cornely <i>et al</i> , 2012). In a further randomised trial, bezlotoxumab (together with standard of care antibiotics) was shown to reduce recurrence of severe CDI compared to standard of care antibiotics alone (10.9% ( $n=6/55$ ) vs 20% ( $n=13/65$ ) respectively) (Wilcox <i>et al</i> , 2017).  The working group noted that there are no studies comparing FMT to fidaxomicin or bezlotoxumab, and only one study comparing a vancomycin taper to FMT (Hota <i>et al</i> , 2017). The working group agreed that in the absence of this evidence, on

Section	Comments	Working group response
		the balance of safety and potential risks, consideration should be given to using antimicrobial/ antitoxin therapy associated with reduced CDI recurrence prior to considering the use of FMT.
8.5.1.1 (iii)	Is there adequate published material or experience to ensure the safety of loperamide? It is usually avoided in C. difficile disease due To increased risk of complications.	We agree that loperamide should not be used expressly for the treatment of CDI diarrhoea. However, a number of studies (references within the guideline) have used a single dose of loperamide after lower GI FMT to retention, and no potential safety issues associated with this use have been identified.

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**Healthcare Infection Society**

**Consultation – The use of faecal microbiota transplant as treatment for recurrent or refractory *Clostridium difficile* infection and other potential indications: joint British Society of Gastroenterology (BSG) and Healthcare Infection Society (HIS) guidelines.**

**Closing date: 5pm on January 2018**

Organisation	NHS Lothian
Title (e.g. Dr, Mr, Ms, Prof)	Dr
Name	Ewan Olson

Job title or role	Consultant Microbiologist
Address and post code	Royal Infirmary of Edinburgh 51 Little France Crescent Edinburgh EH16 4SA
Telephone number	0131 2326048
Email address	ewan.olson@luht.scot.nhs.uk
<b>Please note:</b> comments will only be accepted electronically on this proforma.	

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Section	Comments	Working group response
8.3.4.	<p>Laboratory Screening of donors</p> <p>"Whilst vancomycin-resistant <i>Enterococci</i> (VRE) carriage is relatively common in the community, they are of low pathogenicity, and screening for them was not felt to be justified."</p> <p>VRE can cause life threatening infections that are difficult to treat. Any patient who is VRE positive requires isolation in a sideroom with ensuite facilities.</p> <p>I would suggest that donors should be screened for VRE before accepting stool for donation. If there is a shortage of donor patients</p>	<p>Whilst vancomycin-resistant <i>Enterococci</i> (VRE) carriage is relatively common in the community (probably related to food consumption) (Endtz <i>et al</i>, 1997), community strains of VRE are genetically distinct from (and generally of much lower pathogenicity than) those found nosocomially (Willems <i>et al</i>, 2005); as such, the working group felt that routine screening was not justified. However, the working group acknowledged that the potential infection risk from VRE (and MRSA) would vary regionally dependent upon local prevalence and pathogenicity, and as such recommended that a risk assessment was performed to assess whether screening for these organisms should be considered.</p>



Section	Comments	Working group response
	should be offered VRE positive donations only with informed consent.	

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Healthcare Infection Society

**Consultation – The use of faecal microbiota transplant as treatment for recurrent or refractory *Clostridium difficile* infection and other potential indications: joint British Society of Gastroenterology (BSG) and Healthcare Infection Society (HIS) guidelines.**

Organisation	On behalf of European Study Group for <i>C. difficile</i> (ESGCD), and the National Donor Feces Bank at Leiden University Medical Center (drs. E. Terveer, drs. E. Boeijs-Koppenol, prof. Hein Verspaget, dr. Y van Beurden, drs. R Ooijevaar, dr. Josbert Keller) and Department of Infectious Diseases, University of Koln (dr. Maria Vehreschild).
Title (e.g. Dr, Mr, Ms, Prof)	Prof. Dr.
Name	Ed Kuijper
Job title or role	Head of Experimental Bacteriology
Address and post code	LUMC, Albinusdreef2, 2333 ZA, Leiden
Telephone number	31-71-5263574
Email address	<a href="mailto:e.i.kuijper@lumc.nl">e.i.kuijper@lumc.nl</a>

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Section	Comments	Working group response
general	The literature was searched until April 2017, but please use the recently published document of E.M Terveer et al. entitled "How to: Establish and run a stool bank" and published in Clin Microbiol Infect. 2017 Dec;23(12):924-930. This document has considerable overlap with the proposed guideline, but also shows some important unresolved issues.	This reference has been added. Literature searches have been updated, to January 2018.
Lay summary, line 3	Capsules may also be prepared by use of non-freeze dried microbiota. Also, the possibility of using frozen products in general may be mentioned in this sentence.	We agree that these changes are important, and these amendments have been accordingly.
8.1.1.1	The authors are correct that CDI due to Type 07 responds less to FMT compared with CDI due to other PCR ribotypes. We register all infections by PCR ribotype to obtain more insights in successes and failures associated with strain characteristics and think that this is relevant for future recommendations, such as repeated FMT treatments for specific PCR ribotypes.	We presume that this refers to ribotype 027, and agree that this is important, and further reference has been made to this in <b>Section 10</b> , further research.
recommendation	"FMT should be offered to patients with recurrent CDI who have had at least two recurrences, or those who have had one recurrence and have risk factors for further episodes, including severe CDI (strong)." Please elucidate how this risk assessment can be performed.	The working party noted that these risk factors are well-described in previous studies, and do not require further elucidation within the manuscript.

Section	Comments	Working group response
8.1.1.2	Refractory CDI is also considered as an indication for FMT. Can the authors please provide a recommendation on the number of FMTs that should be used? Are patients on Intensive Care Units with refractory CDI also eligible? in 8.2.1 IC admission can be considered as a contraindication, but there are sufficient publications supporting to apply it for patients with severe CDI at ICU.	<p>In <b>Section 8.2.1</b>, the working group reviewed the literature on contraindications to receiving FMT, and noted that certain studies have made ‘admission to Intensive Care’ such a contraindication. However, the working group have not themselves at any point stated that this is a contraindication to receiving FMT.</p> <p>As stated in <b>Section 8.1.1.2</b>, there are a relatively small number of cases reported in the reviewed literature of refractory CDI. As such, the working group are unable to give recommendations that patients with refractory CDI receiving FMT should be managed in any particular way differently to those with recurrent CDI.</p>
8.1.1.3	Antibiotic treatment of rCDI. Though the literature search was until April 2017, please mention the recent trials of tapered doses of vancomycin and fidaxomicin (PMID 29273269, PMID: 28591789; PMID 29255732).	We agree that these trials are all relevant, and have updated the guideline accordingly.
	Recommendation II is less clear. How have the authors interpreted the literature that a tapered dosage of vancomycin before FMT increases the success rate of FMT? Are these studies also available for fidaxomicin?	There are no studies comparing FMT to fidaxomicin or bezlotoxumab, and only one study comparing a vancomycin taper to FMT (Hota <i>et al</i> , 2017). The safety profile of these medications is well-established from large randomised controlled trials, whilst randomised studies involving FMT have tended to be smaller, and have more variable patient follow-up. Furthermore, FMT remains (in the UK) an unlicensed medicine. As such, on the balance of safety, the working group agreed that antimicrobial/ antitoxin therapy associated with reduced CDI recurrence should be considered prior to FMT. Reflecting the uncertainties in this area within the reviewed literature, the relevant recommendation is ‘conditional’ rather than ‘strong’.

Section	Comments	Working group response
	Recommendation iii is difficult to understand; do the authors recommend to treat severe and complicated CDI not with vancomycin, but with fidaxomicin or vanco+bezlo? If a recurrence occurs, then followed by a FMT?	The wording of this recommendation has been amended, along with expansion of the explanatory text of <b>Section 8.1.1.4</b> .
	A recommendation for FMT treatment in severe (refractory), complicated CDI is missing (e.g. multiple sequential FMTs); should this also be accompanied with anti-CDI antibiotics? See review v. Beurden, Ther Advances in Gast, 2017 and Fischer, Ali Pharm Ther 2015	As stated in <b>Section 8.1.1.2</b> , there are a relatively small number of cases reported in the reviewed literature of refractory CDI. As such, the working group are unable to give recommendations that patients with refractory CDI receiving FMT should be managed in any particular way differently to those with recurrent CDI.
8.1.2.1	We suggest to differentiate between "non-responding" and "late failure". The latter can be defined as a relapse of CDI after an initial response to FMT. For instance, use of antibiotics in the first month after FMT may provoke a new episode of CDI. This new episode doesn't need a FMT and can be treated with conventional anti-CDI treatment, preferably microbiota sparing such as fidaxomicin.	We agree that this distinction is useful, and have amended the guideline accordingly.
8.1.2.2	Should a psychological questionnaire routinely be taken from recipients (before and after FMT) and from donors (regularly)? A ten-week follow-up is too short to recognize long term side-effects of FMT.	The working group did not consider that this was a priority.
8.1.2.3	We consider swallowing disorders a contraindication for upper GI delivery; death of a patient due to aspiration pneumonia with upper GI delivery has been described (PMID: 29026601); this patient had a swallowing disorder following oropharyngeal radiation after surgical removal of a maxillary carcinoma.	We note that this patient received a very large volume (500ml) of nasoduodenal FMT. This guideline recommends a much lower maximum volume with the specific aim of minimising this problem. Nevertheless, we agree that this is an important consideration, and have amended <b>Section 8.1.2.3</b> and <b>Section 8.5.2.2</b> accordingly.
8.2.1	What is the advice of the committee for coeliac patients with recurrent CDI?	The working group did not have any specific advice regarding patients with coeliac disease.
8.2.2	FMT in immunocompromised patients: we think that the presence of neutropenia ( $<0.5 \times 10^9/L$ ) can be considered as a contraindication for FMT, especially if hematological patients are treated with	The working group have recommended that FMT is offered 'with caution' to immunosuppressed patients, reflecting the careful individualised assessment required for each patient.

Section	Comments	Working group response
	selective gut decontamination to prevent translocation and infections with aerobe Gram-negatives. Second, should donors and immunocompromised recipients be matched for the EBV and CMV status to prevent a herpesvirus infection?	We agree with the comment regarding matching donors and immunosuppressed recipients for EBV and CMV status, and have updated <b>Section 8.2.2</b> and <b>Section 8.3.4</b> accordingly.
8.2.3	The effect of FMT on the IBD status for IBD patients with rCDI is under discussion. Is it possible that FMT will result in cure of CDI but an exacerbation of IBD. Should we differentiate UC from CD? Ref 71 suggests that IBD can worsen. The recommendation "strong" is debatable. Is the IBD group not a better candidate for vancomycin tapering, fidaxomicin (tapering) or bezlotoxumab before FMT is given?	We agree that there is evidence that FMT to treat CDI in patients with IBD may be associated with a flare of IBD activity (Qazi <i>et al</i> , 2017); we have updated the recommendation accordingly.
8.3.2	Age and BMI of the donor. We agree with the BMI of the donor but have some difficulties with the age, We consider an age above 50 as a contraindication, based on the risks to develop colon carcinoma and metabolic (diabetes) diseases. Additionally, older people seems to have a less stable gut microbiota.	We note from a recent paper that <i>Bacteroides: Firmicutes</i> ratio and microbial diversity were similar in donors > 60 years compared to younger donors, and donations from older donors had similar efficacy and no higher rate of adverse outcomes (Anand <i>et al</i> , 2017). As such, the working group agreed to uphold their prior recommendation.
8.3.3.	Donor screening history. Donors should also undergo a long term follow-up to recognize microbiota related diseases, including colon malignancies, autoimmune diseases, metabolic diseases and psychiatric illnesses.	We agree with the principle of this statement, and allude to this in <b>Section 8.7.7</b> .
	Please consider to add to the recommendation/evidence: Potential donors should be extensively screened by a questionnaire and a personal interview concerning risk factors for transmissible diseases and factors influencing the intestinal microbiota	We agree with this suggestion, and have amended <b>Section 8.3.3</b> accordingly.
8.3.4	Screening of the donor. Table 4. The Dutch guideline advises screening donors for multi-drug resistant bacteria (MDR), including VRE, MRSA, CPE and ESBL-producing Gram-negatives, and quinolone/aminoglycoside resistant Enterobacteriaceae. Most of the patients with rCDI have much comorbidity and are frequently hospitalized or encounter nosocomially acquired infections, such as	<p>The working group reviewed their recommendation regarding screening for multi-drug resistant bacteria, and <b>Section 8.3.4</b> has been updated accordingly.</p> <p>We agree with the principle of a ‘window period’/ quarantine prior to repeat donor screening in centres using frozen FMT;</p>

Section	Comments	Working group response
	<p>UTI. Infections with MDR are more difficult to treat, mostly with intravenously administered antibiotics. If these patients become colonized with MDR they should be nursed with specific infection control precautions. We also apply a "window period"; donors stools samples are stored in quarantine for 2 months and only become available after a negative second screening.</p> <p>We additionally screen for: <i>Yersinia enterocolitica</i>, <i>Yersinia pseudotuberculosis</i>, <i>Plesiomonas shigelloides</i>, shiga toxin producing <i>E. coli</i> (not only 0157 <i>E. coli</i>), Astrovirus, Sapovirus, Adenovirus, Enterovirus, Parechovirus, Hepatitis E, <i>Entamoeba histolytica</i>, <i>Microsporidium</i> species, <i>Blastocystis hominis</i>, <i>Dientamoeba fragilis</i>, and <i>Strongyloides</i> (if a travel history to Middle and South America, Africa, or Asia is present).</p> <p>We advise to include carriership of <i>E. histolytica</i> and <i>Strongyloides</i> to the mandatory screening, because of the serious infections that occur in immunocompromised patients. We have detected unexpectedly a donor carrying <i>E. histolytica</i> (Terveer, CMI, 2017).</p>	<p><b>Section 8.3.5</b> has been updated accordingly, and a new flow chart to illustrate the process (<b>Figure 1</b>) added.</p> <p>The working group agreed that recommendations should be made to test for Shiga toxin-producing <i>Escherichia coli</i>, hepatitis E IgM, <i>Entamoeba histolytica</i> serology and <i>Strongyloides stercoralis</i> IgG (<b>Table 3</b>). However, the working group consensus was that screening with the other tests suggested is not justified.</p>
8.4.1	<p>Recommendation i. Please elucidate how donors should deliver their stools. We favour the use of specific device systems to prevent contamination with environmental microorganisms.</p> <p>Recommendation ii. Processing within 6 hours is proven effective, consider changing 'conditional' to 'strong' recommendation</p> <p>Recommendation iii. A meta-analysis concludes that less than 50 gram of feces is related to a 4-fold increase in recurrence rates. The recommendation status should be changed to 'strong'.</p>	<p>i. We think that the text as it stands gives sufficient information about best practice in this area.</p> <p>ii. We agree with this suggestion, and have amended <b>Section 8.4.1</b> accordingly.</p> <p>iii. We agree with this suggestion, and have amended <b>Section 8.4.1</b> accordingly.</p>

Section	Comments	Working group response
8.4.2	An important advantage of frozen FMT is the possibility to use a “window period” of, for example, two months. When donors are screened after this window period, the results determine if the stored FMTs can be used.	We have cross-referenced <b>Section 8.4.2</b> to <b>Section 8.3.5</b> , where the concept of a window period/ quarantine is discussed in more detail.
8.4.3	We think that there is not enough evidence to state that feces suspensions can only be used up to six months from preparation. There is no sufficient data that show a decreased efficacy with feces suspensions stored over 6 months. Additionally, multiple stool banks set the expiration date at 1 year after storage.	A trend towards decrease in the viability of certain gut bacterial groups was noted when faecal aliquots were frozen in 10% glycerol for six months (Costello <i>et al</i> , <i>Alimentary Pharm &amp; Ther</i> , 2015), and as such, the working group agreed that six months was the acceptable limit for freezing of an FMT in glycerol. This rationale is now within the text.
	Good practice point: Thawing overnight in a 4C refrigerator is also a good and much used alternative.	None of the working group had sufficient experience with this means of thawing FMT, and as such were unable to make this good practice point.
8.5.1.1.	It is not clear, why the administration of a bowel lavage in upper GI administration, of PPI, of loperamide and of metoclopramide are recommended. There is no evidence to support their use, and all of them are drugs with known side effects. The only reason why they are used is that the first RCT used them. However, the RCT did not assess their importance, and there are many case series showing that FMT has a high success rate even without their use.	All of these interventions have a clear biological or practical rationale for their use. Significant side effects in association with a single dose of these medications are generally rare, and their use has not been associated with adverse outcomes in FMT studies. Our recommendations for their use are only conditional. As such, the working group uphold their recommendations.
8.5.2.1.	Not all capsules necessarily contain lyophilized microbiota, frozen preparations have also been shown to be effective.	We agree with this comment, and have updated the guideline accordingly.
8.5.2.2	Are there studies indicating that 50 ml for upper gastrointestinal have comparable efficacy as 250 ml? If not, this should be more pronounced mentioned, also in the research session. We use at least 50 gram suspended in 200 ml and a slow infusion of 10cc/min.	As described in the text, the working group considered that mass of stool was a more important consideration than volume of diluent. They also noted that as low as 25ml of FMT has been demonstrated to be effective as upper GI FMT (Aas <i>et al</i> , <i>Clin Infect Dis</i> , 2003). However, the working group revised their decision, and now recommend 100ml as the threshold volume for upper GI FMT administration.



Section	Comments	Working group response
8.5.2.4.	The recommendation not to use capsules seems rather strong. It is unlikely that concerning transmission of infection, the risk would differ in any way from other ways of administration. Also, no safety concerns based on endoscopic complications can possibly arise. We would therefore not pronounce a recommendation against use.	We agree with this statement. Of note, whilst the Kao <i>et al</i> , 2017 study (RCT of capsulised vs colonoscopic FMT) was not published at the time of initial searches, it has been identified by updated searches and has now been reviewed by the working group. As such, the guideline has been updated accordingly.
8.6	Consider to add that specific donor microbiota may have better outcomes (e.g. donor B in Moayyedi, gastroenterology, 2015) FMT for other conditions than rCDI. Why have the authors not included the role of FMT to eradicate MDR from the intestinal tract?	Reference to Donor B in this paper has been added to <b>Section 8.6.2.2</b> .  In keeping with NICE methodology, for the consideration of FMT as treatment for non-CDI conditions, only RCTs could be considered. The working group are aware of case studies and case series using FMT to attempt gut decolonisation of multidrug resistant microorganisms. Members of the working party have themselves contributed to the literature in this field. But no RCTs currently exist.
8.6.3.	Consider adding: characterisation of specific CU patient population that would potentially benefit from FMT. "However, recommendations for clinical use for this indication cannot be made until there is clearer evidence of the most appropriate <b>CU patient characteristics</b> , methodology for its preparation, route of delivery, and intensity of administration of FMT"	We agree with this comment, and have updated the guideline accordingly.
8.7.2 and 8.7.4	FMT is considered as a medicinal product under supervision of MHRA and licensing should follow the GMP guidelines. The activities should be performed in a dedicated containment level 2 laboratory with personal protective equipment and a quality assessment system. Does this indicate that FMTs should be prepared under GMP conditions at the Pharmacy Department and not within the Medical Microbiology? Or is this statement too strong?	No. MHRA guidance does not specify where the manufacture should take place. This could be pharmacy, the microbiology laboratory, or another place.
8.7.6	Please consider to add that aliquots of donor FMT materials (and original feces samples) used for patients treatment should be stored,	We agree, and we have updated <b>Sections 6.3</b> and <b>8.7.6</b> accordingly.

Section	Comments	Working group response
	enabling to use these samples when adverse effects after FMT developed. This should also been included in 6.3 (auditing).	
Table 4	PCRs are more sensitive than conventional microscopy and antigen tests for parasites. Second, can the authors please specify the parasites? There is some debate on the significance of Blastocystes spp. and Dientamoeba spp. Why is only E. coli 157 excluded and not other STEC pathogens?	<b>Table 4</b> has been updated to specify Shiga toxin-producing <i>Escherichia coli</i> screening by PCR. The working group did not consider that specific screening for <i>Blastocystis spp</i> or <i>Dientamoeba spp</i> was justified.
<b>Propose to add:</b> Eligibility of patients for FMT	At the NDFB, all requests by the treating physician are evaluated by at least two clinical members of our feces bank board to determine the eligibility of the patient. It is required that patients have a laboratory documented episode of recurrent CDI following at least one course of adequate CDI antibiotic therapy. Recurrent CDI is defined as the re-appearance of diarrhoea ( $\geq 3$ unformed stools per 24 hours for two consecutive days; or $\geq 8$ unformed stools per 48 hours) within eight weeks after cessation of antibiotic therapy in combination with a positive diagnostic test for <i>C. difficile</i> . We strongly recommend a two-stage testing algorithm, as recently advised by the <i>C. difficile</i> working group/ESCMID (ESGCD). Using this algorithm, we reject approximately 20% of all requests for FMT. We would like to add our experience that of 79 candidate patients for FMT, only 75% were considered as suitable candidates for FMT treatment; most rejected requests were patients with underlying IBD who concomitantly carried <i>C. difficile</i> .	Thank you for this comment. Definitions of recurrent CDI are outside of the remit of this working group. Testing is discussed in <b>Section 8.1.1.</b> , where we refer to current ESCMID guidance.
<b>Need for antimicrobial stewardship after FMT (also for 8.5.1.3)</b>	After FMT, we advise that an infectious disease specialist or medical microbiologists should be involved for antibiotic treatment (or prophylaxis) of the patient during the first month after FMT, since 50% of our registered failures were patients who received antibiotics within one month after FMT. Interestingly, all patients responded to conventional anti-CDI treatment and did not need a second FMT. It can be considered to use microbiota sparing fidaxomicin after FMT.	We agree with this comment, and have updated <b>Section 8.5.1.3</b> accordingly.

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### Healthcare Infection Society

**Consultation – The use of faecal microbiota transplant as treatment for recurrent or refractory *Clostridium difficile* infection and other potential indications: joint British Society of Gastroenterology (BSG) and Healthcare Infection Society (HIS) guidelines.**

**Closing date: 5pm on January 2018**

Organisation	OpenBiome
Title (e.g. Dr, Mr, Ms, Prof)	Dr
Name	Majdi Osman
Job title or role	Clinical Program Director, OpenBiome; Visiting Assistant Professor, Harvard Medical School
Address and post code	200 Inner Belt Road, Somerville, MA 02143
Telephone number	+1 (617) 575-2201
Email address	majdi@openbiome.org
<b>Please note:</b> comments will only be accepted electronically on this proforma.	

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Section	Comments	Working group response
8.1.1.1. Recurrent Clostridium difficile infection	<p><b><i>“FMT should be offered to patients with recurrent CDI who have had at least two recurrences, or those who have had one recurrence and have risk factors for further episodes, including severe CDI (strong).”</i></b></p> <p>We agree however for full clarity we would recommend re-wording to:</p> <p><b><i>“FMT should be offered to patients with recurrent CDI who have had at least two recurrences, or those who have had one recurrence and have risk factors for further episodes, including severe and severe-complicated CDI (strong).”</i></b></p>	We agree with this statement, and have updated the guideline accordingly.
8.1.1.2. Refractory Clostridium difficile infection:	<p><b><i>“FMT should be considered in cases of refractory CDI (conditional).”</i></b></p> <p>We agree.</p>	Thank you for this comment.
8.1.1.3. Antimicrobial therapy prior to considering FMT for patients with CDI:	<p><b><i>i. FMT for recurrent CDI should only be considered after failure of antimicrobial anti-C. difficile therapy which has been administered for a minimum of 10 days (conditional).</i></b></p> <p><b><i>ii. Recipients of FMT as treatment for recurrent CDI should have previously been treated with extended/ pulsed vancomycin and/or fidaxomicin (conditional).</i></b></p> <p><b><i>iii. For those with severe or complicated CDI, which appears to be associated with reduced cure rates, consideration should be given to offering patients treatment with medications which are associated with reduced risk of recurrence (e.g. fidaxomicin and bezlotoxumab), before offering FMT (conditional).</i></b></p> <p>We suggest rewording point <i>iii</i>, that recommends fidaxomicin or bezlotoxumab should be offered to patients with severe or</p>	<p>Pre-planned subgroup analysis of patients with severe CDI in a randomised trial demonstrated a significantly lower recurrence rate when treated with fidaxomicin (13.0%, <i>n</i>=12/92) than when treated with vancomycin (26.6%, <i>n</i>=29/209) (Louie <i>et al</i>, 2011); this finding was replicated in another randomised controlled trial, with 8.3% (<i>n</i>=4/48) and 32.6% (<i>n</i>=14/43) experiencing a recurrence respectively (Cornely <i>et al</i>, 2012). In a further randomised trial, bezlotoxumab (together with standard of care antibiotics) was shown to reduce recurrence of severe CDI compared to standard of care antibiotics alone (10.9% (<i>n</i>=6/55) vs 20% (<i>n</i>=13/65) respectively) (Wilcox <i>et al</i>, 2017).</p> <p>The working group noted that there are no studies comparing FMT to fidaxomicin or bezlotoxumab, and only one study</p>

Section	Comments	Working group response
	<p>complicated CDI before FMT. There is little evidence on the role of bezlotuxumab and fidaxomicin in severe or severe-complicated CDI. Although the evidence base is similarly lacking for FMT in severe or severe-complicated disease, there is a growing body of evidence from trials, multiple case series and reports indicating the potential for FMT in this population.</p> <p><b>Bezlotuxumab:</b> The performance of bezlotuxumab has not been evaluated in a severe or severe-complicated population. Results from MODIFY I and II suggest a modest 10% improvement in rates of sustained cure with bezlotuxumab. Importantly, only 15.6% were severe CDI. Based on the modest gains in efficacy and the few severe/severe-complicated patients in the MODIFY trials, we feel that further evidence is required before proposing bezlotuxumab be offered ahead of FMT in this patient population.</p> <p>In comparison, across similar patient populations FMT has demonstrated in several randomized controlled trials reduced risk of recurrence. Based on the available evidence we therefore feel that the statement that bezlotuximab is “associated with reduced risk of recurrence” compared to FMT is not supported by the evidence.</p> <p><b>Fidaxomicin:</b> Similarly, there is a dearth of evidence on the role of fidaxomicin in the severe CDI population. We agree that it has demonstrated superior efficacy compared to vancomycin in the general CDI population. In an RCT comparing extended-pulsed fidaxomicin versus vancomycin for CDI, Guery et al (2017) observed increased recurrence in severe CDI compared to non-severe CDI with an odds ratio 0.57 (95% CI 0.36–0.91) p=0.019. We therefore recommend that fidaxomixin should be offered to patients with severe CDI. However, there is no evidence to suggest that the</p>	<p>comparing a vancomycin taper to FMT (Hota <i>et al</i>, 2017). The working group agreed that in the absence of this evidence, on the balance of safety and potential risks, consideration should be given to using antimicrobial/ antitoxin therapy associated with reduced CDI recurrence prior to considering the use of FMT.</p>

Section	Comments	Working group response
	<p>performance of fidaxomicin would be better than FMT. We acknowledge that access to fidaxomicin is likely to be more timely in settings where FMT is not readily available.</p> <p><b>The role of FMT in severe CDI:</b> In their recent review, Van Beurden et al (2017) reviewed the literature on FMT in severe CDI and found 23 reports (12 case reports; 11 case series) about FMT as treatment for severe or complicated CDI. The patients described (n=200) all had severe or complicated CDI, did not respond to conventional CDI antibiotic treatment and received FMT as last resort treatment. In all studies, patients were treated with (sequential) FMT, whether or not followed by additional antibiotic treatment for CDI. FMT, with or without additional antibiotic CDI treatment, appears to be a promising curative treatment option in patients with severe and complicated CDI who do not respond sufficiently to conventional antibiotic treatment. FMT has been proposed by Fischer et al (2015) as an option utilizing an endoscopic response-guided approach, which may be particularly useful in non-surgical candidates. In an open-label cohort study (n = 17), FMT was delivered by colonoscopy. If pseudomembranes were identified, patients reinitiated oral vancomycin 24 hour after FMT and continued for 5 days. A repeat FMT by colonoscopy was given on day 7. If pseudomembranes persisted, vancomycin was restarted the following day for a 5 days course and a third FMT was offered on day 13. If pseudomembranes were absent during any colonoscopy, no further therapy was initiated. The results were promising with a combined clinical cure rate of 88%.</p> <p>In conclusion, we agree that there is a lack of evidence available to make a strong recommendation on the role of FMT in severe CDI. However, there is insufficient evidence to suggest that fidaxomicin</p>	

Section	Comments	Working group response
	<p>or bezlotuximab would be superior to FMT in this population. On the contrary, the growing pool of experience in using FMT in severe and severe-complicated CDI patients demonstrates that it appears to be generally safe and effective (quality of evidence: 3).</p> <p>We would therefore suggest re-wording point iii to:</p> <p><b><i>iii. For those with severe or complicated CDI, which appears to be associated with reduced cure rates, consideration should be given to offering patients treatment with medications which are associated with reduced risk of recurrence (e.g. fidaxomicin or bezlotuxumab), or offering FMT (conditional).</i></b></p> <p>Fischer M, Sipe BW, Rogers NA, et al. Faecal microbiota transplantation plus selected use of vancomycin for severe-complicated Clostridium difficile infection: description of a protocol with high success rate. Aliment Pharmacol Ther. 2015;42(4):470-476. doi:10.1111/apt.13290.</p> <p>Van Beurden YH, Nieuwdorp M, van de Berg PJEJ, Mulder CJJ, Goorhuis A. Current challenges in the treatment of severe Clostridium difficile infection: early treatment potential of fecal microbiota transplantation. Therapeutic Advances in Gastroenterology. 2017;10(4):373-381. doi:10.1177/1756283X17690480.</p>	
<b>8.1.2.1. Management of FMT failure:</b>	<p><b><i>Further FMT should be offered after initial FMT failure (strong).</i></b></p> <p>We agree.</p>	Thank you for this comment.
<b>8.1.2.2. General approach to follow-up post-FMT:</b>	<p><b><i>All FMT recipients should routinely receive follow-up. Given the relative novelty of FMT and the potential for unexpected sequelae, clinicians should follow-up FMT recipients for long enough to fully</i></b></p>	Thank you for this comment. In light of other comments from the working group and stakeholders, this follow-up period has been adjusted to 'at least eight weeks in total'.



Section	Comments	Working group response
	<i>establish efficacy/ adverse events, and at least ten weeks in total (strong).</i>  We agree.	
8.1.2.3. Management of the FMT recipient:	<i>i. Immediate management after endoscopic administration of FMT should be as per endoscopy unit protocol (strong).</i> <i>ii. Patients should be warned about short term adverse events, in particular the possibility of self-limiting GI symptoms. They should be advised that serious adverse events are rare (strong).</i> <i>iii. After enteral tube administration, patients may have the tube removed and oral water given from 30 minutes post-administration (strong).</i>  We agree.	Thank you for this comment.
8.1.2.4. Definition of cure post-FMT for CDI:	<i>A decision regarding cure/remission from CDI should be recorded during follow-up. However, this has no uniformly-agreed definition, and should be decided on a case-by-case basis (strong).</i>  We agree.	Thank you for this comment.
8.1.2.5. Definition of treatment failure post-FMT for CDI:	<i>Treatment failure/recurrence should be defined on a case-by-case basis. Routine testing for C. difficile toxin after FMT is not recommended, but is appropriate to consider in the case of persistent CDI symptoms/suspected relapse (strong).</i> When testing is to be performed, we would recommend clinicians follow the 2016 European Society of Clinical Microbiology and Infectious Diseases (ESCMID) guidelines for CDI testing, which state that no single commercial test can be used as a stand-alone test for diagnosing CDI, and recommend a 2-step approach (highly sensitive with reflex to highly specific test). These guidelines recommend performing an initial test with a high negative predicative value;	We agree on the use of ESCMID guidelines in CDI testing, and refer to these clearly in <b>Section 8.1.1.1</b> . However, <b>Section 8.1.2.5</b> specifically refers to diagnosing failure post-FMT for CDI rather than initial diagnosis of CDI, and no good uniform definition exists for this. We think that the guidance given, to define treatment failure on a case-by-case basis, is the most fair summary of the current literature on this topic.

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	<p>therefore, if negative, no further testing needs to be done. Specifically, they suggest glutamate dehydrogenase (GDH) EIA or NAAT/PCR testing. Our recommendation is GDH EIA as it is less expensive and has a slightly superior NPV at higher CDI prevalence compared with NAAT/PCR (98 vs 96 at hypothetical CDI prevalence of 50%), and an NPV of 100% at lower CDI prevalence. The second test should be a test with a high positive predictive value, such as EIA for toxin A/B. Obtaining CDI testing at each suspected CDI recurrence and working with institutional laboratories to use an appropriate testing algorithm is a key component to ensuring appropriate patient selection for FMT.</p> <p>As currently worded, the recommendations risk encouraging over testing in a context where patients may develop post-infectious IBS. This concept is highlighted by evidence suggesting that up to 25% of patients referred to an FMT center for “C difficile infection” were found to have an alternative diagnosis, with younger patients being more likely to have a non-CDI diagnosis (Jackson 2016).</p> <p>Jackson M, Olefson S, Machan JT, Kelly CR. A high rate of alternative diagnoses in patients referred for presumed clostridium difficile infection. J Clin Gastroenterol. 2016 Oct;50(9):742-6.</p>	
<b>8.2.1. General approach to co-morbidities and FMT:</b>	<p><b><i>FMT should be offered with caution in patients with decompensated chronic liver disease and should be avoided in those with anaphylactic food allergy (strong).</i></b></p> <p>The authors may want to consider the approach recommended by Allegretti et al (2017). In patients with a severe food allergy, a potential option for FMT could be from a patient identified donor living with the patient (e.g. spouse) who avoids the same allergens.</p>	<p>The working group thought it important to emphasise the ‘good practice point’ that in patients with true anaphylaxis, the risks of FMT administration were likely to outweigh the benefits. As such, this suggestion has not been incorporated.</p>

Section	Comments	Working group response
	Allegretti JR, Kassam Z, Osman M, Budree S, Fischer M, Kelly CR. The 5D framework: a clinical primer for fecal microbiota transplantation to treat <em>Clostridium difficile</em> infection. Gastrointest Endosc [Internet]. 2017 Jul 26; Available from: <a href="http://dx.doi.org/10.1016/j.gie.2017.05.036">http://dx.doi.org/10.1016/j.gie.2017.05.036</a>	
8.2.2. Immunosuppression and FMT:	<b><i>FMT should be offered with caution to immunosuppressed patients, in whom FMT appears efficacious without significant additional adverse effects (strong).</i></b>  We agree.	Thank you for this comment.
8.2.3. Other co-morbidities and FMT:	<b>Recommendation:</b> <b><i>i. FMT should be offered to those with recurrent CDI and inflammatory bowel disease (strong).</i></b> <b><i>ii. FMT should be considered for appropriate patients with recurrent CDI regardless of other comorbidities (conditional).</i></b>  We agree.	Thank you for this comment.
8.3.1. General approach to donor selection:	<b><i>Related or unrelated donors should both be considered acceptable. However, where possible, FMT is best sourced from a centralised stool bank, from a healthy unrelated donor (conditional).</i></b>  We agree.	Thank you for this comment.
8.3.2. Age and BMI restrictions for potential donors:	<b><i>People should only be considered as potential FMT donors if they are ≥18 and ≤60 years old, and have a BMI of &lt;30 kg/m<sup>2</sup> (conditional).</i></b>  We agree.	Thank you for this comment.
8.3.3. General approach to the donor screening assessment:	<b><i>A donor-screening history/ questionnaire is mandatory (Table 2) (strong).</i></b> <b><i>1. Receipt of antimicrobials within the past three months.</i></b>	

Section	Comments	Working group response
	<ol style="list-style-type: none"> <li>2. <i>Known prior exposure to HIV and/ or viral hepatitis, and known previous or latent tuberculosis.</i></li> <li>3. <i>Risk factors for blood-borne viruses - including high risk sexual behaviours, use of illicit drugs, any tattoo/ body piercing/ needlestick injury/ blood transfusion/ acupuncture, all within previous six months.</i></li> <li>4. <i>Receipt of a live attenuated virus within the past six months.</i></li> <li>5. <i>Underlying gastrointestinal conditions (e.g. history of IBD, IBS, chronic diarrhoea, chronic constipation, coeliac disease, bowel resection or bariatric surgery).</i></li> <li>6. <i>Family history of any significant gastrointestinal conditions (e.g. family history of IBD, or colorectal cancer).</i></li> <li>7. <i>History of atopy (e.g. asthma, eosinophilic disorders).</i></li> <li>8. <i>Any systemic autoimmune conditions.</i></li> <li>9. <i>Any metabolic conditions, including diabetes and obesity.</i></li> <li>10. <i>Any neurological or psychiatric conditions, or known risk of prion disease.</i></li> <li>11. <i>History of chronic pain syndromes, including chronic fatigue syndrome and fibromyalgia.</i></li> <li>12. <i>History of any malignancy.</i></li> <li>13. <i>Taking particular regular medications, or such medications within the past three months, i.e. antimicrobials, proton pump inhibitors, immunosuppression, chemotherapy</i></li> <li>14. <i>History of receiving growth hormone, insulin from cows, or clotting factor concentrates.</i></li> <li>15. <i>History of receiving an experimental medicine or vaccine within the past six months.</i></li> </ol>	

Section	Comments	Working group response
8.3.4. Laboratory screening of potential donors:	<p><b>Blood and stool screening of donors is mandatory (Tables 2 and 3) (strong).</b></p> <p><b>Table 3: Recommended blood screening for stool donors:</b></p> <p><b>Pathogen screening:</b></p> <ul style="list-style-type: none"><li>• <b>Hepatitis A IgM</b></li><li>• <b>Hepatitis B (HBsAg and HBcAb)</b></li><li>• <b>Hepatitis C antibody</b></li><li>• <b>Hepatitis E IgM</b></li><li>• <b>HIV -1 and -2 antibodies</b></li><li>• <b>HTLV-1 and -2 antibodies</b></li><li>• <b>Treponema pallidum antibodies (TPHA, VDRL)</b></li><li>• <b>Epstein-Barr virus IgM</b></li><li>• <b>Cytomegalovirus IgM</b></li><li>• <b>Strongyloides stercoralis IgG</b></li><li>• <b>Entamoeba histolytica serology</b></li></ul> <p><b>General/ metabolic screening:</b></p> <ul style="list-style-type: none"><li>• <b>Full blood count with differential.</b></li><li>• <b>Creatinine and electrolytes</b></li><li>• <b>Liver enzymes (including albumin, bilirubin, aminotransferases, gamma-glutamyltransferase and alkaline phosphatase).</b></li><li>• <b>C-reactive protein</b></li></ul> <p><b>Table 4: Recommended stool screening for stool donors:</b></p> <ul style="list-style-type: none"><li>• <b>Clostridium difficile PCR</b></li><li>• <b>Campylobacter, Salmonella, and Shigella by standard stool culture and/ or PCR</b></li><li>• <b>Escherichia coli 0157 H7 by culture and/or PCR</b></li></ul>	<p>We agree with the comment regarding matching donors and immunosuppressed recipients for EBV and CMV status, and have updated <b>Section 8.2.2</b> and <b>Section 8.3.4</b> accordingly.</p> <p>The working group did not think that screening for adenovirus was justified.</p> <p>Whilst vancomycin-resistant <i>Enterococci</i> (VRE) carriage is relatively common in the community (probably related to food consumption) (Endtz <i>et al</i>, 1997), the form of VRE in the community is genetically distinct from that found nosocomially, with much lower pathogenicity in community forms (Willems <i>et al</i>, 2005). As such, the working group strongly opined that routine screening was not justified. However, it was acknowledged that the potential infection risk from VRE (and MRSA) would vary regionally depending on local prevalence and pathogenicity, and as such a local risk assessment has been recommended to decide whether screening for these organisms should be considered.</p>

Section	Comments	Working group response
	<ul style="list-style-type: none"> <li>• <b><i>Multi-drug resistant bacteria, specifically carbapenemase-producing Enterobacteriaceae.</i></b></li> <li>• <b><i>Stool ova, cysts and parasite analysis, including for Microsporidia.</i></b></li> <li>• <b><i>Faecal antigen for Cryptosporidium and Giardia.</i></b></li> <li>• <b><i>Acid fast stain for Cyclospora and Isospora.</i></b></li> <li>• <b><i>Helicobacter pylori faecal antigen.</i></b></li> <li>• <b><i>Norovirus and Rotavirus PCR.</i></b></li> </ul> <p>We recommend:</p> <p><b>CMV and EBV:</b> Given the high rates of carriage for both EBV and CMV in a healthy, adult population, excluding EBV or CMV positive donors would make it prohibitively difficult to identify suitable donors to provide access to care (Bate et al). Moreover, excluding EBV or CMV positive candidates is not expected to provide a significant benefit to the majority of the patients that would be served by a centralized stool bank, who are not severely immunocompromised.</p> <p>Given the need to ensure a reliable supply of material for the vast majority of rCDI patients while protecting severely immunocompromised patients, until now OpenBiome has chosen not to test for EBV and CMV. Instead, we treat material as presumptively CMV and EBV positive and discourage use in severely immunocompromised patients who are seronegative for CMV or EBV.</p> <p>We are sensitive to the fact that this leaves clinicians with an additional challenge for managing these already difficult cases (severely immunocompromised rCDI patients). Should FMT be indicated then we would suggest that <b>in the immunocompromised</b></p>	

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	<p>patient at risk of CMV or EBV infection either: 1) CMV and EBV testing of the recipient to confirm positive serology, in which case FMT may be considered after extensive discussion of the risks, benefits, and alternatives in the informed consent process; or 2) the use of a directed donor with matching serology.</p> <p>Bate SL, Dollard SC, Cannon MJ. Cytomegalovirus seroprevalence in the United States: the national health and nutrition examination surveys, 1988-2004. Clin Infect Dis. 2010;50:1439–1447.</p> <p><b>Adenovirus:</b> We recommend including adenovirus on stool in addition to norovirus and rotavirus.</p> <p><b>Vancomycin resistant enterococcus (VRE):</b> VRE should be specifically mentioned in “Multi-drug resistant bacteria”. VRE is a leading cause for donor exclusion despite prospective donors having no known risk factors for colonization.</p>	
8.3.5. Final donor checks prior to donation:	<p><b>Further final screening should take place prior to collection of a stool sample for processing into FMT (strong).</b></p> <p>We agree.</p>	Thank you for this comment. In light of this and other comments, the recommendation on repeat screening has been strengthened.
8.4.1. General principles of FMT preparation:	<p><b>Recommendation:</b></p> <p>i. Donor stool collection should follow a standard protocol (strong).</p> <p>ii. Donor stool should be processed within 6 hours of defecation (conditional).</p> <p>iii. Both aerobically and anaerobically prepared FMT treatments should be considered suitable when preparing FMT for the treatment of recurrent CDI (strong).</p>	Thank you for this comment.



Section	Comments	Working group response
	<p>iv. <i>Sterile 0.9% saline should be considered as an appropriate diluent for FMT production, and cryoprotectant such as glycerol should be added for frozen FMT (strong).</i></p> <p>v. <i>Consider <math>\geq 50\text{g}</math> of stool for use in FMT preparation (conditional).</i></p> <p><b>Good practice points:</b></p> <p>i. <i>Stool should be mixed 1:5 with diluent to make the initial faecal emulsion (conditional).</i></p> <p>ii. <i>Homogenisation and filtration of FMT should be undertaken in a closed disposable system (conditional).</i></p> <p>We agree.</p>	
<b>8.4.2. Fresh vs frozen FMT:</b>	<p><i>The use of banked frozen FMT material should be considered preferable to fresh preparations for CDI (strong).</i></p> <p>We agree.</p>	Thank you for this comment.
<b>8.4.3. Use of frozen FMT:</b>	<p><b>Recommendation:</b></p> <p><i>FMT material stored frozen at <math>-80^{\circ}\text{C}</math> should be regarded as having a maximum shelf life of six months from preparation (strong).</i></p> <p><b>Good practice point:</b></p> <p><i>Consider thawing frozen FMT should at ambient temperature and using within six hours of thawing (conditional).</i></p> <p>We agree.</p>	Thank you for this comment.
<b>8.5.1. Use of specific medications in the period around FMT administration:</b>	<p><b>Recommendation:</b></p> <p>i. <i>Bowel lavage should be administered prior to FMT via the lower GI route, and bowel lavage should be considered prior to FMT via the upper GI route; polyethylene glycol preparation is preferred (conditional).</i></p>	Thank you for this comment.

Section	Comments	Working group response
8.5.1.1. General principles of FMT administration:	<p><i>ii. For upper GI FMT administration, a proton pump inhibitor should be considered, e.g. the evening before and morning of delivery (conditional).</i></p> <p><i>iii. Loperamide (or other anti-motility drugs) should be considered following lower GI FMT delivery (conditional).</i></p> <p><b>Good practice point:</b></p> <p><i>i. Prokinetics (such as metoclopramide) should be considered prior to FMT via the upper GI route (conditional).</i></p> <p><i>ii. Best practice for prevention of further transmission of CDI should be applied throughout when administering FMT to patients with CDI (nursing with enteric precautions, sporicidal treatment of endoscope, etc).</i></p> <p>We agree.</p>	
8.5.1.2. Additional antibiotics pre-FMT:	<p><i>Consider further antimicrobial treatment for CDI for at least 72 hours prior to FMT (conditional).</i></p> <p>We agree.</p>	Thank you for this comment.
8.5.1.3. Washout period between antibiotic use and FMT:	<p><i>To minimise any deleterious effect of antimicrobials on the FMT material, there should be a minimum washout period of 24 hours between the last dose of antibiotic and treatment with FMT (strong).</i></p> <p>We agree.</p>	Thank you for this comment.
8.5.2.2. Upper gastrointestinal tract administration of FMT:	<p><b>Recommendation:</b></p> <p><i>i. Upper GI administration of FMT as treatment for recurrent or refractory CDI should be used where clinically appropriate (strong).</i></p> <p><i>ii. Where upper GI administration is considered most appropriate, FMT administration should be via nasogastric,</i></p>	Thank you for this comment. In light of further discussion by the working group, the maximum volume of FMT recommended by upper GI administration is now 100ml.

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	<p><i>nasoduodenal, or nasojejunal tube, or alternatively via upper GI endoscopy. Administration via a permanent feeding tube is also appropriate (strong).</i></p> <p><b>Good practice point:</b>  <i>It is recommended that no more than 50ml of FMT is administered to the upper GI tract (conditional).</i></p> <p>We agree.</p>	
<p><b>8.5.2.3. Lower gastrointestinal tract administration of FMT:</b></p>	<p><b>Recommendation:</b></p> <ul style="list-style-type: none"> <li><i>i. Colonoscopic administration of FMT as treatment for recurrent or refractory CDI should be used where appropriate (strong).</i></li> <li><i>ii. Where colonoscopic administration is employed, consider preferential delivery to the caecum or terminal ileum, as this appears to give the highest efficacy rate (conditional).</i></li> <li><i>iii. FMT via enema should be used as a lower GI option when colonoscopic delivery is not possible (strong).</i></li> </ul> <p>We recommend rewording point <i>iii</i>. Although there is limited data, flexible sigmoidoscopy may be the preferred route of delivery where colonoscopic delivery is not possible. Several experts have advised less invasive modalities such sigmoidoscopy in high risk patients (Brandt 2013; Kelly 2014). This may provide a more effective method for delivering material as proximally as possible and improving retention. We therefore recommend re-wording point <i>iii</i> to:</p> <p><b><i>FMT via enema should be used as a lower GI option when colonoscopic or flexible sigmoidoscopy delivery is not possible (strong).</i></b></p>	<p>We agree with this suggestion, and have updated the guideline accordingly.</p>

Section	Comments	Working group response
	<p>Brandt LJ, Aroniadis OC. An overview of fecal microbiota transplantation: Techniques, indications, and outcomes. <i>Gastrointest Endosc.</i> 2013 Aug;78(2):240-9.</p> <p>Kelly CR, Ihunnah C, Fischer M, Khoruts A, Surawicz C, Afzali A, et al. Fecal microbiota transplant for treatment of clostridium difficile infection in immunocompromised patients. <i>Am J Gastroenterol.</i> 2014 Jul;109(7):1065-71.</p>	
8.5.2.4. Capsulised FMT:	<p><b><i>Capsulised FMT holds promise as a treatment option for recurrent CDI, but further evidence regarding its safety and efficacy is awaited, and it should not be considered for use at present (conditional).</i></b></p> <p>There is a growing body of evidence on encapsulated FMT and the delivery modality presents a potential option in circumstances where it may be inappropriate, contraindicated, or contrary to patient preferences to deliver material via traditional routes of administration for CDI.</p> <p>In terms of patient perceptions, Zipursky and colleagues report that more aesthetically appealing FMT formulations, such as capsules, would both eliminate potential barriers to treatment and reduce the necessity for healthcare resources and procedure time for clinicians. Capsules appear well tolerated. For example, the mean time of 30 capsule administration is approximately 20 minutes (range 10-30 minute) (Allegretti, unpublished data).</p> <p>Although the optimal dose is still under investigation (as with other FMT delivery modalities), there have been several studies that have shown equivalent efficacy rates. Youngster and colleagues reported their experience with a capsule formulation that averaged 1.6 grams</p>	<p>We largely agree with this comment. Whilst the Kao <i>et al</i>, 2017 study was not published at the time of initial searches, it has been identified by updated searches and has now been reviewed by the working group. The guideline has been updated accordingly.</p>

Section	Comments	Working group response
	<p>of stool per capsule in which they dosed 15 capsules on 2 consecutive days. They reported a 70% cure rate after an initial dose in a cohort of 140 patients. Those that failed to achieve cure were re-treated, bringing the cumulative cure rate up to 90%.</p> <p>Similarly, Hirsch and colleagues demonstrated a clinical cure rate of 68% in the 19 participants, using capsules containing purified, concentrated, and cryopreserved fecal bacteria and this increased to 89% with retreatment.</p> <p>Allegretti and colleagues conducted the first dose-finding study for FMT capsules (0.75 grams of stool per capsule with upper GI release) assessing 30 capsules once (low dose) versus 30 capsules on 2 consecutive days (high dose). Efficacy rates between the groups were similar on initial dose (70%) and there were no adverse events reported.</p> <p>Lastly the largest randomized control trial to date of FMT used encapsulated FMT with good safety and efficacy outcomes equivalent to colonoscopy FMT. In Kao et al's non-inferiority randomized clinical trial (cited in the guidelines) that included 116 adults with rCDI, the proportion without recurrence over 12 weeks was 96.2% after a single treatment in a group treated with oral capsules and in a group treated via colonoscopy. Given this 1+ level of evidence, in addition to multiple smaller studies of encapsulated FMT, we feel that there is a good body of evidence to support the short-term safety of encapsulated FMT. We agree that further evidence is needed on optimal dosing and formulation, however this applies to all delivery modalities.</p>	

Section	Comments	Working group response
	<p>We agree that capsule availability is very limited in the UK at present however this shouldn't preclude guidelines recommending this as a potential FMT delivery option.</p> <p>We therefore recommend rewording the 8.5.2.4 to:</p> <p><b><i>Capsulised FMT holds promise as a treatment option for recurrent CDI and should be offered to patients as a potential treatment modality. Capsule preparations should follow a standard protocol. Further evidence regarding its optimal dosing and formulation is needed (conditional).</i></b></p> <p>Allegretti J*, Fischer M*, Papa E, Elliot R, Klank M, Mendolia G, et al. Fecal microbiota transplantation delivered via oral capsules achieves microbial engraftment similar to traditional delivery modalities: Safety, efficacy and engraftment results from a multi-center cluster randomized dose-finding study. Digestive Disease Week 2016.</p> <p>Hirsch BE, Saraiya N, Poeth K, Schwartz RM, Epstein ME, Honig G. Effectiveness of fecal- derived microbiota transfer using orally administered capsules for recurrent clostridium difficile infection. BMC Infect Dis. 2015 Apr 17;15:191,015-0930-z.</p> <p>Youngster I, Russell GH, Pindar C, Ziv-Baran T, Sauk J, Hohmann EL. Oral, capsulized, frozen fecal microbiota transplantation for relapsing clostridium difficile infection. JAMA. 2014 Nov 5;312(17):1772-8.</p> <p>Zipursky JS, Sidorsky TI, Freedman CA, Sidorsky MN, Kirkland KB. Patient attitudes toward the use of fecal microbiota transplantation</p>	

Section	Comments	Working group response
	in the treatment of recurrent clostridium difficile infection. Clin Infect Dis. 2012 Dec;55(12):1652-8.	
<b>8.6. What is the clinical effectiveness of faecal microbiota transplant in treating conditions other than Clostridium difficile infection?</b>	<b><i>FMT is not currently recommended as treatment for inflammatory bowel disease. There is insufficient evidence to recommend FMT for any other gastrointestinal or non-gastrointestinal disease (strong).</i></b> We agree.	Thank you for this comment.
<b>8.7. Basic requirements for implementing a FMT service</b>	<b><i>The development of FMT centres should be encouraged (strong).</i></b> We agree.	Thank you for this comment.
<b>8.7.5. FMT manufacturing:</b>	<b><i>Ensure traceability of supply (strong).</i></b> We agree.	Thank you for this comment.
<b>FMT in patients with IBD</b>	We recommend emphasizing the importance of counselling patients with IBD on the risk of flare or worsening IBD activity post-FMT.	We agree with this comment, and have updated <b>Section 8.2.3.</b> accordingly.
<b>FMT in paediatric populations</b>	A recommendation on paediatric FMT should be include. The evidence base is limited but safety and efficacy appears comparable to adult FMT. Patients and caregivers should be counselled on the unknown long-term risks of FMT.  Recommendation: <b><i>i. FMT should be offered to paediatric patients with recurrent CDI. ii. Paediatric patients and caregivers should be counselled on the unknown short and long-term risks of FMT.</i></b>	FMT in the paediatric setting is outside of the remit of this working group. We have updated <b>Section 5.4</b> to clarify this.

**Closing date:** Please forward this electronically by 5pm on January 2018 at the very latest to [consultations@his.org.uk](mailto:consultations@his.org.uk)



Supplementary Material 3 for *Gut*

**The use of faecal microbiota transplant as treatment for recurrent or refractory *Clostridium difficile* infection and other potential indications: joint British Society of Gastroenterology (BSG) and Healthcare Infection Society (HIS) guidelines.**

**Supplementary Material 3: Basic requirements for implementing a FMT service:**

**1. Basic requirements for implementing a FMT service:**

**1.1. General considerations:**

Although it is possible to prepare and administer FMT on an individual patient basis in a single hospital, the regulatory requirements are more readily fulfilled by a specialist centre approach for the production of a safe FMT product. This particularly applies to record keeping and staff expertise in quality control and production. Recent European consensus advice suggests that FMT should be administered in a referral centre<sup>1</sup>, however an alternative approach which limits the need for patient transfer is to undertake controlled production in a large centre and transport treatment to the patient, a supply model which has been well established in the USA (OpenBiome)<sup>2</sup> and has also been successfully replicated in the UK in a large centre in Birmingham, which has supplied FMT to nine NHS Trusts across three regions<sup>3</sup>. This service design only requires that a responsible clinician is capable of administering the FMT safely at the satellite clinical site. It also eliminates the need for patient transfer between clinical sites, which in the case of severe CDI may not be practical.

The working group encouraged the use of frozen FMT material supplied from a carefully controlled production site. This allows donor screening more closely to meet regulatory requirements, ensuring that the window period between donor testing and FMT production is maintained to a minimum. The costs of donor screening are substantially reduced using this supply model, as a single donor can provide multiple FMT donations under a single screening period.

The working group also noted that given the novelty of FMT, awareness of this as a potential treatment option for recurrent or refractory CDI may be low amongst certain groups of clinicians. For instance, clinicians working in primary care, or those whose practice is not located near to an FMT centre, are likely to have less knowledge about the potential suitability of FMT for patients with CDI, or be unaware of referral pathways. As such, there is a responsibility for FMT centres to raise awareness and educate as wide a range of clinicians as possible about the potential role for FMT.

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Furthermore, microbiology staff processing stool samples for *C difficile* assays from the community should proactively liaise with primary care teams where recurrent positive tests are received from a single patient to raise awareness and suggest the option of FMT.

Similarly, given the expectation that FMT and/ or other 'microbiome therapeutics' are likely to play an increasing role within medicine over future years, there is also an expectation for FMT centres to not only educate about the potential role for FMT, but also to train relevant healthcare professionals in the practicalities of delivering an FMT service, to enable longer-term ongoing provision of services. This is likely to be most of relevance to specialty trainee and consultant physicians specialising in gastroenterology, infectious diseases and/ or medical microbiology, but potentially to other healthcare professionals too, including infection prevention and control nurses, infectious diseases pharmacists, etc.

**Recommendations:**

- i. The development of FMT centres should be encouraged (GRADE of evidence: very low; strength of recommendation: strong).*
- ii. We suggest that FMT centres should work to raise awareness about FMT as a treatment option amongst clinicians caring for patients with CDI, and provide training to relevant healthcare professionals on the practicalities of delivering an FMT service (GRADE of evidence: very low; strength of recommendation: weak).*

**1.2. Legal aspects and clinical governance:**

In the United Kingdom, FMT is now considered a medicinal product based on the definitions of purpose and efficacy, in The Medicines Directive 2001/83 and The Human Medicines Regulations<sup>4</sup>. As the competent authority for medicines regulation, the Medicines and Healthcare products Regulatory Agency (MHRA) has indicated that the approach to regulation will be proportionate, depending on factors such as supply being within or outside a legal entity and FMT production scale. Specifically:

- When FMT is supplied on prescription on a named patient basis, then supply under a pharmacy exemption may be used subject to ensuring proper governance and traceability<sup>4</sup>.
- If production scale reaches an 'industrial' level, defined 'by virtue of the batch sizes, the extent of processing and/ or whether potential use includes supply between legal entities'<sup>4</sup>, the route to regulation is via adherence to HMR and formal Manufacturer's 'Specials' (MS) license.

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- If a supply is to a clinical trial, then an MIA (IMP) manufacturing license is required (further information on license applications<sup>5</sup> and specials<sup>6</sup> is available online).

Centres establishing an FMT service should undertake steps to ensure practice meets the required compliance levels and seek guidance from the MHRA. If pharmacy exemption is applied, there should be justifiable processes in place to ensure traceability, health and safety, governance and to prevent cross-contamination. FMT is regulated as a medicine, rather than a tissue, but no products have been licensed following an assessment against the criteria of safety, quality and efficacy, for there is a possible risk that donor screening protocols will not be sufficiently considered, a step which is critical to the quality of the product and therefore safety of the patient<sup>7</sup>. To mitigate this, it is advisable that donor screening protocols are under regularly review and risk assessment, and to ensure that consideration is also given to the Guide to Quality and Safety Assurance for Human Tissues and Cells for Patient Treatment, particularly Annex B related to donor testing<sup>8</sup>. When formal licencing is sought, this is overseen by a Production Manager and Quality Control Manager if under an MS, or by a Qualified Person if under an MIA (IMP). Both should follow the Good Manufacturing Practice (GMP) guidelines, found within The Orange Guide Rules and Guidance for Pharmaceutical Manufacturers and Distributors 2017<sup>9</sup>, or at: [https://ec.europa.eu/health/documents/eudralex/vol-4\\_en](https://ec.europa.eu/health/documents/eudralex/vol-4_en).

The working group noted that outside the UK, the legal and regulatory framework relating to FMT was highly variable between different countries. They agreed that FMT should only be administered after appropriate approval from the competent body of each country.

**Recommendation:**

***In the UK, FMT must be manufactured in accordance with MHRA guidance for human medicines regulation. When FMT is supplied on a named patient basis, within a single organisation, a pharmacy exemption may be used, subject to ensuring proper governance and traceability. All centres that are processing and distributing FMT should seek guidance from the MHRA and where necessary obtain appropriate licenses prior to establishing an FMT service. This is a legal requirement. In countries other than the UK, FMT should only be manufactured following appropriate approval from the national authority of that country (GRADE of evidence: very low; strength of recommendation: strong).***

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**1.3. Multidisciplinary teams:**

To promote safe and high quality FMT supply, it is strongly recommended that providers adopt a multidisciplinary team approach. The choice of the team required is subject to the scale of production, but should involve as a minimum a clinical gastroenterologist, microbiologist/infectious diseases clinician, state-registered experienced healthcare scientist and pharmacist. Governance and quality expertise will be required, which may be provided by consultation. If FMT production is to be under a 'specials' licence, the team should be expanded to include a Qualified Person, Quality Manager and Production Manager, all with GMP training.

**Recommendation:**

***We recommend that a multidisciplinary team should be formed to deliver FMT services (GRADE of evidence: very low; strength of recommendation: strong).***

**1.4. Infrastructure:**

Dedicated laboratory facilities for FMT production are recommended to ensure that the process adheres to Health and Safety requirements, to reduce the risk of cross-contamination, and to facilitate standardisation of the production process. In some studies, FMT has been prepared in a clinical environment<sup>10</sup>; however, this may not be advisable because of the risks of cross-contamination. The manipulation of human stool should be conducted in a Containment Level 2 laboratory according to current Health and Safety guidance (Health and Safety at Work Act 1974, COSHH Control of Substances Hazardous to Health Regulations, 2002), and at least within a microbiological safety cabinet which provides user protection (Class I) or, ideally, user and product protection (Class II). To meet the requirements of GMP, this facility should be sole use or be risk assessed for multipurpose use with adequate separation of different activities. The working group recommend that the facility complies with the new GMP production facility classification of 'clean not sterile'. The use of personal protective equipment - such as laboratory coat, gloves and face mask - is also recommended to prevent production contamination. It is essential to risk assess the process and develop control measures to reduce microbial ingress into the facility and monitor the microbiological cleanliness of the production suite. FMT preparation under a 'specials' licence should ensure that the production process is integrated into a Quality Management System, to safeguard production and maintain the minimum criteria for audit, monitoring, standard operating procedures, document control, training, facilities, equipment and storage. With regard to storage, it is essential that the freezer system has real-time temperature monitoring which provides notification outside pre-set limits.

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**Recommendation:**

***We recommend utilisation of suitable laboratory facilities and infrastructure for FMT production (GRADE of evidence: very low; strength of recommendation: strong).***

**1.5. FMT manufacturing:**

It is strongly recommended to employ a batch numbering system to track FMT preparations from production to use. It should be possible from records to identify an individual FMT aliquot, trace it to a specific donation, and identify all other FMT aliquots prepared from the same donation. It must also be clear which FMT aliquots patients have received, which should be verifiable from the donor to the patient and vice-versa. It is therefore strongly recommended that a treatment directory be maintained documenting all production and use of FMT, and that an unambiguous record is created in the patients' clinical notes to identify the specific FMT batch number. Further to this, it is also recommended that treatment directories also record clinical outcome, such as that developed in the USA<sup>11</sup> and Germany<sup>12</sup> to standardise and improve future clinical practice.

**Recommendation:**

***We recommend ensuring the traceability of supply (GRADE of evidence: very low; strength of recommendation: strong).***

**1.6. FMT production quality control:**

Safety and clinical governance is a central responsibility for FMT centres, particularly in light of the absence of phase III licensing trials for FMT, which would normally be required for a novel medicinal product. Reporting and investigating adverse events and reactions contributes to knowledge of the FMT safety profile, while also identifying previously unknown safety issues. Governance structures and processes must be in place to monitor, notify and investigate all FMT-related adverse events or reactions locally, and FMT users are encouraged to use the MHRA Yellow Card Scheme for formal notification. FMT supply should be suspended if serious adverse events or reactions occur which are directly attributable to FMT, and there should be a clear documented pathway to achieve this. To facilitate a 'look-back exercise' if required, it is advisable to store documentation and reference samples, both product-based and donor/ patient-based. Specifically, retention of production documentation should be for at least five years after the use of the batch; retention of reference FMT

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samples (and stool samples from donors and recipients) should be for at least one year after the last use. Retention of excipient samples should be for at least one year after expiry of the excipient.

**Recommendation:**

***We recommend monitoring, notification and investigation of all adverse events and reactions related to FMT (GRADE of evidence: very low; strength of recommendation: strong).***

**1.7. Donor screening governance:**

The testing requirements for donor screening have been discussed previously; however, it is worth noting here the pertinent clinical governance issues which should be addressed. Donor anonymity should be maintained at all times. The laboratory undertaking testing of donor samples should be competent for such activity, demonstrable by accreditation with the United Kingdom Accreditation Service (UKAS). The results of donor testing should remain confidential. There should be appropriate standard operating procedures to ensure that the outcome of donor screening is built into a robust FMT batch release process. To ensure unbiased autonomy during donor screening, it is suggested that a clinician independent to the FMT production team is responsible for ratifying FMT donors prior to donation. Finally, the duration of donor follow-up should be considered and extend beyond the period of active donation to capture acute and chronic health changes.

**Recommendation:**

***We recommend ensuring the clinical governance of donor screening (GRADE of evidence: very low; strength of recommendation: strong).***

**2. References:**

1. Cammarota G, Ianiro G, Tilg H, et al. European consensus conference on faecal microbiota transplantation in clinical practice. *Gut*. 2017;66(4):569-580. doi:10.1136/gutjnl-2016-313017.
2. Home — OpenBiome. <http://www.openbiome.org/home/>. Accessed October 19, 2017.
3. Quraishi MN, Segal J, Mullish B, et al. National survey of practice of faecal microbiota

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- transplantation for *Clostridium difficile* infection in the UK. *J Hosp Infect.* 2016. doi:10.1016/j.jhin.2016.10.023.
4. Faecal Microbiota Transplantation (FMT) MHRA's position. [http://www.bsg.org.uk/images/stories/docs/clinical/guidance/fmt\\_mhra\\_position\\_june2015.pdf](http://www.bsg.org.uk/images/stories/docs/clinical/guidance/fmt_mhra_position_june2015.pdf). Accessed October 3, 2017.
5. Medicines and Healthcare products Regulatory Agency. Apply for manufacturer or wholesaler of medicines licences - GOV.UK. <https://www.gov.uk/guidance/apply-for-manufacturer-or-wholesaler-of-medicines-licences#apply-for-a-manufacturerimporter-licence>. Accessed February 15, 2018.
6. Medicines and Healthcare products Regulatory Agency. Supply unlicensed medicinal products (specials) - GOV.UK. <https://www.gov.uk/government/publications/supply-unlicensed-medicinal-products-specials>. Accessed February 15, 2018.
7. Edelstein CA, Kassam Z, Daw J, Smith MB, Kelly CR. The regulation of fecal microbiota for transplantation: An international perspective for policy and public health. *Clin Res Regul Aff.* 2015;32(3):99-107. doi:10.3109/10601333.2015.1046602.
8. Human Tissue Authority Guide to Quality and Safety Assurance for Human Tissues and Cells for Patient Treatment. 2010. [https://www.hta.gov.uk/sites/default/files/Annex\\_-\\_Guide\\_to\\_Quality\\_and\\_Safety\\_Assurance\\_for\\_Tissues\\_and\\_Cells\\_for\\_Patient\\_Treatment.pdf](https://www.hta.gov.uk/sites/default/files/Annex_-_Guide_to_Quality_and_Safety_Assurance_for_Tissues_and_Cells_for_Patient_Treatment.pdf). Accessed October 9, 2017.
9. Great Britain. Medicines and Healthcare products Regulatory Agency. *Rules and Guidance for Pharmaceutical Manufacturers and Distributors 2017.*
10. Allegretti JR, Korzenik JR, Hamilton MJ. Fecal microbiota transplantation via colonoscopy for recurrent *C. difficile* Infection. *J Vis Exp.* 2014;(94). doi:10.3791/52154.
11. Kelly CR, Kim AM, Laine L, Wu GD. The AGA's Fecal Microbiota Transplantation National Registry: An Important Step Toward Understanding Risks and Benefits of Microbiota Therapeutics. *Gastroenterology.* 2017;152(4):681-684. doi:10.1053/j.gastro.2017.01.028.
12. Vehreschild M. MicroTrans - A Multicenter Registry of Fecal Microbiota Transplantation (MicroTrans). doi:10.1016/j.cmi.2015.06.027.